



## Whole-Genome Sequence of Aneurinibacillus sp. Ricciae\_BoGa-3, Isolated from Riccia fluitans

Marvin Hildebrandt, <sup>a</sup> Isabell E. Bleile, <sup>a</sup> <sup>(b)</sup> Felix Althoff, <sup>c</sup> <sup>(b)</sup> Sabine Zachgo, <sup>c</sup> <sup>(b)</sup> Andrea Bräutigam, <sup>a,b</sup> <sup>(b)</sup> Bart Verwaaijen<sup>a,b,d</sup>

<sup>a</sup>Bielefeld University, Computational Biology, Faculty of Biology, Bielefeld, Germany

<sup>b</sup>CeBiTec, Bielefeld University, Bielefeld, Germany

<sup>c</sup>Osnabrück University, Department of Botany, Osnabrück, Germany

<sup>d</sup>Department of Genetics, Martin-Luther-University Halle-Wittenberg, Halle, Germany

Marvin Hildebrandt and Isabell E. Bleile contributed equally. Marvin Hildebrandt was placed on the first position as he was the main author of the manuscript text.

**ABSTRACT** Here, we present the Nanopore-only genome sequence of *Aneurinibacillus* sp. *Ricciae\_BoGa-3*. It was isolated from *Riccia fluitans* ecotype BoGa-3 and its source was Botanical Garden Osnabrück (Germany). The complete circular genome is 4,981,254 bp with a GC content of 44.8%.

Members of the Gram-positive, endospore-forming, and rod-shaped bacterial genus Aneurinibacillus (1) occupy diverse habitats like the plant rhizosphere (2), geothermal soil (3), or marine environments (4). Several Aneurinibacillus species are known to produce useful metabolites, such as antibiotics (5) and biosurfactants (4), or exhibit plant-growth-promoting traits, such as phosphate solubilization and growth inhibition of plant pathogens (2). Of this genus, so far, only seven type strains have been published according to the Type Strain Genome Server (TYGS) (6, 7). Therefore, there may be a great potential within this genus for further discoveries of species that have useful properties for agricultural or biotechnological applications (8).

We isolated *Aneurinibacillus* sp. *Ricciae\_BoGa-3* from a laboratory-grown *Riccia fluitans* (floating crystalwort) (9) line, ecotype BoGa, named after its original source, the Botanical Garden of Osnabrück University (Germany) (10, 11). The medium used was 1/ 2 Gamborg B5 medium with 1% glucose. Plants were grown at room temperature with a 16:8 day:night regime, and after colonies had formed around the plants, DNA was isolated from a single colony, with the NucleoSpin microbial DNA minikit (Macherey-Nagel, Düren, Germany). Sequencing was performed with the kit SQK-LSK109 on a GridION device with a R9.4.1 flow cell (Oxford Nanopore, Oxford, UK). Next, we performed base calling at super accuracy (Guppy v5.0.11), assembly (Canu v2.1.1) (12), and polishing with Racon (v1.4.20) (13) in combination with BWA (v0.7.17) (14) and Medaka (v1.4.3). The final contig was circularized and oriented manually. Default settings were used for all tools.

Raw sequencing generated 1.25 million reads, an  $N_{50}$  of 9.07 kilobases, and 3.22 total gigabases. Assembly showed  $511 \times$  coverage, a GC content of 44.8%, and a length of 4,981,254 bp. Annotation showed 5,938 total genes and 5,472 coding genes. Genome completeness was determined with BUSCO and included 98.3% complete (C), 96.7% single copy (S), 1.6% duplicated (D), 1.1% fragmented (F), and 0.6% missing (M) orthologues genes (15, 16). Annotation was based on NCBI PGAP (v6.4) annotation of CP116887 on 1 January 2023 (17).

For the genome sequence of *Aneurinibacillus* sp. *Ricciae\_BoGa-3* and the seven published type strains, a phylogenetic network was calculated using SplitsTree (18) (Fig. 1) with default settings on an alignment of 16S sequences created with Clustal Omega (19). Complementary to the 16S-based network, TYGS (6, 7) was used to calculate a Editor David A. Baltrus, University of Arizona Copyright © 2023 Hildebrandt et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Bart Verwaaijen, bart.verwaaijen@genetik.uni-halle.de. The authors declare no conflict of interest.

