

Article

The Finally Rewarding Search for A Cytotoxic Isosteviol Derivative

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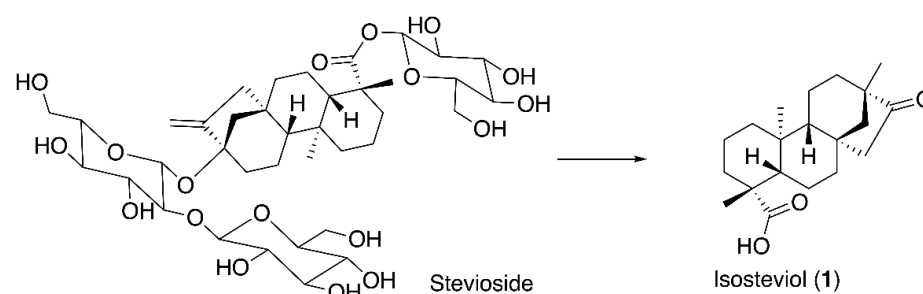
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Abstract: Acid hydrolysis of stevioside resulted in a 63% yield of isosteviol (**1**), which served as a starting material for the preparation of numerous amides. These compounds were tested for cytotoxic activity, employing a panel of human tumor cell lines, and almost all amides were found to be non-cytotoxic. Only the combination of isosteviol, a (homo)-piperazinyl spacer and rhodamine B or rhodamine 101 unit proved to be particularly suitable. These spaced rhodamine conjugates exhibited cytotoxic activity in the sub-micromolar concentration range. In this regard, the homopiperazinyl-spaced derivatives were found to be better than those compounds with piperazinyl spacers, and rhodamine 101 conjugates were more cytotoxic than rhodamine B hybrids.

Keywords: stevioside; isosteviol; rhodamine conjugates; cytotoxicity

1. Introduction

At least since the introduction and approval of stevioside [1–5] as a sweetener and substitute for sugar, increased attention has been paid to its aglycone steviol and its rearrangement product isosteviol (**1**, Scheme 1) [3,4]. Stevioside (Scheme 1) is now present in numerous food and consumer goods in daily life, as it is a non-caloric sugar substitute with a sweetening power of 250–300 times sweeter than sucrose. In addition, numerous therapeutic benefits are attributed to it, including antihyperglycemic, antihypertensive, anti-inflammatory, antitumor, and immunomodulatory properties [6–11]. Alkaline as well as enzymatic hydrolysis of stevioside leads to steviol, while acid hydrolysis leads to isosteviol (**1**) [12–17]. Isosteviol is a diterpene of the *ent*-beyerane = stachane type, i.e., a 13-methyl-17-norkaurane; it is obtained biosynthetically from the cyclization of a pimarane cation intermediate without subsequent rearrangement.



Scheme 1. Hydrolysis (methanolic HCl, Δ , 2 h, 63%) of stevioside to isosteviol (**1**).

Isosteviol has been reported to be a suppressor of human DNA topoisomerase II [18] and of mammalian DNA polymerases, thus explaining its antitumor activity [19,20]. Parent stevioside was reported to have minor cytotoxic effects on several human cancer cell lines,



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thereby inducing apoptosis and cell cycle arrest in the G₂/M phase [21–38]. Since previous work also indicated an effect on mitochondrial permeability transition [35], it was natural to transfer our own previous findings on pentacyclic triterpenes to the diterpene isosteviol. Of particular interest were benzyl and pyridine-amides, (iso)-quinoline amides, and especially (homo)-piperazinyl-spacered rhodamine B and rhodamine 101 conjugates [39–58].

Quinoline (and isoquinoline) derivatives of pentacyclic triterpenes had shown high cytotoxicity with partly excellent tumor cell/non-tumor cell selectivity [56], while several benzylamides were of good cytotoxicity but also excellent selectivity [43,46–48,53,55,58–60]; rhodamine B conjugates of triterpenoic acid held excellent cytotoxic activity; for some derivatives even sub-nanomolar cytotoxicity has been reported [51].

Previous results from our labs revealed a dependance on the type of spacer between the triterpene and the cationic rhodamine moiety. Thereby, piperazinyl and homopiperazinyl spacer proved especially useful; hence, these two spacers were included in this study [45,46,51,55,57].

Many solid tumor cells hold a trans-membrane potential exceeding that of “normal” cells, thus allowing a selective accumulation of lipophilic delocalized cations in the tumor cell mitochondria [61–68]. Our previous studies on these cationic hybrids were limited to pentacyclic triterpenoid hybrids; the first results obtained for diterpenoid abietylamine indicated some potential also for non-pentacyclic triterpenoids [69–72]. It seems that, depending on both the kind of terpene as well as on the chosen cation, different modes of action are involved. While for triterpenoid-safirinium compounds [73], as well as for an aza-BODIPY [74], an accumulation in the endoplasmic reticulum was observed; several rhodamine conjugates interact in the mitochondria [46], and even an almost complete shut-down of the mitochondrial ATP synthesis has been observed for an Asiatic acid-derived hybrid [51].

2. Results

Stevioside (obtained from different local vendors) was hydrolyzed in methanolic hydrochloric acid, and isosteviol (**1**) was obtained in a 63% isolated yield. Due to some ambiguity in the assignment of its ¹H and ¹³C NMR spectra [75–77], a complete analysis was undertaken. However, it quickly became apparent that the usual 1D and 2D NMR experiments (e.g., gHSQC and gHMBC) did not allow a complete and accurate assignment of all signals, since even in these spectra, some signals overlapped very strongly. In addition, data from the literature contradicted each other with respect to the assignment of the quaternary carbons. To solve this problem, further NMR experiments were carried out, with which it was possible to directly separate the connectivity in the carbon framework. Two methods are suitable for this purpose: On the one hand, this is the classical INADEQUATE method [78–81], which requires larger substance amounts but also long measurement times due to the low sensitivity, or, on the other hand, the 1,1-ADEQUATE experiment [82–84]. In contrast to the INADEQUATE experiment, which is based on a correlation of the ¹J coupling between two vicinal ¹³C nuclei, the 1,1-ADEQUATE experiment uses an INEPT transfer between ¹H and ¹³C adjacent nuclei and the subsequent formation of double quantum coherence, thereby circumventing the problem of the low natural abundance of ¹³C nuclei.

So far, no INADEQUATE or 1,1-ADEQUATE spectra have been reported in the literature specifically for isosteviol. Due to the very good solubility of isosteviol in chloroform (520 mg in 0.7 mL) and yet sufficiently low viscosity at the measuring temperature used (27 °C), it was possible to perform both experiments and compare the results. While the INADEQUATE experiment required a measuring time of 80 h, 20 h was sufficient for the ADEQUATE experiment. The results are shown in Figures 1 and 2 (in the ADEQUATE experiment, the chemical shift is shown in ppm instead of the double quantum coherence frequency, for better comparability). It was shown that both methods are very well suited for an exact and doubtless assignment. However, it is also evident that the INADEQUATE spectrum is somewhat easier to interpret, since no overlapping proton signals have to

be taken into account. Both NMR experiments, however, allow a doubtless and reliable assignment of all signals.

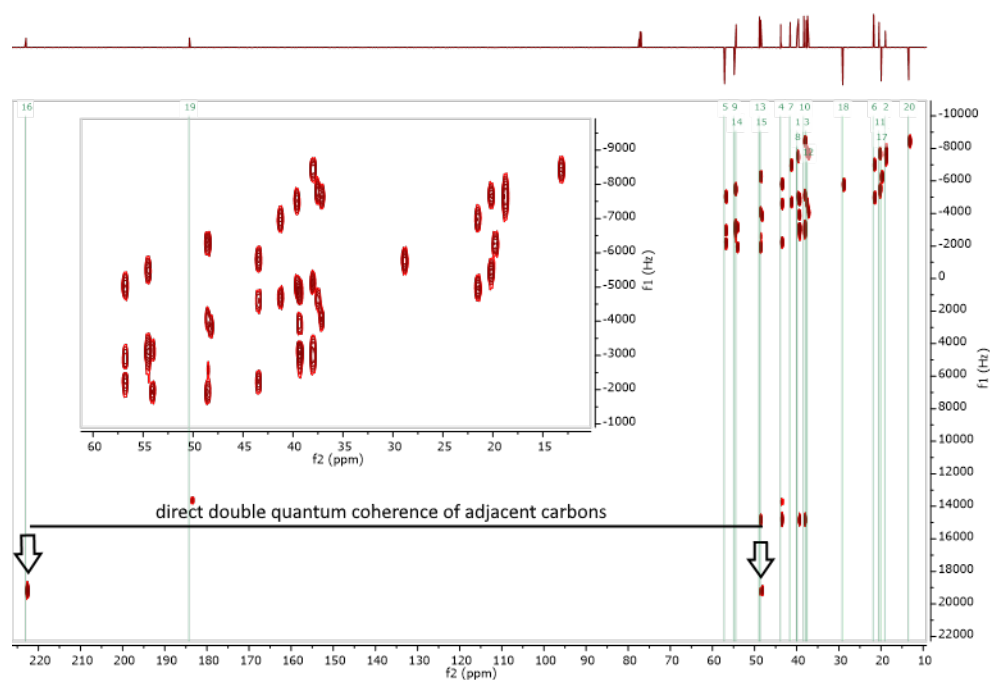


Figure 1. INADEQUATE spectrum of **1** in CDCl_3 ; total acquisition time: 80 h.

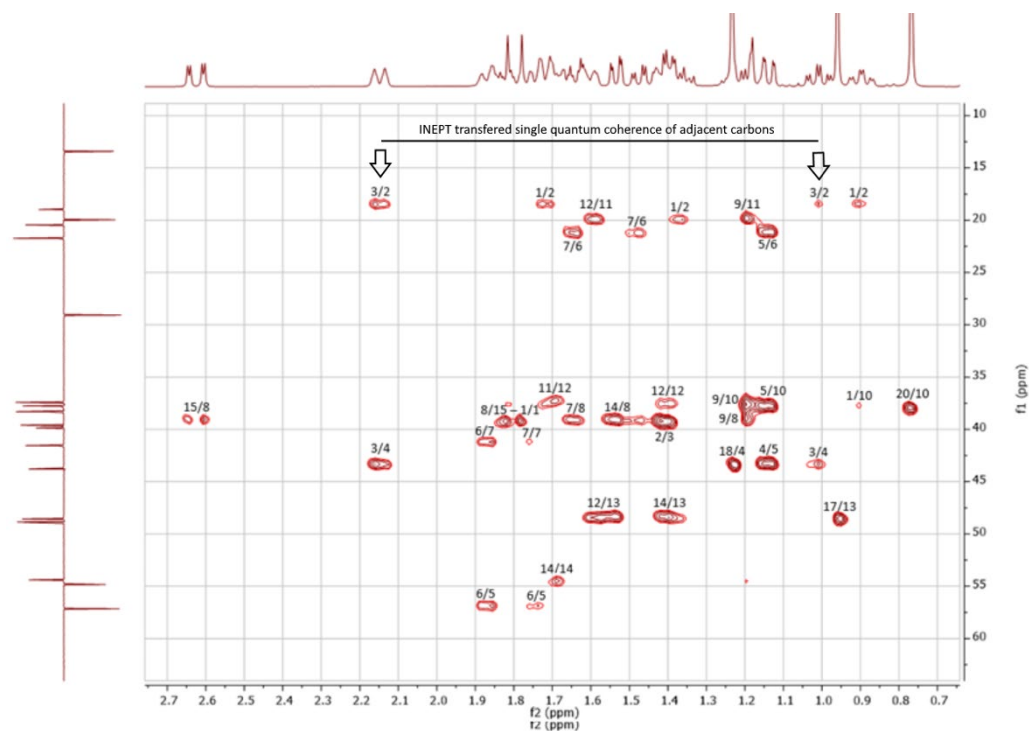
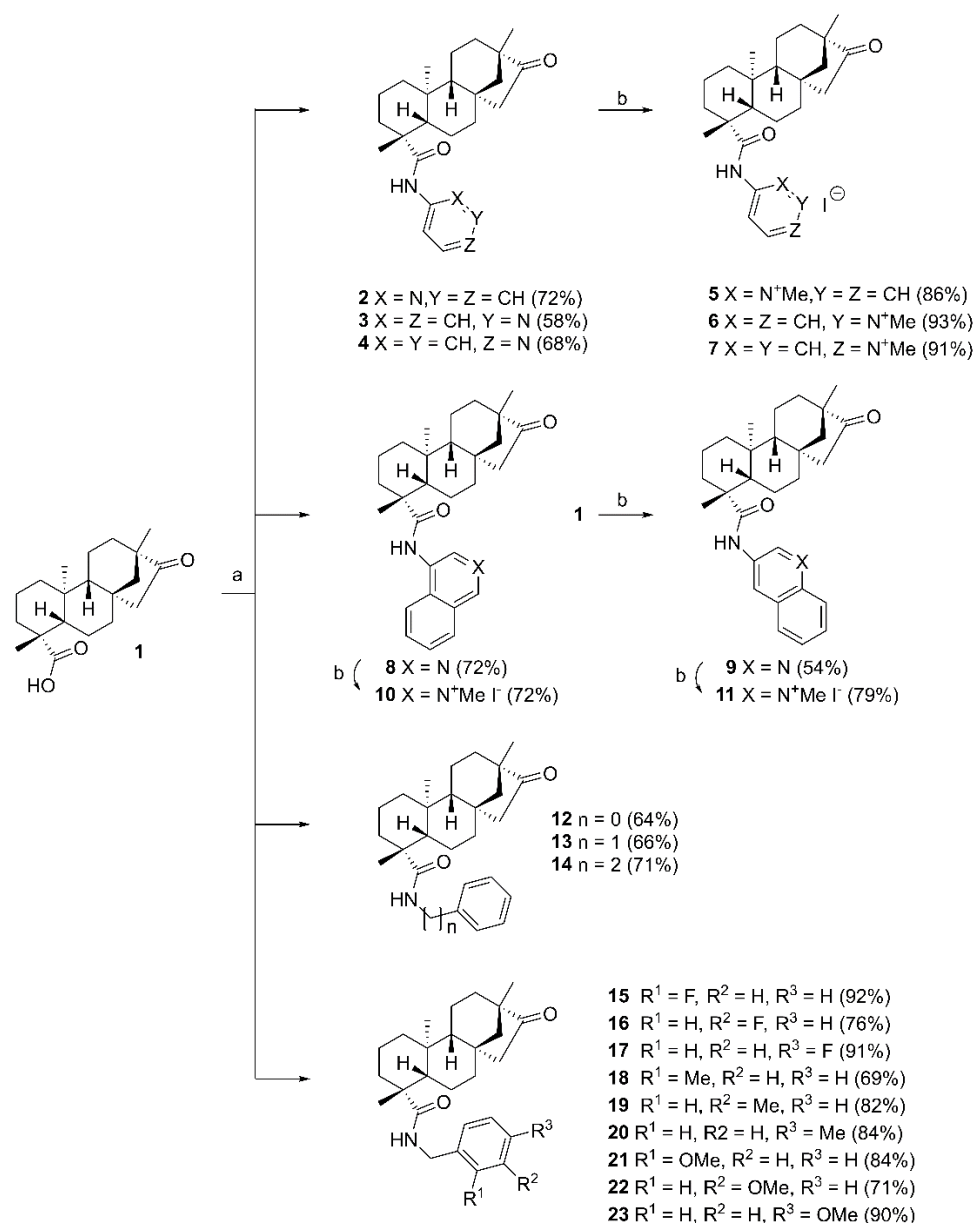


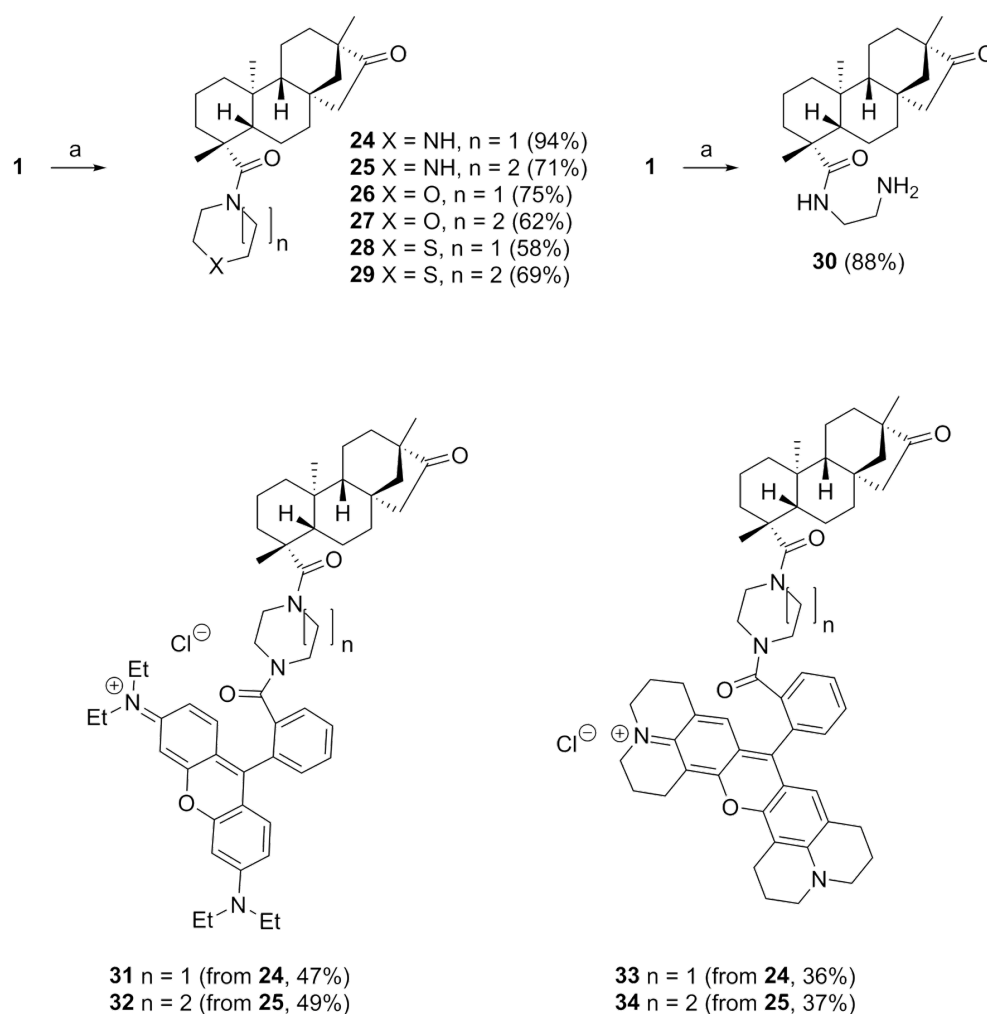
Figure 2. 1,1-ADEQUATE spectrum of **1** in CDCl_3 ; total acquisition time: 20 h.

Compound **1** was transformed in situ with oxalyl chloride into the corresponding acid chloride followed by the addition of the corresponding amine (Scheme 2). The pyridine amides **2–4**, the quinoline amides **8** and **9**, the amides **12–23**, the (homo)-piperazinyl amides **24** and **25** (Scheme 3), the (homo)-morpholinyl amides **26** and **27**, and the (homo)-thiomorpholinyl amides **28** and **29** were obtained; compound **30** was synthesized from **1**

and ethylene diamine in 88% yield. Rhodamine B or rhodamine 101 was activated with oxalyl chloride in the same manner, followed by the reaction with either **26** and **27** to yield **31** and **32** (from rhodamine B) or **33** and **34** (from rhodamine 101), respectively. We refrained from using **30** as a starting material to prepare the corresponding rhodamine conjugates, since it has since become apparent that these conjugates exist preferentially in a non-cationic but neutral spirocyclic form. These electrically neutral molecules proved to be hardly cytotoxic, since they obviously cannot interact with membranes.



Scheme 2. Synthesis of isosteviol derived pyridinyl-, (iso)-quinolinyl- and benzylamides. Reactions and conditions: (a) (COCl)₂, DCM, DMF (cat.), 1 h, 23 °C, then amine in DCM, 23 °C, 1 h; (b) CH₃I, DCM, 23 °C, 2 h.



Scheme 3. Synthesis and structure of amides **24–30** as well as rhodamine B (**31**, **32**) and rhodamine 101 conjugates **33** and **34**. Reactions and conditions: (a) $(\text{COCl})_2$, DCM, DMF (cat.), 1 h, 23 °C, then amine in DCM, 23 °C, 1 h.

For comparison, quaternization was performed with iodomethane, and this provided **5–7** as well as **10** and **11**, respectively.

Isosteviol (**1**) and all derivatives were subjected to sulforhodamine B (SRB) assays employing a panel of human tumor cell lines and non-malignant murine fibroblasts NIH 3T3 and HEK293 cells for comparison. The results from these assays are compiled in Table 1.

The SRB assays showed neither the parent compound isosteviol (**1**) nor almost none of the amides to hold any cytotoxic effect on the human tumor cell lines; they were also non-cytotoxic for the non-malignant cell lines NIH 3T3 and HEK293. The (homo)-piperazinyl amides **24** and **25**, however, showed slight cytotoxic effect, with the homopiperazinyl amide **25** performing slightly better than the piperazinyl-spacered compound **24**. A significant improvement was made with the (homo)-piperazinyl rhodamine-B-spacered compounds **31** and **32**, and the rhodamine 101 hybrids **33** and **34** were even more cytotoxic than the rhodamine B analogs **31** and **32**. The selectivity to distinguish between malignant and non-malignant cell lines, however, was low.

These results emphasize once again that for high cytotoxic activity of terpene/rhodamine hybrids, the interplay between the selected terpene, spacer and lipophilic cation is crucial. If the cationic part is not lipophilic enough (as in compounds **6–7**, **10** and **11**), no cytotoxic activity is obtained. The values obtained for isosteviol derivatives are basically much smaller than those previously obtained for pentacyclic triterpenes. On the other hand, a pronounced cytotoxicity can be achieved also for non-cytotoxic isosteviol if an

appropriate rhodamine residue is added to the terpenoid backbone via a suitable spacer. Once again, the homopiperazinyl spacer proves to be superior to the piperazinyl spacer.

Table 1. Cytotoxicity of compounds: isosteviol (1) and compounds 2–34 (EC₅₀-values in μ M from SRB-assays) after 72 h of treatment; the values are averaged from three independent experiments performed each in triplicate, confidence interval CI = 95%; mean \pm standard mean error; malignant cell lines tested: A375 (melanoma), HT29 (colon adenocarcinoma), MCF-7 (breast adenocarcinoma), A2780 (ovarian carcinoma), HeLa (cervical adenocarcinoma); non-malignant cell lines tested: NIH 3T3 (murine fibroblast), HEK293 (embryonic kidney); positive control: doxorubicin (DOX); n.d. not determined.

	A375	HT29	MCF7	A2780	HeLa	NIH 3T3	HEK293
1	>20	>20	>20	>20	>20	>20	>20
2–23	>20	>20	>20	>20	>20	>20	>20
24	>20	13.5 \pm 2.1	>20	>20	>20	11.2 \pm 1.1	>20
25	14.3 \pm 1.7	11.2 \pm 1.3	11.1 \pm 0.9	13.0 \pm 1.3	>20	11.9 \pm 2.50	>20
26–30	>20	>20	>20	>20	>20	>20	>20
31	0.91 \pm 0.02	0.81 \pm 0.04	0.42 \pm 0.04	0.40 \pm 0.02	0.81 \pm 0.03	1.28 \pm 0.11	0.40 \pm 0.05
32	0.94 \pm 0.15	0.90 \pm 0.06	0.36 \pm 0.06	0.29 \pm 0.08	1.34 \pm 0.9	1.33 \pm 0.09	0.63 \pm 0.07
33	0.44 \pm 0.01	0.42 \pm 0.04	0.23 \pm 0.02	0.19 \pm 0.01	0.45 \pm 0.06	0.59 \pm 0.02	0.20 \pm 0.02
34	0.40 \pm 0.02	0.47 \pm 0.09	0.23 \pm 0.02	0.16 \pm 0.02	0.44 \pm 0.07	0.08 \pm 0.07	0.25 \pm 0.04
DOX	n.d.	0.91 \pm 0.01	1.10 \pm 0.30	0.01 \pm 0.01	n.d.	0.41 \pm 0.07	n.d.

3. Experiment

NMR spectra were recorded using the Varian spectrometers (Darmstadt, Germany) DD2 and VNMRS (400 and 500 MHz, respectively). MS spectra were taken on a Advion expressionL CMS mass spectrometer (Ithaca, USA; positive ion polarity mode, solvent: methanol, solvent flow: 0.2 mL/min, spray voltage: 5.17 kV, source voltage: 77 V, APCI corona discharge: 4.2 μ A, capillary temperature: 250 $^{\circ}$ C, capillary voltage: 180 V, sheath gas: N₂). Thin-layer chromatography was performed on pre-coated silica gel plates supplied by Macherey-Nagel (Düren, Germany). IR spectra were recorded on a Spectrum 1000 FT-IR-spectrometer from Perkin Elmer (Rodgau, Germany). The UV/Vis-spectra were recorded on a Lambda 14 spectrometer from Perkin Elmer (Rodgau, Germany); optical rotations were measured using a JASCO-P2000 instrument (JASCO Germany GmbH, Pfungstadt, Germany). The melting points were determined using the Leica hot stage microscope Galen III (Leica Biosystems, Nussloch, Germany) and are uncorrected. The solvents were dried according to usual procedures. Microanalyses were performed with an Elementar Vario EL (CHNS) instrument (Elementar Analysensysteme GmbH, Elementar-Straße 1, D-63505 Langenselbold, Germany). All dry solvents were distilled over respective drying agents, except for DMF, which was distilled and stored under argon and molecular sieve. Reactions using air- or moisture-sensitive reagents were carried out under argon atmosphere in dried glassware. Triethylamine was stored over potassium hydroxide. Biological assays were performed as previously reported, employing cell lines obtained from the Department of Oncology [Martin-Luther-University Halle Wittenberg; they were bought from ATCC: malignant: A 375, HT29, MCF7 and A2780; non-malignant: NIH 3T3]. Rhodamine B and stevioside were obtained from local vendors and used as received.