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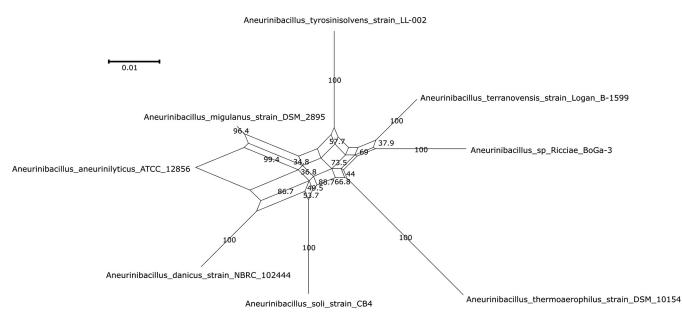


FIG 1 Phylogenetic network of A. ricciae and the seven Aneurinibacillus type strains calculated with Clustal Omega and SplitsTree (18). The alignment was calculated using 1,500-bp-long 16S RNA sequences. Bootstrap values from 1,000 replications are shown as labels on the corresponding edges.

whole-proteome-based tree (not shown). The obtained average branch support for the tree is 98.1%. The network and tree both support the finding that *Aneurinibacillus* sp. *Ricciae\_BoGa-3* is closest to *Aneurinibacillus terranovensis* but with enough phyloge-netic distance to indicate that *Aneurinibacillus* sp. *Ricciae\_BoGa-3* is distinct from previously known species. Neither the addition of not-type-strain species to the network nor a BLAST search against the NCBI nonredundant (nr) database (20, 21) for the 16S sequence provide additional candidates for more closely related species. The digital DNA-DNA hybridization (dDDH) values for *Aneurinibacillus* sp. *Ricciae\_BoGa-3* and *A. terranovensis* for all three distance formulas are significantly below the 70% cutoff (22).

Although *Aneurinibacillus* sp. *Ricciae\_BoGa-3* formed colonies when cocultivated with its host, it did not grow on 1/2 Gamborg medium with 1% glucose. This finding suggests a dependency on one or more plant exudates. According to a mapping of genes on KEGG pathways (23, 24), the bacterium may be auxotrophic for several B vitamins. These findings combined with the existing literature base of the genus indicate that *Aneurinibacillus* sp. *Ricciae\_BoGa-3* is a plant-associated species and possible symbiont. The genome presented here was determined with Nanopore only and despite deep sequencing ( $511 \times$ ) might still contain Nanopore sequencing-related errors.

**Data availability.** This whole-genome shotgun project has been deposited in GenBank under the BioProject no. PRJNA914707, BioSample no. SAMN32348813, and accession no. CP116887. Raw sequence reads can be found in SRA under SRR23191365.

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## REFERENCES

- Subhash Y, Kim H, Lee S-S. 2017. Aneurinibacillus sediminis sp. nov., isolated from lagoon sediments. Int J Syst Evol Microbiol 67:2544–2548. https://doi.org/10.1099/ijsem.0.001963.
- Chauhan A, Balgir PP, Shirkot CK. 2014. Characterization of Aneurinibacillus aneurinilyticus strain CKMV1 as a plant growth promoting rhizobacteria. Int J Agric Environ Biotechol 7:37. https://doi.org/10.5958/j.2230-732X.7.1.006.
- 3. Allan RN, Lebbe L, Heyrman J, De Vos P, Buchanan CJ, Logan NA. 2005.

Brevibacillus levickii sp. nov. and Aneurinibacillus terranovensis sp. nov., two novel thermoacidophiles isolated from geothermal soils of northern Victoria Land, Antarctica. Int J Syst Evol Microbiol 55:1039–1050. https:// doi.org/10.1099/ijs.0.63397-0.