For the SRB assay: cells were seeded into 96-well plates on day zero at appropriate cell densities to prevent confluence of the cells during the period of the experiment. After 24 h, the cells were treated with different concentrations (1, 3, 7, 12, 20 and 30 μ M), but the final concentration of DMSO/DMF never exceeded 0.5%, which was non-toxic to the cells. After 72 h of treatment, the supernatant media from the 96-well plates were discarded, and then the cells were fixed with 10% trichloroacetic acid and allowed to rest at 4 $^{\circ}$ C. After 24 h of fixation, the cells were washed in a strip washer and then dyed with SRB solution (200 μ L, 10 mM) for 20 min. Then the plates were washed four times with 1% acetic acid to remove the excess dye and allowed to air-dry overnight. Tris base solution (200 μ L, 10 mM)

was added to each well. The absorbance was measured with a 96-well plate reader from Tecan Spectra.

3.1. General Procedure for the Synthesis of Amides (GPA)

A solution of **1** (1 equiv.) in dry DCM (10 mL) was treated with oxalyl chloride (4 equiv.) and DMF (catal.) for 1 h. The volatiles were evaporated under reduced pressure. To a solution of the residue in dry DCM (10 mL) the corresponding amine (3 equiv.) was added, and the mixture was stirred at room temperature for 1 h. The usual aqueous workup, followed by chromatography, gave amides.

3.2. General Procedure for the Quaternization (GPB)

To a solution of **2–4**, **8**, or **9** in dry DCM (3 mL), iodomethane (3 mL, 0.05 mmol) was added, and the mixture was stirred at room temperature for 2 h. The volatiles were evaporated under reduced pressure, and the residue was subjected to chromatography to afford **5–6**, **10**, and **11**.

3.3. General Procedure for the Synthesis of Rhodamine Conjugates (GPC)

The respective rhodamine was dissolved in dry dichloromethane (10 mL) and mixed with oxalyl chloride (4 eq.) and catalytic amounts of DMF. Following the conditions of GPA as described above, the residue was dissolved in dry DCM (10 mL) and compounds **22** or **23** (3 eq.) were added. Stirring at room temperature was continued for 1 h. The usual aq. work-up followed by chromatography furnished the conjugates **31–34**.

3.4. 16-Oxostachan-18-oic Acid (1, Isosteviol)

A suspension of stevioside (43.0 g, from different local vendors) in MeOH (250 mL) and conc. aq. HCl (43 mL) was heated under reflux for 2 h; stirring at room temperature was continued overnight. Precipitation with water (600 mL) gave a solid that was recrystallized from EtOH (150 mL), and **1** (10.8 g, 63%) was obtained as a colorless solid; $R_f = 0.69$ (SiO₂, CHCl₃/MeOH, 9:1); m.p. 230 °C (lit.: [85] 228–230 °C); $[\alpha]_D^{20} = -84.45^\circ$ ($c = 0.164$, CHCl₃); IR (ATR): $\nu = 2959w, 2922w, 2851w, 1736m, 1690m, 1455w, 1264w$ cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 2.62$ (*dd*, $J = 18.6, 3.7$ Hz, 1H, 15-H), 2.15 (*dt*, $J = 13.1, 4.0$ Hz, 1H, 3-H), 1.91 – 1.79 (*m*, 3H, 2-H, 6-H, 15-H), 1.79 – 1.67 (*m*, 3H, 1-H, 6-H, 11-H), 1.64 (*dt*, $J = 13.3, 3.1$ Hz, 2H, 7-H), 1.62 – 1.57 (*m*, 1H, 12-H), 1.53 (*dd*, $J = 11.6, 2.7$ Hz, 1H, 14-H), 1.48 (*dd*, $J = 13.6, 4.0$ Hz, 1H, 7-H), 1.44 – 1.33 (*m*, 3H, 2-H, 12-H, 14-H), 1.23 (*s*, 3H, 20-H), 1.21 – 1.17 (*m*, 1H, 9-H), 1.14 (*dd*, $J = 12.1, 2.3$ Hz, 1H, 5-H), 1.01 (*td*, $J = 13.6, 4.2$ Hz, 1H, 3-H), 0.96 (*s*, 3H, 17-H), 0.90 (*td*, $J = 13.3, 4.3$ Hz, 1H, 1-H), 0.77 (*s*, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): $\delta = 222.94$ (C-16), 184.16 (C-18), 57.13 (C-5), 54.84 (C-9), 54.38 (C-14), 48.85 (C-13), 48.55 (C-15), 43.79 (C-4), 41.55 (C-7), 39.86 (C-1), 39.59 (C-8), 38.30 (C-10), 37.75 (C-3), 37.42 (C-12), 29.07 (C-20), 21.73 (C-6), 20.45 (C-11), 19.95 (C-17), 18.97 (C-2), 13.42 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 317 (100%, [M-H]⁻).

3.5. 16-Oxo-N-pyridin-2-yl-stachan-18-amide (2)

Following GPA from **1** (300 mg, 0.94 mmol), oxalyl chloride (0.4 mL (4.7 mmol), 2-aminopyridine (354 mg, 3.76 mmol), NEt₃ (0.7 mL, 5.0 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 9:1), **2** (267 mg, 72%) was obtained as a colorless solid; $R_f = 0.16$ (SiO₂, hexanes/ethyl acetate, 8:2); m.p. 185.5 °C; $[\alpha]_D^{20} = -73.47^\circ$ ($c = 0.034$, MeOH); UV-Vis (MeOH): λ_{max} (log ϵ) = 278.52 nm (0.37); IR (ATR) (ATR): $\nu = 3442w, 2926m, 2847m, 1735s, 1683m, 1592w, 1576m, 1504m, 1453w, 1427s, 1295m, 1253w, 1210w, 1177w, 1147m, 1131w, 1109w, 1090w, 1050w, 1016w, 929w, 868w, 778m, 753m, 695w, 666w, 611w, 518w, 410w$ cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 8.50$ (*s*, 1H, N-H), 8.29 – 8.22 (*m*, 2H, 22-H, 25-H), 7.77 (*ddd*, $J = 8.7, 7.3, 1.9$ Hz, 1H, 23-H), 7.08 (*ddd*, $J = 7.3, 5.1, 1.0$ Hz, 1H, 24-H), 2.62 (*dd*, $J = 18.6, 3.8$ Hz, 1H, 15-H), 2.29 (*td*, $J = 14.6, 3.6$ Hz, 1H, 3-H), 2.14 – 2.07 (*m*, 1H, 6-H), 1.94 (*dt*, $J = 13.9, 3.7$ Hz, 1H, 2-H), 1.86 (*dd*, $J = 12.8, 3.0$ Hz, 1H, 6-H), 1.81 (*d*, $J = 18.6$ Hz, 1H, 15-H), 1.78 – 1.77 (*m*, 1H, 1-H), 1.74 (*dt*, $J = 13.4, 3.3$ Hz, 1H, 7-H), 1.71 – 1.67 (*m*, 1H, 11-H),

1.64 – 1.60 (*m*, 1H, 12-H), 1.60 – 1.55 (*m*, 2H, 2-H, 14-H), 1.51 (*dd*, $J = 13.5, 3.7$ Hz, 1H, 7-H), 1.42 (*dd*, $J = 11.6, 3.8$ Hz, 1H, 14-H), 1.40 – 1.35 (*m*, 1H, 12-H), 1.33 (*s*, 3H, 20-H), 1.29 – 1.19 (*m*, 4H, 3-H, 5-H, 9-H, 11-H), 1.02 – 0.98 (*m*, 1H, 1-H), 0.97 (*s*, 3H, 17-H), 0.79 (*s*, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): $\delta = 222.23$ (C-16), 175.98 (C-18), 151.26 (C-21), 146.07 (C-25), 139.83 (C-23), 119.73 (C-24), 114.85 (C-22), 57.83 (C-5), 54.96 (C-9), 54.33 (C-14), 48.82 (C-13), 48.48 (C-15), 45.20 (C-4), 41.74 (C-7), 40.16 (C-1), 39.62 (C-8), 38.34 (C-3), 38.25 (C-10), 37.39 (C-12), 29.73 (C-20), 22.33 (C-6), 20.49 (C-11), 19.96 (C-17), 19.28 (C-2), 13.78 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 , 4:1): m/z (%) = 395 (100%, $[\text{M}+\text{H}]^+$), 811 (70%, $[\text{2M}+\text{Na}]^+$); analysis calcd for $\text{C}_{25}\text{H}_{34}\text{N}_2\text{O}_2$ (394.55): C 76.10, H 8.69, N 7.10; found: C 75.96, H 8.83, 6.95.

3.6. 16-Oxo-N-pyridin-3-yl-stachan-18-amide (3)

Following GPA from **1** (400 mg, 1.3 mmol), oxalyl chloride (0.55 mL, 6.5 mmol), 3-aminopyridine (612 mg, 6.5 mmol), NEt_3 (0.72 mL, 5.2 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **3** (297 mg, 58%) was obtained as a colorless solid; $R_f = 0.68$ (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1); m.p. 162 °C; $[\alpha]_D^{20} = -45.77^\circ$ ($c = 0.043$, MeOH); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 240.10 nm (0.52); IR (ATR): $\nu = 3326\text{br}, 2926\text{m}, 2848\text{m}, 1733\text{m}, 1664\text{m}, 1523\text{m}, 1480\text{m}, 1414\text{m}, 1326\text{w}, 1267\text{w}, 1191\text{w}, 1150\text{w}, 1131\text{w}, 1109\text{w}, 1028\text{w}, 977\text{w}, 802\text{w}, 750\text{s}, 706\text{m}, 665\text{w}, 532\text{w}$ cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 8.79$ (*s*, 1H, N-H), 8.47 – 8.26 (*m*, 2H, 22-H, 25-H), 7.83 (*s*, 1H, 24-H), 7.38 (*s*, 1H, 23-H), 2.62 (*dd*, $J = 18.6, 3.7$ Hz, 1H, 15-H), 2.28 (*d*, $J = 14.5$ Hz, 1H, 3-H), 2.08 (*d*, $J = 13.4$ Hz, 1H, 6-H), 1.96 – 1.65 (*m*, 6H, 1-H, 2-H, 6-H, 7-H, 11-H, 15-H), 1.65 – 1.48 (*m*, 4H, 2-H, 7-H, 12-H, 14-H), 1.42 (*dd*, $J = 11.6, 3.7$ Hz, 1H, 14-H), 1.40 – 1.36 (*m*, 1H, 12-H), 1.34 (*s*, 3H, 20-H), 1.31 – 1.19 (*m*, 4H, 3-H, 5-H, 9-H, 11-H), 1.01 (*dd*, $J = 13.2, 4.3$ Hz, 1H, 1-H), 0.97 (*s*, 3H, 17-H), 0.80 (*s*, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): $\delta = 222.12$ (C-16), 176.11 (C-18), 129.68 (C-22), 124.41 (C-23), 57.88 (C-5), 54.96 (C-9), 54.35 (C-14), 48.83 (C-13), 48.49 (C-15), 45.03 (C-4), 41.77 (C-7), 40.16 (C-1), 39.62 (C-8), 38.33 (C-3), 38.28 (C-10), 37.38 (C-12), 29.87 (C-20), 22.41 (C-6), 20.50 (C-11), 19.97 (C-17), 19.37 (C-2), 13.86 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 , 4:1): m/z (%) = 393 (100%, $[\text{M}-\text{H}]^-$), 429 (30%, $[\text{M}+\text{Cl}]^-$); analysis calcd for $\text{C}_{25}\text{H}_{34}\text{N}_2\text{O}_2$ (394.55): C 76.10, H 8.69, N 7.10; found: C 75.97, H 8.91, N 6.96.

3.7. 16-Oxo-N-pyridin-4-yl-stachan-18-amide (4)

Following GPA from **1** (400 mg, 1.25 mmol), oxalyl chloride (0.54 mL, 6.3 mmol), 4-aminopyridine (593 mg, 6.3 mmol), NEt_3 (0.7 mL, 5 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **4** (335 mg, 68%) was obtained as a colorless solid; $R_f = 0.27$ (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1); m.p. 155 °C; $[\alpha]_D^{20} = -59.56^\circ$ ($c = 0.160$, CHCl_3); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 247.92 nm (0.47); IR (ATR): $\nu = 2926\text{m}, 2845\text{m}, 1732\text{m}, 1669\text{m}, 1585\text{m}, 1504\text{s}, 1454\text{m}, 1412\text{w}, 1325\text{m}, 1284\text{w}, 1209\text{w}, 1127\text{m}, 1089\text{w}, 977\text{w}, 826\text{m}, 750\text{s}, 665\text{w}, 525\text{m}$ cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 8.48$ (*s*, 1H, N-H), 7.99 (*s*, 2H, 22-H, 25-H), 7.77 (*s*, 2H, 24-H, 23-H), 2.61 (*dd*, $J = 18.6, 3.7$ Hz, 1H, 15-H), 2.30 (*d*, $J = 14.4$ Hz, 1H, 3-H), 2.07 (*t*, $J = 12.8$ Hz, 1H, 6-H), 1.89 – 1.77 (*m*, 5H, 1-H, 2-H, 6-H, 7-H, 11-H), 1.72 (*m*, 3H, 2-H, 7-H, 15-H), 1.64 – 1.48 (*m*, 3H, 12-H, 14-H), 1.46 – 1.36 (*m*, 1H, 12-H), 1.34 (*s*, 3H, 20-H), 1.31 – 1.19 (*m*, 5H, 1-H, 3-H, 5-H, 9-H, 11-H), 0.98 (*s*, 3H, 17-H), 0.77 (*s*, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): $\delta = 221.82$ (C-16), 176.29 (C-18), 147.93 (C-21), 147.39 (C-23, C-24), 114.30 (C-22, C-25), 57.70 (C-5), 54.81 (C-9), 54.15 (C-14), 48.67 (C-13), 48.30 (C-15), 45.34 (C-4), 41.58 (C-7), 39.92 (C-1), 39.46 (C-8), 38.17 (C-3), 38.12 (C-10), 37.19 (C-12), 29.46 (C-20), 22.21 (C-6), 20.35 (C-11), 19.80 (C-17), 19.12 (C-2), 13.78 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 , 4:1): m/z (%) = 395 (100%, $[\text{M}+\text{H}]^+$); analysis calcd for $\text{C}_{25}\text{H}_{34}\text{N}_2\text{O}_2$ (394.55): C 76.10, H 8.69, N 7.10; found: C 75.97, H 8.88, N 6.92.

3.8. N-(1-Methylpyridinium-2-yl)-16-oxostachan-18-amide Iodide (5)

Following GPB from **2** (223 mg, 0.56 mmol), iodomethane (3.0 mL, 0.05 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **5** (258 mg, 86%) was obtained as a yellowish solid; $R_f = 0.82$ (SiO_2 , ethyl acetate/ MeOH , 8:2); m.p. 155 °C; $[\alpha]_D^{20} = -114.62^\circ$ ($c = 0.032$, CHCl_3); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 283.98 nm (0.37); IR (ATR): $\nu = 2930\text{m}, 2878\text{m}$,

2832w, 1733s, 1636s, 1596m, 1549m, 1506s, 1444m, 1376s, 1364m, 1320w, 1258w, 1231m, 1177m, 1155s, 1131w, 1105w, 1051w, 974w, 888w, 856m, 783m, 773m, 756m, 730w, 636w, 565w, 533w, 507w, 420w cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 7.84 (*d*, *J* = 9.2 Hz, 1H, N-H), 7.43 (*dd*, *J* = 6.9, 1.7 Hz, 1H, 25-H), 7.37 (*m*, 2H, 22-H, 23-H), 6.35 (*td*, *J* = 6.7, 1.4 Hz, 1H, 24-H), 3.67 (*s*, 3H, 26-H), 2.65 (*dd*, *J* = 18.7, 3.8 Hz, 1H, 15-H), 2.37 (*d*, *J* = 12.9 Hz, 1H, 3-H), 2.03 (*d*, *J* = 10.3 Hz, 1H, 6-H), 1.93 (*ddd*, *J* = 14.2, 11.7, 3.2 Hz, 1H, 12-H), 1.77 (*d*, *J* = 18.7 Hz, 1H, 14-H), 1.73 – 1.47 (*m*, 5H, 1-H, 2-H, 6-H, 7-H, 15-H), 1.48 – 1.32 (*m*, 5H, 2-H, 7-H, 11-H, 12-H, 14-H), 1.24 (*s*, 3H, 20-H), 1.23 – 1.12 (*m*, 4H, 3-H, 5-H, 9-H, 11-H), 0.97 (*s*, 3H, 17-H), 0.87 (*d*, *J* = 7.1 Hz, 1H, 1-H), 0.81 (*s*, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 223.06 (C-16), 187.23 (C-18), 158.17 (C-21), 138.55 (C-23), 138.47 (C-25), 120.20 (C-24), 109.36 (C-22), 58.06 (C-5), 54.78 (C-9), 54.45 (C-14), 48.76 (C-13), 48.59 (C-15), 46.31 (C-26), 42.14 (C-4), 41.15 (C-7), 40.54 (C-1), 39.60 (C-8), 39.07 (C-3), 38.29 (C-10), 37.49 (C-12), 30.49 (C-20), 22.49 (C-6), 20.41 (C-11), 19.90 (C-17), 19.59 (C-2), 14.11 (C-19) ppm; MS (ESI, MeOH/CHCl₃ 4:1): *m/z* (%) = 410 (17%, [M+H-I]⁺); analysis calcd for C₂₆H₃₇N₂O₂I (536.50): C 58.21, H 6.95, N 5.22; found: C 57.97, H 7.13, N 5.01.

3.9. N-(1-Methylpyridinium-3-yl)-16-oxostachan-18-amide Iodide (6)

Following GPB from **3** (233 mg, 0.59 mmol), iodomethane (3.0 mL, 0.05 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 9:1), **6** (294 mg, 93%) was obtained as a yellowish solid; R_f = 0.82 (SiO₂, ethyl acetate/MeOH, 8:2); m.p. 176 °C; [α]_D²⁰ = 50.44° (*c* = 0.154, CHCl₃); UV-Vis (MeOH): λ_{max} (log ε) = 220.38 nm (1.10); IR (ATR): ν = 3328br, 2926m, 2849m, 1731m, 1684m, 1526m, 1507s, 1451m, 1319m, 1241w, 1153w, 1127w, 1106w, 1034w, 963w, 748s, 699s, 592w, 529m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 10.07 (*s*, 1H, 25-H), 9.32 (*d*, *J* = 8.8 Hz, 1H, 22-H), 9.17 (*s*, 1H, N-H), 8.63 (*d*, *J* = 5.8 Hz, 1H, 24-H), 7.90 (*dd*, *J* = 8.7, 5.9 Hz, 1H, 23-H), 4.47 (*s*, 3H, 26-H), 2.66 (*dt*, *J* = 13.8, 2.6 Hz, 1H, 3-H), 2.58 (*dd*, *J* = 18.6, 3.6 Hz, 1H, 15-H), 2.25 – 2.12 (*m*, 1H, 6-H), 1.89 – 1.43 (*m*, 11H, 1-H, 2-H, 6-H, 7-H, 11-H, 12-H, 14-H, 15-H), 1.41 (*s*, 3H, 20-H), 1.39 – 1.11 (*m*, 5H, 3-H, 5-H, 9-H, 11-H, 12-H), 1.02 – 0.97 (*m*, 1H, 1-H), 0.96 (*s*, 3H, 17-H), 0.73 (*s*, 3H, 19-H) ppm; ¹³C NMR (101 MHz, CDCl₃): δ = 222.57 (C-16), 177.86 (C-18), 140.26 (C-21), 138.56 (C-24), 136.93 (C-25), 136.37 (C-22), 127.80 (C-23), 58.11 (C-5), 54.89 (C-9), 54.28 (C-14), 49.30 (C-26), 48.83 (C-13), 48.62 (C-15), 45.76 (C-4), 41.60 (C-7), 39.91 (C-1), 39.62 (C-8), 38.36 (C-3), 38.07 (C-10), 37.37 (C-12), 29.72 (C-20), 22.61 (C-6), 20.49 (C-11), 19.94 (C-17), 19.68 (C-2), 14.29 (C-19) ppm; MS (ESI, MeOH/CHCl₃ 4:1): *m/z* (%) = 410 (16%, [M+H-I]⁺); analysis calcd for C₂₆H₃₇N₂O₂I (536.50): C 58.21, H 6.95, N 5.22; found: C 58.01, H 7.17, N 4.96.

3.10. N-(1-Methylpyridinium-4-yl)-16-oxostachan-18-amide Iodide (7)

Following GPB from **4** (153 mg, 0.39 mmol), iodomethane (3.0 mL, 0.05 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 9:1), **7** (190 mg, 91%) was obtained as a yellowish solid; R_f = 0.82 (SiO₂, ethyl acetate/MeOH, 8:2); m.p. 197 °C; [α]_D²⁰ = −58.15° (*c* = 0.149, CHCl₃); UV-Vis (MeOH): λ_{max} (log ε) = 275.94 nm (0.76); IR (ATR): ν = 2926m, 2849m, 1730m, 1671m, 1641m, 1586m, 1515s, 1446m, 1317m, 1198m, 1174w, 1120m, 1086m, 1023w, 977w, 845w, 748s, 661w, 519m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 9.48 (*s*, 1H, N-H), 8.90 – 8.83 (*m*, 2H, 23-H, 24-H), 8.70 – 8.63 (*m*, 2H, 22-H, 25-H), 4.42 (*s*, 3H, 26-H), 2.75 (*d*, *J* = 14.6 Hz, 1H, 15-H), 2.57 (*dd*, *J* = 18.6, 3.6 Hz, 2H, 1-H, 3-H), 2.22 (*m*, 2H, 6-H, 15-H), 1.86 – 1.63 (*m*, 8H, 2-H, 6-H, 7-H, 11-H, 12-H, 14-H), 1.63 – 1.48 (*m*, 3H, 7-H, 11-H, 14-H), 1.46 (*s*, 3H, 20-H), 1.44 – 1.30 (*m*, 2H, 5-H, 9-H), 1.27 – 1.12 (*m*, 2H, 1-H, 3-H), 0.96 (*s*, 3H, 17-H), 0.70 (*s*, 3H, 19-H) ppm; ¹³C NMR (101 MHz, CDCl₃): δ = 222.33 (C-16), 178.30 (C-18), 152.95 (C-21), 144.44 (C-23, C-24), 116.98 (C-22, C-25), 58.10 (C-5), 54.84 (C-9), 54.13 (C-14), 48.67 (C-13), 48.44 (C-15), 47.49 (C-26), 46.45 (C-4), 41.46 (C-7), 39.69 (C-1), 39.46 (C-8), 38.21 (C-10), 38.11 (C-3), 37.22 (C-12), 29.16 (C-20), 22.37 (C-6), 20.33 (C-11), 19.80 (C-17), 19.43 (C-2), 14.23 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) = 410 (36%, [M+H-I]⁺); analysis calcd for C₂₆H₃₇N₂O₂I (536.50): C 58.21, H 6.95, N 5.22; found: C 58.00, H 7.19, N 4.86.

3.11. 16-Oxo-N-isoquinolin-4-yl-stachan-18-amide (8)

Following GPA (microwave-assisted) from **1** (250 mg, 0.78 mmol), oxalyl chloride (0.3 mL, 3.12 mmol), 4-amino-isoquinoline (337 mg, 2.34 mmol), NEt_3 (0.3 mL, 2.34 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **8** (250 mg, 72%) was obtained as a colorless solid; $R_f = 0.17$ (SiO_2 , hexanes/ethyl acetate, 1:1); m.p. 91 °C; $[\alpha]_D^{20} = -39.67^\circ$ ($c = 0.031$, MeOH); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 217.81 nm (2.45); IR (ATR): $\nu = 3320w$, 2924m, 2847m, 1734s, 1653m, 1586w, 1511m, 1488s, 1451s, 1410m, 1391m, 1321w, 1278w, 1254w, 1225w, 1169w, 1110w, 1017w, 976w, 883w, 861w, 778m, 749s, 632w, 579w, 507w, 491w, 464w cm^{-1} ; $^1\text{H NMR}$ (500 MHz, CDCl_3): $\delta = 9.13$ (s, 1H, 23-H), 8.88 (s, 1H, 22-H), 8.09 (d, $J = 8.2$ Hz, 1H, 25-H), 7.97 (s, 1H, N-H), 7.88 – 7.80 (m, 2H, 27-H, 28-H), 7.71 (ddd, $J = 8.1$, 5.9, 2.0 Hz, 1H, 26-H), 2.63 (dd, $J = 18.6$, 3.7 Hz, 1H, 15-H), 2.38 (dt, $J = 15.3$, 3.7 Hz, 1H, 3-H), 2.14 – 2.09 (m, 1H, 6-H), 2.02 (dt, $J = 13.9$, 3.1 Hz, 1H, 2-H), 1.93 (dd, $J = 12.4$, 2.9 Hz, 1H, 6-H), 1.88 – 1.83 (m, 1H, 1-H), 1.80 (d, $J = 18.6$ Hz, 1H, 15-H), 1.76 – 1.69 (m, 2H, 7-H, 11-H), 1.69 – 1.66 (m, 1H, 2-H), 1.66 – 1.62 (m, 1H, 12-H), 1.61 – 1.51 (m, 2H, 7-H, 14-H), 1.47 (s, 3H, 20-H), 1.44 (dd, $J = 11.7$, 3.8 Hz, 1H, 14-H), 1.41 – 1.36 (m, 2H, 3-H, 12-H), 1.31 (dd, $J = 12.3$, 2.1 Hz, 1H, 5-H), 1.29 – 1.21 (m, 2H, 9-H, 11-H), 1.05 (td, $J = 13.6$, 4.7 Hz, 1H, 1-H), 0.98 (s, 3H, 17-H), 0.89 (s, 3H, 19-H) ppm; $^{13}\text{C NMR}$ (126 MHz, CDCl_3): $\delta = 222.10$ (C-16), 176.10 (C-18), 147.58 (C-23), 135.05 (C-22), 132.33 (C-27), 131.60 (C-21), 129.86 (C-29), 129.08 (C-25), 128.61 (C-24), 128.47 (C-26), 120.97 (C-28), 57.80 (C-5), 54.94 (C-9), 54.34 (C-14), 48.82 (C-13), 48.44 (C-15), 45.17 (C-4), 41.79 (C-7), 40.21 (C-1), 39.63 (C-8), 38.54 (C-3), 38.38 (C-10), 37.36 (C-12), 30.35 (C-20), 22.49 (C-6), 20.51 (C-11), 19.96 (C-17), 19.50 (C-2), 14.19 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 4:1): m/z (%) = 443 (100%, $[\text{M-H}]^-$); analysis calcd for $\text{C}_{29}\text{H}_{36}\text{N}_2\text{O}_2$ (444.62): C 78.34, H 8.16, N 6.30; found: C 78.16, H 7.95, N 6.15.