 Balan SS, Kumar CG, Jayalakshmi S. 2017. Aneurinifactin, a new lipopeptide biosurfactant produced by a marine Aneurinibacillus aneurinilyticus SBP-11 isolated from Gulf of Mannar: purification, characterization and its biological evaluation. Microbiol Res 194:1-9. https://doi.org/10.1016/j .micres.2016.10.005.

- Berditsch M, Afonin S, Ulrich AS. 2007. The ability of Aneurinibacillus migulanus (Bacillus brevis) to produce the antibiotic gramicidin S is correlated with phenotype variation. Appl Environ Microbiol 73:6620–6628. https://doi.org/10.1128/AEM.00881-07.
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. Nat Commun 10: 2182. https://doi.org/10.1038/s41467-019-10210-3.
- Meier-Kolthoff JP, Carbasse JS, Peinado-Olarte RL, Göker M. 2022. TYGS and LPSN: a database tandem for fast and reliable genome-based classification and nomenclature of prokaryotes. Nucleic Acids Res 50:D801–D807. https:// doi.org/10.1093/nar/gkab902.
- Kamli MR, Alzahrani NAY, Hajrah NH, Sabir JSM, Malik A. 2021. Genome-driven discovery of enzymes with industrial implications from the genus Aneurinibacillus. Microorganisms 9:499. https://doi.org/10.3390/microorganisms9030499.
- 9. Edwards SR. 1999. English names for British bryophytes. British Bryological Society, London, United Kingdom.
- Althoff F, Zachgo S. 2020. Transformation of Riccia fluitans, an amphibious liverwort dynamically responding to environmental changes. Int J Mol Sci 21:5410. https://doi.org/10.3390/ijms21155410.
- Althoff F, Wegner L, Ehlers K, Buschmann H, Zachgo S. 2022. Developmental plasticity of the amphibious liverwort Riccia fluitans. Front Plant Sci 13:909327. https://doi.org/10.3389/fpls.2022.909327.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722–736. https://doi.org/10.1101/ gr.215087.116.
- Vaser R, Sović I, Nagarajan N, Šikić M. 2017. Fast and accurate de novo genome assembly from long uncorrected reads. Genome Res 27:737–746. https://doi.org/10.1101/gr.214270.116.
- 14. Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv https://doi.org/10.48550/arXiv.1303.3997.
- Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. Bioinformatics 31:3210–3212. https://doi.org/ 10.1093/bioinformatics/btv351.

- Manni M, Berkeley MR, Seppey M, Simão FA, Zdobnov EM. 2021. BUSCO update: novel and streamlined workflows along with broader and deeper phylogenetic coverage for scoring of eukaryotic, prokaryotic, and viral genomes. Mol Biol Evol 38:4647–4654. https://doi.org/10.1093/molbev/ msab199.
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. Nucleic Acids Res 49:D1020–D1028. https://doi.org/10.1093/ nar/gkaa1105.
- Huson DH, Bryant D. 2006. Application of phylogenetic networks in evolutionary studies. Mol Biol Evol 23:254–267. https://doi.org/10.1093/ molbev/msj030.
- Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, Lopez R, McWilliam H, Remmert M, Söding J, Thompson JD, Higgins DG. 2011. Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. Mol Syst Biol 7:539. https://doi.org/10.1038/msb.2011.75.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- Johnson M, Zaretskaya I, Raytselis Y, Merezhuk Y, McGinnis S, Madden TL. 2008. NCBI BLAST: a better Web interface. Nucleic Acids Res 36:W5–W9. https://doi.org/10.1093/nar/gkn201.
- Meier-Kolthoff JP, Auch AF, Klenk H-P, Göker M. 2013. Genome sequencebased species delimitation with confidence intervals and improved distance functions. BMC Bioinformatics 14:60. https://doi.org/10.1186/1471 -2105-14-60.
- 23. Kanehisa M, Goto S. 2000. KEGG: Kyoto Encyclopedia of Genes and Genomes. Nucleic Acids Res 28:27–30. https://doi.org/10.1093/nar/28.1 .27.
- Kanehisa M, Sato Y, Kawashima M. 2022. KEGG mapping tools for uncovering hidden features in biological data. Protein Sci 31:47–53. https://doi .org/10.1002/pro.4172.