3.12. 16-Oxo-N-quinolin-5-yl-stachan-18-amide (9)

Following GPA (microwave-assisted) from **1** (250 mg (0.78 mmol), oxalyl chloride (0.3 mL (3.12 mmol), 5-aminoquinoline (337 mg, 2.34 mmol), NEt_3 (0.3 mL, 2.34 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **9** (187 mg, 54%) was obtained as a colorless solid; $R_f = 0.15$ (SiO_2 , hexanes/ethyl acetate, 1:1); m.p. 90 °C; $[\alpha]_D^{20} = -35.71^\circ$ ($c = 0.056$, MeOH); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 229.71 nm (0.86); IR (ATR): $\nu = 3337w$, 2924m, 2847m, 1734m, 1649m, 1594w, 1510w, 1485m, 1452m, 1397w, 1318w, 1261w, 1171w, 1131w, 1110w, 976w, 862w, 798s, 750s, 656w, 589w, 498w, 467w cm^{-1} ; $^1\text{H NMR}$ (500 MHz, CDCl_3): $\delta = 8.78$ (dd, $J = 4.5$, 1.5 Hz, 1H, 26-H), 8.31 (d, $J = 8.4$ Hz, 1H, 22-H), 8.02 (d, $J = 8.1$ Hz, 1H, N-H), 7.77 (dd, $J = 7.6$, 1.1 Hz, 2H, 27-H, 28-H), 7.71 (dd, $J = 8.4$, 7.5 Hz, 1H, 24-H), 7.49 (dd, $J = 8.5$, 4.4 Hz, 1H, 23-H), 2.64 (dd, $J = 18.5$, 3.7 Hz, 1H, 15-H), 2.37 (d, $J = 14.4$ Hz, 1H, 3-H), 2.09 (td, $J = 13.8$, 6.9 Hz, 1H, 6-H), 2.05 – 1.84 (m, 3H, 1-H, 2-H, 6-H), 1.80 (d, $J = 18.6$ Hz, 1H, 15-H), 1.77 – 1.59 (m, 4H, 2-H, 7-H, 11-H, 12-H), 1.60 – 1.51 (m, 2H, 7-H, 14-H), 1.47 (s, 3H, 20-H), 1.44 – 1.33 (m, 3H, 3-H, 12-H, 14-H), 1.34 – 1.19 (m, 3H, 5-H, 9-H, 11-H), 1.05 (td, $J = 13.2$, 4.4 Hz, 1H, 1-H), 0.98 (s, 3H, 17-H), 0.91 (s, 3H, 19-H) ppm; $^{13}\text{C NMR}$ (126 MHz, CDCl_3): $\delta = 222.03$ (C-16), 176.18 (C-18), 148.08 (C-23), 133.47 (C-22), 130.62 (C-27), 125.31 (C-21), 124.06 (C-29), 123.41 (C-25, C-26), 120.78 (C-24, C-28), 57.72 (C-5), 54.77 (C-9), 54.20 (C-14), 48.69 (C-13), 48.34 (C-15), 44.89 (C-4), 41.66 (C-7), 40.09 (C-1), 39.51 (C-8), 38.38 (C-3), 38.24 (C-10), 37.24 (C-12), 30.28 (C-20), 22.36 (C-6), 20.38 (C-11), 19.82 (C-17), 19.40 (C-2), 14.09 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 , 4:1): m/z (%) = 445 (90%, $[\text{M+H}]^+$), 911 (100%, $[\text{2M+Na}]^+$); analysis calcd for $\text{C}_{29}\text{H}_{36}\text{N}_2\text{O}_2$ (444.62): C 78.34, H 8.16, N 6.30; found: C 78.07, H 8.36, N 6.13.

3.13. N-(1-Methylisoquinolinium-4-yl)-16-oxostachan-18-amide Iodide (10)

Following GPB from **8** (97 mg, 0.22 mmol), iodomethane (3.0 mL, 0.05 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **10** (93 mg, 72%) was obtained a yellowish solid; $R_f = 0.15$ (SiO_2 , hexanes/ethyl acetate, 1:1); m.p. 188 °C; $[\alpha]_D^{20} = -46.89^\circ$ ($c = 0.110$, CHCl_3); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 228.82 nm (1.50); IR (ATR): $\nu = 3225w$, 3078w, 2926m, 2841m, 1730s, 1690m, 1643w, 1608w, 1519m, 1499m, 1472m, 1445s, 1415w, 1352m, 1259m, 1219w, 1187w, 1157m, 1133w, 1109w, 980w, 868w, 783m, 752w, 638w, 590m, 553w,

524w, 512w, 445w, 425w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 10.40 (s, 1H, 23-H), 9.06 (d, J = 1.4 Hz, 1H, 22-H), 8.76 (s, 1H, N-H), 8.54 (d, J = 8.3 Hz, 1H, 28-H), 8.19 – 8.08 (m, 2H, 25-H, 26-H), 7.92 (ddd, J = 8.2, 4.9, 3.1 Hz, 1H, 27-H), 4.52 (s, 3H, 30-H), 2.65 – 2.53 (m, 2H, 3-H, 15-H), 2.21 – 2.12 (m, 1H, 6-H), 1.97 – 1.92 (m, 1H, 2-H), 1.89 (dd, J = 12.8, 2.9 Hz, 1H, 6-H), 1.81 (d, J = 18.7 Hz, 1H, 15-H), 1.81 – 1.65 (m, 4H, 1-H, 2-H, 7-H, 11-H), 1.63 – 1.52 (m, 3H, 7-H, 12-H, 14-H), 1.49 (s, 3H, 20-H), 1.43 (dd, J = 11.8, 3.7 Hz, 1H, 14-H), 1.40 – 1.34 (m, 2H, 3-H, 12-H), 1.32 (dd, J = 12.4, 2.0 Hz, 1H, 5-H), 1.28 – 1.22 (m, 1H, 9-H), 1.19 (dd, J = 12.7, 5.2 Hz, 1H, 11-H), 1.04 (td, J = 13.1, 4.3 Hz, 1H, 1-H), 0.96 (s, 3H, 17-H), 0.79 (s, 3H, 19-H) ppm; ¹³C NMR (101 MHz, CDCl₃): δ = 222.21 (C-16), 177.11 (C-18), 145.95 (C-23), 136.91 (C-26), 134.29 (C-29), 131.83 (C-21), 131.53 (C-27), 131.31 (C-28), 129.06 (C-22), 127.99 (C-24), 122.37 (C-25), 57.92 (C-5), 54.83 (C-9), 54.24 (C-14), 48.89 (C-30), 48.82 (C-13), 48.49 (C-15), 45.73 (C-4), 41.66 (C-7), 39.97 (C-1), 39.63 (C-8), 38.37 (C-3), 38.32 (C-10), 37.32 (C-12), 30.19 (C-20), 22.54 (C-6), 20.49 (C-11), 19.93 (C-17), 19.64 (C-2), 14.48 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 460 (21%, [M+H-I]⁺); analysis calcd for C₃₀H₃₉N₂O₂I (586.56): C 61.43, H 6.70, N 4.78; found: C 71.20, H 6.97, N 4.44.

3.14. N-(1-Methylquinolinium-5-yl)-16-oxostachan-18-amide Iodide (11)

Following GPB from **9** (60 mg, 0.14 mmol), iodomethane (3.0 mL, 0.05 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 9:1), **11** (65 mg, 79%) was obtained as a yellowish solid; R_f = 0.53 (SiO₂, CHCl₃/MeOH, 8:2); m.p. 195 °C; [α]_D²⁰ = -61.93° (c = 0.108, MeOH); UV-Vis (MeOH): λ_{max} (log ε) = 219.17 nm (0.92); IR (ATR): ν = 3428m, 2924m, 2848m, 1732s, 1667w, 1622m, 1592m, 1533m, 1492m, 1450s, 1406w, 1368w, 1336w, 1279w, 1244m, 1161m, 1128w, 1111w, 1088w, 1029w, 977w, 929w, 862w, 793s, 750m, 696w, 559w, 527m, 463m cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 9.76 (d, J = 5.7 Hz, 1H, 28-H), 9.28 (d, J = 8.6 Hz, 1H, 26-H), 8.77 (s, 1H, N-H), 8.22 – 8.05 (m, 2H, 22-H, 27-H), 7.98 (dd, J = 8.6, 5.7 Hz, 2H, 23-H, 24-H), 4.71 (s, 3H, 30-H), 2.72 – 2.61 (m, 1H, 15-H), 2.59 (d, J = 3.6 Hz, 1H, 3-H), 2.16 (d, J = 13.4 Hz, 1H, 6-H), 2.00 – 1.78 (m, 4H, 1-H, 2-H, 6-H, 15-H), 1.77 – 1.55 (m, 6H, 2-H, 7-H, 11-H, 12-H, 14-H), 1.52 (s, 3H, 20-H), 1.49 – 1.30 (m, 3H, 3-H, 12-H, 14-H), 1.30 – 1.18 (m, 3H, 5-H, 9-H, 11-H), 1.05 (td, J = 13.4, 4.4 Hz, 1H, 1-H), 0.98 (s, 3H, 17-H), 0.87 (s, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.33 (C-16), 177.27 (C-18), 149.55 (C-23), 144.31 (C-22), 139.08 (C-27), 136.96 (C-21), 136.19 (C-29), 127.35 (C-26), 126.15 (C-25), 120.98 (C-24), 115.24 (C-28), 57.94 (C-5), 54.77 (C-9), 54.19 (C-14), 48.71 (C-13, C-15), 48.43 (C-30), 46.84 (C-4), 45.23 (C-7), 41.61 (C-1), 39.97 (C-8), 39.53 (C-3), 38.27 (C-10), 37.27 (C-12), 30.20 (C-20), 22.46 (C-6), 20.39 (C-11), 19.83 (C-17), 19.80 (C-2), 14.38 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 460 (35%, [M+H-I]⁺); analysis calcd for C₃₀H₃₉N₂O₂I (586.56): C 61.43, H 6.70, N 4.78; found: C 61.26, H 6.91, N 4.55.

3.15. 16-Oxo-N-phenyl-stachan-18-amide (12)

Following GPA from **1** (300 mg, 0.94 mmol), aniline (0.35 mL, 3.76 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 9:1), **12** (237 mg, 64%) was obtained as an off-white solid; m.p. = 65 °C; [α]_D²⁰ = -57.68° (c = 0.108, MeOH); UV-Vis (MeOH): λ_{max} (log ε) = 238.27 nm (1.23); IR (ATR): ν = 3371w, 2925m, 2847m, 1730s, 1667m, 1595w, 1519w, 1499s, 1434s, 1306m, 1237w, 1150w, 1131w, 1109w, 1028w, 976w, 748s, 691s, 596w, 527m cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 7.47 – 7.43 (m, 2H, 22-H, 26-H), 7.33 – 7.27 (m, 2H, 23-H, 25-H), 7.12 – 7.07 (m, 1H, 24-H), 6.68 – 6.71 (m, 1H, N-H), 2.63 (dd, J = 18.6, 3.8 Hz, 1H, 15-H), 2.18 (dd, J = 14.6, 2.1 Hz, 1H, 3-H), 2.08 – 2.01 (m, 1H, 6-H), 1.95 – 1.89 (m, 1H, 2-H), 1.86 (dd, J = 12.7, 3.1 Hz, 1H, 6-H), 1.81 (s, 1H, 1-H), 1.80 (d, J = 18.7 Hz, 1H, 15-H), 1.72 (dt, J = 13.6, 3.4 Hz, 1H, 7-H), 1.70 – 1.67 (m, 1H, 11-H), 1.63 – 1.59 (m, 1H, 12-H), 1.57 – 1.47 (m, 3H, 2-H, 7-H, 14-H), 1.41 – 1.37 (m, 1H, 12-H), 1.30 (s, 3H, 20-H), 1.27 – 1.19 (m, 4H, 3-H, 5-H, 9-H, 11-H), 1.03 – 0.99 (m, 1H, 1-H), 0.98 (s, 3H, 17-H), 0.82 (s, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.07 (C-16), 174.88 (C-18), 137.82 (C-21), 129.00 (C-23, C-25), 124.35 (C-24), 120.41 (C-22, C-26), 57.69 (C-5), 54.80 (C-9), 54.21 (C-14), 48.69 (C-13), 48.32 (C-15), 44.53 (C-4), 41.69 (C-7), 40.14 (C-1), 39.49 (C-8), 38.27 (C-3), 38.17 (C-10), 37.25 (C-12), 29.91 (C-20), 22.30 (C-6), 20.36 (C-11), 19.84 (C-17), 19.17 (C-2), 13.62 (C-19) ppm;

MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 392 (100%, [M-H]⁻); analysis calcd for C₂₆H₃₅NO₂ (393.57): C 79.35, H 8.96, N 3.56; found: C 79.17, H 9.18, N 3.23.

3.16. *N*-Benzyl-16-oxostachan-18-amide (13)

Following GPA from **1** (250 mg, 0.78 mmol), benzylamine (0.44 mL, 4.0 mmol), and chromatography (SiO₂, hexanes/ethyl acetate, 95:5), **13** (210 mg, 66%) was obtained as a colorless solid; R_f = 0.28 (SiO₂, hexanes/ethyl acetate, 8:2); m.p. = 71 °C; $[\alpha]_D^{20}$ = -65.44° (c = 0.122, MeOH); IR (ATR): ν = 3371 w , 2925 m , 2847 m , 1730 s , 1667 m , 1595 w , 1519 w , 1499 s , 1434 s , 1306 m , 1237 w , 1150 w , 1131 w , 1109 w , 1028 w , 976 w , 748 s , 691 s , 596 w , 527 m cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 7.36 – 7.26 (m, 5H, 23-H, 24-H, 25-H, 26-H, 27-H), 5.86 (t, J = 5.5 Hz, 1H, N-H), 4.40 (dd, J = 5.5, 1.7 Hz, 2H, 21-H), 2.63 (dd, J = 18.7, 3.8 Hz, 1H, 15-H), 2.06 – 2.00 (m, 1H, 3-H), 1.97 – 1.92 (m, 1H, 6-H), 1.84 – 1.71 (m, 4H, 1-H, 2-H, 6-H, 15-H), 1.69 – 1.63 (m, 2H, 7-H, 11-H), 1.62 – 1.57 (m, 1H, 12-H), 1.54 (dd, J = 11.6, 2.7 Hz, 1H, 14-H), 1.50 – 1.42 (m, 2H, 2-H, 7-H), 1.41 – 1.32 (m, 2H, 12-H, 14-H), 1.21 (s, 3H, 20-H), 1.20 – 1.10 (m, 4H, 3-H, 5-H, 9-H, 11-H), 0.97 (s, 3H, 17-H), 0.96 – 0.89 (m, 1H, 1-H), 0.74 (s, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.50 (C-16), 176.51 (C-18), 138.64 (C-22), 128.85 (C-24, C-26), 128.11 (C-23, C-27), 127.63 (C-25), 57.77 (C-5), 54.89 (C-9), 54.41 (C-14), 48.84 (C-13), 48.52 (C-15), 43.85 (C-4, C-21), 41.84 (C-7), 40.30 (C-1), 39.63 (C-8), 38.27 (C-3), 38.23 (C-10), 37.43 (C-12), 30.34 (C-20), 22.40 (C-6), 20.47 (C-11), 19.99 (C-17), 19.36 (C-2), 13.74 (C-19) ppm; MS (ESI, MeOH/CHCl₃ 4:1): m/z (%) = 408 (100%, [M+H]⁺), 430 (95%, [M+Na]⁺), 837 (80%, [2M+Na]⁺); analysis calcd for C₂₇H₃₇NO₂ (407.60): C 79.56, H 9.15, N 3.44; found: C 79.31, H 9.36, N 3.20.

3.17. 16-Oxo-*N*-(2-phenylethyl)-stachan-18-amide (14)

Following GPA from **1** (500 mg, 1.57 mmol), phenethylamine (0.8 mL, 6.28 mmol), and chromatography (SiO₂, hexanes/ethyl acetate, 95:5), **14** (470 mg, 71%) was obtained as a colorless solid; R_f = 0.18 (SiO₂, hexanes/ethyl acetate, 8:2); m.p. = 82 °C; $[\alpha]_D^{20}$ = -53.68° (c = 0.106, MeOH); UV-Vis (MeOH): λ_{max} (log ϵ) = 213.81 nm (3.41); IR (ATR): ν = 3371 w , 2925 m , 2847 m , 1730 s , 1667 m , 1595 w , 1519 w , 1499 s , 1434 s , 1306 m , 1237 w , 1150 w , 1131 w , 1109 w , 1028 w , 976 w , 748 s , 691 s , 596 w , 527 m cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 7.34 – 7.07 (m, 5H, 24-H, 25-H, 26-H, 27-H, 28-H), 5.56 (s, 1H, N-H), 3.57 – 3.43 (m, 2H, 21-H), 2.88 – 2.72 (m, 2H, 22-H), 2.55 (dd, J = 18.7, 3.7 Hz, 1H, 15-H), 1.94 (d, J = 14.1 Hz, 1H, 3-H), 1.82 – 1.76 (m, 1H, 6-H), 1.72 (d, J = 18.7 Hz, 1H, 15-H), 1.69 – 1.60 (m, 3H, 1-H, 2-H, 11-H), 1.59 – 1.53 (m, 3H, 6-H, 7-H, 12-H), 1.50 (dd, J = 11.6, 2.7 Hz, 2H, 14-H), 1.41 (dd, J = 13.2, 3.3 Hz, 1H, 7-H), 1.38 – 1.29 (m, 3H, 2-H, 12-H, 14-H), 1.17 (dd, J = 12.9, 5.2 Hz, 1H, 11-H), 1.14 – 1.12 (m, 1H, 9-H), 1.10 (s, 3H, 20-H), 1.06 – 1.02 (m, 2H, 3-H, 5-H), 0.95 (s, 3H, 17-H), 0.85 (td, J = 13.4, 12.0, 6.9 Hz, 1H, 1-H), 0.66 (s, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.36 (C-16), 176.76 (C-18), 139.03 (C-23), 128.80 (C-24, C-25, C-27, C-28), 126.65 (C-26), 57.44 (C-5), 54.78 (C-9), 54.30 (C-14), 48.73 (C-13), 48.54 (C-15), 43.70 (C-4), 41.73 (C-7), 40.52 (C-21), 40.23 (C-1), 39.51 (C-8), 38.17 (C-3), 38.11 (C-10), 37.34 (C-12), 35.25 (C-22), 30.24 (C-20), 22.14 (C-6), 20.37 (C-11), 19.92 (C-17), 19.12 (C-2), 13.52 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 420 (100%, [M-H]⁻); analysis calcd for C₂₈H₃₉NO₂ (421.63): C 79.63, H 9.32, N 3.32; found: C 79.41, H 9.57, N 3.01.

3.18. *N*-(2-Fluorobenzyl)-16-oxostachan-18-amide (15)

Following GPA from **1** (300 mg, 0.94 mmol), 2-fluorobenzylamine (0.32 mL, 2.82 mmol), and chromatography (SiO₂, hexanes/ethyl acetate, 95:5), **15** (368 mg, 92%) was obtained as a colorless solid; R_f = 0.32 (SiO₂, hexanes/ethyl acetate, 8:2); m.p. = 60 °C; $[\alpha]_D^{20}$ = -51.6° (c = 0.05, MeOH); UV-Vis (MeOH): λ_{max} (log ϵ) = 238.27 nm (1.23); IR (ATR): ν = 3371 w , 2925 m , 2847 m , 1730 s , 1667 m , 1595 w , 1519 w , 1499 s , 1434 s , 1306 m , 1237 w , 1150 w , 1131 w , 1109 w , 1028 w , 976 w , 748 s , 691 s , 596 w , 527 m cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 7.38 – 7.33 (m, 1H, 23-H), 7.28 – 7.22 (m, 1H, 25-H), 7.10 – 7.06 (m, 1H, 24-H), 7.06 – 7.01 (m, 1H, 26-H), 6.00 (t, J = 5.7 Hz, 1H, N-H), 4.46 – 4.37 (m, 2H, 21-H), 2.56 (dd, J = 18.7, 3.8 Hz, 1H, 15-H), 2.06 – 2.01 (m, 1H, 3-H), 1.95 – 1.90 (m, 1H, 6-H), 1.82 – 1.73

(m, 3H, 2-H, 6-H, 15-H), 1.72 – 1.61 (m, 3H, 1-H, 7-H, 11-H), 1.60 – 1.55 (m, 1H, 12-H), 1.52 (dd, $J = 11.6, 2.8$ Hz, 1H, 14-H), 1.49 – 1.42 (m, 2H, 2-H, 7-H), 1.38 (dd, $J = 11.6, 3.9$ Hz, 1H, 14-H), 1.35 – 1.30 (m, 1H, 12-H), 1.17 (s, 3H, 20-H), 1.16 – 1.12 (m, 3H, 3-H, 9-H, 11-H), 1.10 (dd, $J = 12.2, 2.2$ Hz, 1H, 5-H), 0.96 (s, 3H, 17-H), 0.94 – 0.85 (m, 1H, 1-H), 0.55 (s, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): ^{13}C NMR (126 MHz, CDCl_3): $\delta = 222.55$ (C-16), 176.57 (C-18), 161.36 ($d, J = 245.4$ Hz, C-27), 131.14 ($d, J = 4.4$ Hz, C-23), 129.47 ($d, J = 8.2$ Hz, C-25), 125.37 ($d, J = 14.7$ Hz, C-22), 124.40 ($d, J = 3.8$ Hz, C-24), 115.37 ($d, J = 21.2$ Hz, C-26), 57.71 (C-5), 54.84 (C-9), 54.39 (C-14), 48.83 (C-13), 48.47 (C-15), 43.85 (C-4), 41.82 (C-7), 40.27 (C-1), 39.61 (C-10), 38.20 (C-3), 38.15 (C-8), 38.10 (C-21), 37.43 (C-12), 30.19 (C-20), 22.27 (C-6), 20.44 (C-11), 19.98 (C-17), 19.22 (C-2), 13.27 (C-19) ppm; ^{19}F NMR (470 MHz, CDCl_3): $\delta = -119.37 - 119.47$ (m) ppm; MS (ESI, MeOH/ CHCl_3 , 4:1): m/z (%) = 424 (100%, $[\text{M}-\text{H}]^-$); analysis calcd for $\text{C}_{27}\text{H}_{36}\text{FNO}_2$ (425.59): C 76.20, H 8.53, N 3.29; found: C 75.96, H 8.77, N 2.97.

3.19. *N*-(3-Fluorobenzyl)-16-oxostachan-18-amide (16)

Following GPA from **1** (300 mg, 0.94 mmol), 3-fluorobenzylamine (0.32 mL, 2.82 mmol), and chromatography (SiO_2 , hexanes/ethyl acetate, 95:5), **16** (304 mg, 38%) was obtained as a colorless solid; $R_f = 0.32$ (SiO_2 , hexanes/ethyl acetate, 8:2); m.p. = 60 °C; $[\alpha]_D^{20} = -51.6^\circ$ ($c = 0.05$, MeOH); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 238.27 nm (1.23); IR (ATR): $\nu = 3371w, 2925m, 2847m, 1730s, 1667m, 1595w, 1519w, 1499s, 1434s, 1306m, 1237w, 1150w, 1131w, 1109w, 1028w, 976w, 748s, 691s, 596w, 527m \text{ cm}^{-1}$; ^1H NMR (500 MHz, CDCl_3): $\delta = 7.31 - 7.26$ (m, 1H, 24-H), 7.07 – 7.02 (m, 1H, 23-H), 6.99 – 6.92 (m, 2H, 25-H, 27-H), 5.93 (t, $J = 5.8$ Hz, 1H, N-H), 4.44 – 4.34 (m, 2H, 21-H), 2.61 (dd, $J = 18.6, 3.8$ Hz, 1H, 15-H), 2.07 – 2.01 (m, 1H, 3-H), 1.97 – 1.92 (m, 1H, 6-H), 1.83 – 1.72 (m, 4H, 1-H, 2-H, 6-H, 15-H), 1.70 – 1.64 (m, 2H, 7-H, 11-H), 1.62 – 1.57 (m, 1H, 12-H), 1.54 (dd, $J = 11.6, 2.8$ Hz, 1H, 14-H), 1.50 – 1.43 (m, 2H, 2-H, 7-H), 1.40 (dd, $J = 11.7, 4.0$ Hz, 1H, 14-H), 1.35 (dd, $J = 12.6, 5.2$ Hz, 1H, 12-H), 1.21 (s, 3H, 3-H, 9-H, 11-H), 1.27 – 1.16 (m, 3H, 20-H), 1.13 (dd, $J = 12.3, 2.2$ Hz, 1H, 5-H), 0.96 (s, 3H, 17-H), 0.95 – 0.88 (m, 1H, 1-H), 0.72 (s, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): $\delta = 222.44$ (C-16), 176.65 (C-18), 164.09 ($d, J = 246.4$ Hz, C-26), 141.30 ($d, J = 7.1$ Hz, C-22), 130.33 ($d, J = 8.1$ Hz, C-24), 123.53 ($d, J = 2.9$ Hz, C-23), 114.86 ($d, J = 21.5$ Hz, C-27), 114.47 ($d, J = 21.0$ Hz, C-25), 57.73 (C-5), 54.88 (C-9), 54.39 (C-14), 48.83 (C-13), 48.50 (C-15), 43.91 (C-4), 43.24 (C-21), 41.82 (C-7), 40.26 (C-1), 39.61 (C-8), 38.25 (C-3), 38.22 (C-10), 37.42 (C-12), 30.33 (C-20), 22.40 (C-6), 20.47 (C-11), 19.98 (C-17), 19.35 (C-2), 13.74 (C-19) ppm; ^{19}F NMR (470 MHz, CDCl_3): $\delta = -112.73$ (td, $J = 9.2, 5.9$ Hz) ppm; MS (ESI, MeOH/ CHCl_3 , 4:1): m/z (%) = 424 (100%, $[\text{M}-\text{H}]^-$); analysis calcd for $\text{C}_{27}\text{H}_{36}\text{FNO}_2$ (425.59): C 76.20, H 8.53, N 3.29; found: C 75.86, H 8.71, N 3.11.

3.20. *N*-(4-Fluorobenzyl)-16-oxostachan-18-amide (17)

Following GPA from **1** (250 mg, 0.78 mmol), 4-fluoro-benzylamine (0.27 mL, 2.34 mmol), and chromatography (SiO_2 , hexanes/ethyl acetate, 95:5), **17** (302 mg, 91%) was obtained as a colorless solid; $R_f = 0.2$ (SiO_2 , hexanes/ethyl acetate, 8:2); m.p. = 62 °C; $[\alpha]_D^{20} = -87.89^\circ$ ($c = 0.09$, MeOH); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 211 nm (3.10); IR (ATR): $\nu = 3371w, 2925m, 2847m, 1730s, 1667m, 1595w, 1519w, 1499s, 1434s, 1306m, 1237w, 1150w, 1131w, 1109w, 1028w, 976w, 748s, 691s, 596w, 527m \text{ cm}^{-1}$; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.27 - 7.20$ (m, 2H, 23-H, 27-H), 7.05 – 6.93 (m, 2H, 24-H, 26-H), 5.88 (t, $J = 5.7$ Hz, 1H, N-H), 4.43 – 4.29 (m, 2H, 21-H), 2.62 (dd, $J = 18.6, 3.7$ Hz, 1H, 15-H), 2.06 – 1.98 (m, 1H, 3-H), 1.98 – 1.90 (m, 1H, 6-H), 1.83 – 1.63 (m, 6H, 1-H, 2-H, 6-H, 7-H, 11-H, 15-H), 1.62 – 1.57 (m, 1H, 12-H), 1.54 (dd, $J = 11.6, 2.7$ Hz, 1H, 14-H), 1.50 – 1.31 (m, 4H, 2-H, 7-H, 12-H, 14-H), 1.28 – 1.21 (m, 2H, 3-H, 11-H), 1.20 (s, 3H, 20-H), 1.18 (s, 1H, 9-H), 1.13 (dd, $J = 12.1, 2.2$ Hz, 1H, 5-H), 0.96 (s, 3H, 17-H), 0.90 (dd, $J = 13.8, 3.9$ Hz, 1H, 1-H), 0.72 (s, 3H, 19-H) ppm; ^{13}C NMR (101 MHz, CDCl_3): $\delta = 222.45$ (C-16), 176.55 (C-18), 162.28 ($d, J = 245.7$ Hz, C-25), 134.50 ($d, J = 3.0$ Hz, C-22), 129.76 ($d, J = 7.6$ Hz, C-23, C-27), 115.66 ($d, J = 21.4$ Hz, C-24, C-26), 57.74 (C-5), 54.86 (C-9), 54.39 (C-14), 48.82 (C-13), 48.51 (C-15), 43.86 (C-4), 43.06 (C-21), 41.82 (C-7), 40.27 (C-1), 39.62 (C-8), 38.23 (C-3, C-10), 37.41 (C-12), 30.31 (C-20), 22.38 (C-6), 20.47

(C-11), 19.97 (C-17), 19.34 (C-2), 13.75 (C-19) ppm; ^{19}F NMR (470 MHz, CDCl_3): $\delta = -114.97$ (ddd, $J = 14.0, 8.8, 5.2$ Hz) ppm; MS (ESI, $\text{MeOH}/\text{CHCl}_3$, 4:1): m/z (%) = 424 (100%, $[\text{M}-\text{H}]^-$); analysis calcd for $\text{C}_{27}\text{H}_{36}\text{FNO}_2$ (425.59): C 76.20, H 8.53, N 3.29; found: C 75.87, H 8.76, N 3.03.

3.21. *N*-(2-Methylbenzyl)-16-oxostachan-18-amide (18)

Following GPA from **1** (465 mg, 1.46 mmol), 2-methyl-benzylamine (0.75 mL, 5.8 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **18** (425 mg, 69%) was obtained as a colorless solid; $R_f = 0.87$ (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1); m.p. = 75 °C; $[\alpha]_D^{20} = -57.17^\circ$ ($c = 0.16$, CHCl_3); IR (ATR): $\nu = 3391w, 2924m, 2847m, 1732m, 1644m, 1511m, 1454m, 1238w, 1189w, 1109w, 1005w, 976w, 740s, 695w, 665w, 589w, 506w, 456w, 430w$ cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 7.25 - 7.09$ (m, 4H, 23-H, 24-H, 25-H, 26-H), 5.67 (t, $J = 5.2$ Hz, 1H, N-H), 4.46 - 4.32 (m, 2H, 21-H), 2.63 (dd, $J = 18.6, 3.8$ Hz, 1H, 15-H), 2.33 (s, 3H, 28-H), 2.07 - 1.98 (m, 1H, 3-H), 1.98 - 1.90 (m, 1H, 6-H), 1.85 - 1.56 (m, 7H, 1-H, 2-H, 6-H, 7-H, 11-H, 12-H, 15-H), 1.54 (dd, $J = 11.6, 2.8$ Hz, 1H, 14-H), 1.50 - 1.32 (m, 4H, 2-H, 7-H, 12-H, 14-H), 1.21 (s, 3H, 20-H), 1.19 (s, 3H, 3-H, 9-H, 11-H), 1.13 (dd, $J = 12.1, 2.1$ Hz, 1H, 5-H), 0.97 (s, 3H, 17-H), 0.95 - 0.89 (m, 1H, 1-H), 0.76 (s, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): $\delta = 222.49$ (C-16), 176.46 (C-18), 136.58 (C-22), 136.19 (C-27), 130.73 (C-26), 128.97 (C-23), 127.93 (C-25), 126.38 (C-24), 57.81 (C-5), 54.91 (C-9), 54.43 (C-14), 48.85 (C-13), 48.53 (C-15), 43.93 (C-4), 42.01 (C-21), 41.86 (C-7), 40.33 (C-1), 39.64 (C-8), 38.28 (C-3), 38.22 (C-10), 37.45 (C-12), 30.39 (C-20), 22.39 (C-6), 20.49 (C-11), 20.00 (C-17), 19.39 (C-2), 19.20 (C-28), 13.73 (C-19) ppm; MS (ESI, $\text{MeOH}/\text{CHCl}_3$, 4:1): m/z (%) = 420 (100%, $[\text{M}-\text{H}]^-$); analysis calcd for $\text{C}_{28}\text{H}_{39}\text{NO}_2$ (421.63): C 79.76, H 9.32, N 3.32; found: C 79.51, H 9.48, N 3.09.

3.22. *N*-(3-Methylbenzyl)-16-oxostachan-18-amide (19)

Following GPA from **1** (418 mg, 1.31 mmol), 3-methyl-benzylamine (0.66 mL, 3.24 mmol) and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **19** (453 mg, 82%) was obtained as a colorless solid; $R_f = 0.76$ (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1); m.p. = 74.5 °C; $[\alpha]_D^{20} = -62.8^\circ$ ($c = 0.12$, CHCl_3); IR (ATR): $\nu = 3389w, 2923m, 2847m, 1733s, 1641s, 1609w, 1513s, 1453s, 1402w, 1374w, 1352w, 1316w, 1239m, 1187w, 1134w, 1109w, 1088w, 1028w, 1012w, 976w, 928w, 875w, 754m, 697m, 665w, 589w, 569w, 507w, 455w$ cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.26 - 6.98$ (m, 4H, 23-H, 24-H, 25-H, 27-H), 5.82 (t, $J = 5.5$ Hz, 1H, N-H), 4.41 - 4.31 (m, 2H, 21-H), 2.63 (dd, $J = 18.7, 3.8$ Hz, 1H, 15-H), 2.34 (s, 3H, 28-H), 2.07 - 1.99 (m, 1H, 6-H), 1.99 - 1.91 (m, 1H, 3-H), 1.87 - 1.57 (m, 7H, 1-H, 2-H, 6-H, 7-H, 11-H, 12-H, 15-H), 1.54 (dd, $J = 11.6, 2.7$ Hz, 1H, 14-H), 1.52 - 1.32 (m, 4H, 2-H, 7-H, 12-H, 14-H), 1.21 (s, 3H, 20-H), 1.31 - 1.13 (m, 3H, 3-H, 9-H, 11-H), 1.13 (dd, $J = 12.1, 2.3$ Hz, 1H, 5-H), 0.97 (s, 3H, 17-H), 0.95 - 0.87 (m, 1H, 1-H), 0.76 (s, 3H, 19-H) ppm; ^{13}C NMR (101 MHz, CDCl_3): $\delta = 222.49$ (C-16), 176.47 (C-18), 138.59 (C-22), 138.55 (C-26), 128.94 (C-24), 128.77 (C-27), 128.36 (C-25), 125.06 (C-23), 57.79 (C-5), 54.92 (C-9), 54.44 (C-14), 48.85 (C-13), 48.54 (C-15), 43.86 (C-21), 43.83 (C-4), 41.87 (C-7), 40.33 (C-1), 39.64 (C-10), 38.29 (C-3), 38.25 (C-8), 37.45 (C-12), 30.36 (C-20), 22.42 (C-6), 21.54 (C-28), 20.49 (C-11), 20.00 (C-17), 19.37 (C-2), 13.74 (C-19) ppm; MS (ESI, $\text{MeOH}/\text{CHCl}_3$, 4:1): m/z (%) = 420 (100%, $[\text{M}-\text{H}]^-$); analysis calcd for $\text{C}_{28}\text{H}_{39}\text{NO}_2$ (421.63): C 79.76, H 9.32, N 3.32; found: C 79.58, H 9.47, N 3.14.

3.23. *N*-(4-Methylbenzyl)-16-oxostachan-18-amide (20)

Following GPA from **1** (460 mg, 1.44 mmol), 4-methyl-benzylamine (0.74 mL, 5.8 mmol), and chromatography (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1), **20** (510 mg, 84%) was obtained as a colorless solid; $R_f = 0.8$ (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1); m.p. = 76 °C; $[\alpha]_D^{20} = -71.55^\circ$ ($c = 0.06$, CHCl_3); IR (ATR): $\nu = 3388w, 2923m, 2847m, 1734s, 1642s, 1514s, 1453s, 1318w, 1240m, 1182m, 1109w, 1008w, 976w, 814w, 749m, 695w, 586w, 506w, 473m$ cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 7.20 - 7.06$ (m, 4H, 23-H, 24-H, 26-H, 27-H), 5.80 (t, $J = 5.5$ Hz, 1H, N-H), 4.44 - 4.30 (m, 2H, 21-H), 2.64 (dd, $J = 18.6, 3.8$ Hz, 1H, 15-H), 2.34 (s, 3H, 28-H), 2.08 - 1.99 (m, 1H, 3-H), 1.99 - 1.90 (m, 1H, 6-H), 1.85 - 1.70 (m, 4H, 1-H, 2-H, 6-H, 15-H),

1.67 (dt, $J = 13.2, 3.4$ Hz, 2H, 7-H, 11-H), 1.63 – 1.57 (m, 1H, 12-H), 1.54 (dd, $J = 11.6, 2.8$ Hz, 1H, 14-H), 1.50 – 1.31 (m, 4H, 2-H, 7-H, 12-H, 14-H), 1.21 (s, 3H, 20-H), 1.29 – 1.13 (m, 3H, 3-H, 9-H, 11-H), 1.13 (dd, $J = 12.1, 2.1$ Hz, 1H, 5-H), 0.97 (s, 3H, 17-H), 0.92 (td, $J = 13.2, 4.5$ Hz, 1H, 1-H), 0.76 (s, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): $\delta = 222.53$ (C-16), 176.44 (C-18), 137.34 (C-22), 135.59 (C-25), 129.54 (C-24, C-26), 128.12 (C-23, C-27), 57.79 (C-5), 54.91 (C-9), 54.44 (C-14), 48.86 (C-13), 48.54 (C-15), 43.84 (C-4), 43.63 (C-21), 41.87 (C-7), 40.33 (C-1), 39.65 (C-8), 38.28 (C-3), 38.25 (C-10), 37.46 (C-12), 30.35 (C-20), 22.41 (C-6), 21.25 (C-28), 20.49 (C-11), 20.00 (C-17), 19.38 (C-2), 13.77 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 4:1): m/z (%) = 420 (100%, $[\text{M}-\text{H}]^-$); analysis calcd for $\text{C}_{28}\text{H}_{39}\text{NO}_2$ (421.63): C 79.63, H 9.32, N 3.32; found: C 79.42, H 9.50, N 3.06.

3.24. *N*-(2-Methoxybenzyl)-16-oxostachan-18-amide (21)

Following GPA from **1** (460 mg, 1.44 mmol), 2-methoxy-benzylamine (0.75 mL, 5.76 mmol), and chromatography (SiO_2 , hexanes/ethyl acetate, 95:5), **21** (529 mg, 84%) was obtained as a colorless solid; $R_f = 0.87$ (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1); m.p. = 56 °C; $[\alpha]_D^{20} = -64.66^\circ$ ($c = 0.115$, MeOH); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 270 nm (0.09); IR (ATR): $\nu = 3392w, 2924m, 2846m, 1734s, 1648m, 1602w, 1492s, 1457m, 1401w, 1370w, 1317w, 1289w, 1240s, 1171w, 1112m, 1043w, 1028m, 976w, 929w, 855w, 815w, 750s, 696w, 665w, 615w, 586w, 531w, 507w, 490w$ cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): $\delta = 7.30 - 7.19$ (m, 2H, 23-H, 25-H), 6.95 – 6.83 (m, 2H, 24-H, 26-H), 6.21 (t, $J = 5.7$ Hz, 1H, N-H), 4.40 – 4.33 (m, 2H, 21-H), 3.85 (s, 3H, 28-H), 2.54 (dd, $J = 18.6, 3.8$ Hz, 1H, 15-H), 2.08 – 1.98 (m, 1H, 3-H), 1.96 – 1.89 (m, 1H, 6-H), 1.81 – 1.59 (m, 6H, 1-H, 2-H, 6-H, 7-H, 11-H, 15-H), 1.59 – 1.54 (m, 1H, 12-H), 1.53 – 1.49 (m, 1H, 14-H), 1.49 – 1.29 (m, 4H, 2-H, 7-H, 12-H, 14-H), 1.16 (s, 3H, 20-H), 1.15 – 1.10 (m, 3H, 3-H, 9-H, 11-H), 1.09 – 1.05 (m, 1H, 5-H), 0.95 (s, 3H, 17-H), 0.88 (td, $J = 13.1, 4.4$ Hz, 1H, 1-H), 0.51 (s, 3H, 19-H) ppm; ^{13}C NMR (126 MHz, CDCl_3): $\delta = 222.51$ (C-16), 176.27 (C-18), 157.63 (C-27), 130.56 (C-25), 129.00 (C-23), 126.30 (C-22), 120.87 (C-24), 110.21 (C-26), 57.72 (C-5), 55.30 (C-28), 54.83 (C-9), 54.39 (C-14), 48.80 (C-13), 48.51 (C-15), 43.74 (C-4), 41.90 (C-7), 40.32 (C-1), 40.00 (C-21), 39.57 (C-8), 38.19 (C-3), 38.11 (C-10), 37.41 (C-12), 30.19 (C-20), 22.18 (C-6), 20.41 (C-11), 19.97 (C-17), 19.08 (C-2), 13.07 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 , 4:1): m/z (%) = 438 (90%, $[\text{M}+\text{H}]^+$), 897 (100%, $[\text{2M}+\text{Na}]^+$); analysis calcd for $\text{C}_{28}\text{H}_{39}\text{NO}_3$ (437.62): C 76.85, H 8.98, N 3.20; found: C 76.60, H 9.22, N 2.97.

3.25. *N*-(3-Methoxybenzyl)-16-oxostachan-18-amide (22)

Following GPA from **1** (500 mg, 1.57 mmol), 3-methoxy-benzylamine (0.82 mL, 6.28 mmol), and chromatography (SiO_2 , hexanes/ethyl acetate, 95:5), **22** (498 mg, 71%) was obtained as a colorless solid; $R_f = 0.89$ (SiO_2 , $\text{CHCl}_3/\text{MeOH}$, 9:1); m.p. = 58.5 °C; $[\alpha]_D^{20} = -56.57^\circ$ ($c = 0.069$, MeOH); UV-Vis (MeOH): λ_{max} ($\log \epsilon$) = 273 nm (0.17); IR (ATR): $\nu = 3390w, 2925m, 2846m, 1733s, 1650m, 1601m, 1586m, 1507s, 1489s, 1454s, 1352w, 1316w, 1262s, 1189w, 1151m, 1110w, 1087w, 1043m, 1008w, 976w, 874w, 854w, 776m, 738m, 694m, 569w, 554w, 507w, 465w$ cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.28 - 7.21$ (m, 1H, 24-H), 6.90 – 6.79 (m, 3H, 23-H, 25-H, 27-H), 5.85 (t, $J = 5.6$ Hz, 1H, N-H), 4.42 – 4.29 (m, 2H, 21-H), 3.79 (s, 3H, 28-H), 2.63 (dd, $J = 18.6, 3.8$ Hz, 1H, 15-H), 2.07 – 1.99 (m, 1H, 3-H), 1.99 – 1.91 (m, 1H, 6-H), 1.87 – 1.64 (m, 6H, 1-H, 2-H, 6-H, 7-H, 11-H, 15-H), 1.63 – 1.57 (m, 1H, 12-H), 1.54 (dd, $J = 11.6, 2.7$ Hz, 1H, 14-H), 1.51 – 1.31 (m, 4H, 2-H, 7-H, 12-H, 14-H), 1.22 (s, 3H, 20-H), 1.19 (s, 3H, 3-H, 9-H, 11-H), 1.13 (m, 1H, 5-H), 0.97 (s, 3H, 17-H), 0.94 – 0.88 (m, 1H, 1-H), 0.76 (s, 3H, 19-H) ppm; ^{13}C NMR (101 MHz, CDCl_3): $\delta = 222.48$ (C-16), 176.50 (C-18), 160.04 (C-26), 140.26 (C-22), 129.89 (C-24), 120.27 (C-23), 113.62 (C-25), 113.10 (C-27), 57.77 (C-5), 55.41 (C-28), 54.91 (C-9), 54.42 (C-14), 48.84 (C-13), 48.53 (C-15), 43.89 (C-4), 43.77 (C-21), 41.85 (C-7), 40.32 (C-1), 39.63 (C-8), 38.28 (C-3), 38.24 (C-10), 37.44 (C-12), 30.36 (C-20), 22.41 (C-6), 20.48 (C-11), 19.99 (C-17), 19.37 (C-2), 13.77 (C-19) ppm; MS (ESI, MeOH/ CHCl_3 4:1): m/z (%) = 436 (100%, $[\text{M}-\text{H}]^-$); analysis calcd for $\text{C}_{28}\text{H}_{39}\text{NO}_3$ (437.62): C 76.85, H 8.98, N 3.20; found: C 76.58, H 9.13, N 3.07.

3.26. *N*-(4-Methoxybenzyl)-16-oxostachan-18-amide (23)

Following GPA from **1** (500 mg, 1.57 mmol), 3-methoxy-benzylamine (0.61 mL, 4.71 mmol), and chromatography (SiO₂, hexanes/ethyl acetate, 95:5), **23** (618 mg, 90%) was obtained as a colorless solid; R_f = 0.9 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 59 °C; [α]_D²⁰ = −59.89° (c = 0.1, MeOH); UV-Vis (MeOH): λ_{max} (log ε) = 224 nm (0.66); IR (ATR): ν = 3396w, 2925m, 2846m, 1733m, 1642m, 1612w, 1511s, 1453m, 1356w, 1317w, 1301w, 1245s, 1179m, 1159w, 1109w, 1087w, 1032m, 976w, 929w, 828m, 751m, 696w, 665w, 588w, 563w, 508m, 415w cm^{−1}; ¹H NMR (400 MHz, CDCl₃): δ = 7.20 – 7.13 (m, 2H, 23-H, 27-H), 6.88 – 6.82 (m, 2H, 24-H, 26-H), 5.79 (t, J = 5.0 Hz, 1H, N-H), 4.40 – 4.26 (m, 2H, 21-H), 3.80 (s, 3H, 28-H), 2.63 (dd, J = 18.7, 3.8 Hz, 1H, 15-H), 2.05 – 1.98 (m, 1H, 3-H), 1.98 – 1.91 (m, 1H, 6-H), 1.85 – 1.64 (m, 6H, 1-H, 2-H, 6-H, 7-H, 11-H, 15-H), 1.63 – 1.57 (m, 1H, 12-H), 1.54 (dd, J = 11.6, 2.7 Hz, 1H, 14-H), 1.42 (m, 4H, 2-H, 7-H, 12-H, 14-H), 1.29 – 1.13 (m, 3H, 3-H, 9-H, 11-H), 1.20 (s, 3H, 20-H), 1.12 (m, 1H, 5-H), 0.97 (s, 3H, 17-H), 0.95 – 0.86 (m, 1H, 1-H), 0.75 (s, 3H, 19-H) ppm; ¹³C NMR (101 MHz, CDCl₃): δ = 222.53 (C-16), 176.41 (C-18), 159.16 (C-25), 130.71 (C-22), 129.44 (C-23, C-27), 114.24 (C-24, C-26), 57.78 (C-5), 55.44 (C-28), 54.89 (C-9), 54.42 (C-14), 48.85 (C-13), 48.53 (C-15), 43.82 (C-4), 43.31 (C-21), 41.85 (C-7), 40.3 (C-1), 39.64 (C-8), 38.24 (C-3, C-10), 37.45 (C-12), 30.33 (C-20), 22.40 (C-6), 20.48 (C-11), 19.99 (C-17), 19.36 (C-2), 13.76 (C-19) ppm; MS (ESI, MeOH/CHCl₃ 4:1): m/z (%) = 436 (100%, [M-H][−]); analysis calcd for C₂₈H₃₉NO₃ (437.62): C 76.85, H 8.98, N 3.20; found: C 76.63, H 9.17, N 3.01.

3.27. 18-Oxo-18-piperazin-1-yl-stachan-16-one (24)

Following GPA from **1** (2.0 g, 6.28 mmol), piperazine (3.1 g, 36 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **24** (2.28 g, 94%) was obtained as a colorless solid; R_f = 0.5 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 137 °C; [α]_D²⁰ = −26.12° (c = 0.117, CHCl₃); IR (ATR): ν = 2855w, 1726s, 1640s, 1453m, 1397m, 1328w, 1313w, 1252w, 1220w, 1177m, 1142w, 1108w, 1056w, 1030m, 978w, 796m, 590w, 556w, 519w, 505w cm^{−1}; ¹H NMR (500 MHz, CDCl₃): δ = 4.28 (s, 1H, N-H), 3.72 – 3.57 (m, 4H, 21-H, 24-H), 2.99 – 2.86 (m, 4H, 22-H, 23-H), 2.72 (dd, J = 18.7, 3.8 Hz, 1H, 15-H), 2.28 (dd, J = 14.2, 3.7 Hz, 1H, 3-H), 2.16 – 2.04 (m, 1H, 6-H), 1.83 – 1.75 (m, 2H, 6-H, 15-H), 1.71 – 1.62 (m, 3H, 1-H, 7-H, 11-H), 1.61 – 1.57 (m, 1H, 12-H), 1.55 (s, 1H, 2-H), 1.53 (dd, J = 11.6, 2.8 Hz, 1H, 14-H), 1.45 (dt, J = 6.4, 2.8 Hz, 1H, 2-H), 1.41 (dd, J = 11.3, 3.7 Hz, 1H, 7-H), 1.42 – 1.36 (m, 1H, 14-H), 1.34 (dd, J = 12.3, 5.2 Hz, 1H, 12-H), 1.28 (s, 3H, 20-H), 1.24 (td, J = 12.9, 4.9 Hz, 1H, 11-H), 1.20 – 1.11 (m, 2H, 3-H, 9-H), 1.01 (dd, J = 11.8, 1.8 Hz, 1H, 5-H), 0.96 (s, 3H, 17-H), 0.91 (td, J = 13.2, 4.3 Hz, 1H, 1-H), 0.82 (s, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.79 (C-16), 176.61 (C-18), 61.99 (C-5), 56.23 (C-9), 54.57 (C-14), 48.84 (C-13), 48.68 (C-15), 46.96 (C-22), 46.28 (C-4), 46.13 (C-21), 45.88 (C-25), 45.59 (C-24), 42.60 (C-7), 40.90 (C-1), 39.81 (C-3), 39.78 (C-8), 38.73 (C-10), 37.49 (C-12), 28.14 (C-20), 22.64 (C-6), 20.54 (C-11), 20.02 (C-17), 19.99 (C-2), 16.19 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 387 (100%, [M+H]⁺), 795 (30%, [2M+Na]⁺); analysis calcd for C₂₄H₃₈N₂O₂ (386.58): C 74.57, H 9.91, N 7.23; found: C 74.41, H 10.08, N 7.02.

3.28. 18-(1,4-Diazepan-1-yl)-18-oxostachan-16-one (25)

Following GPA from **1** (2.0 g, 6.28 mmol), homopiperazine (2.52 g, 36 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **25** (1.78 g, 71%) was obtained as a colorless solid; R_f = 0.4 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 162 °C; [α]_D²⁰ = −17.37° (c = 0.019, CHCl₃); IR (ATR): ν = 2922m, 1735s, 1628s, 1458m, 1401m, 1172m cm^{−1}; ¹H NMR (500 MHz, CDCl₃): δ = 5.29 (s, 1H, N-H), 3.74 – 3.57 (m, 4H, 21-H, 25-H), 3.14 – 2.86 (m, 4H, 22-H, 23-H), 2.76 – 2.64 (m, 1H, 15-H), 2.34 (td, J = 14.5, 3.5 Hz, 1H, 3-H), 2.18 – 2.08 (m, 1H, 6-H), 1.97 (ttt, J = 12.8, 8.9, 4.0 Hz, 2H, 24-H), 1.87 – 1.75 (m, 2H, 6-H, 15-H), 1.73 – 1.56 (m, 5H, 1-H, 2-H, 7-H, 11-H, 12-H), 1.53 (dd, J = 11.5, 2.7 Hz, 1H, 14-H), 1.50 – 1.32 (m, 4H, 2-H, 7-H, 11-H, 12-H), 1.29 (s, 3H, 20-H), 1.28 – 1.10 (m, 3H, 3-H, 9-H, 11-H), 1.01 (dd, J = 11.8, 1.9 Hz, 1H, 5-H), 0.96 (s, 3H, 17-H), 0.95 – 0.90 (m, 1H, 1-H), 0.83 (s, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.84 (C-16), 176.77 (C-18), 62.63 (C-5), 56.25 (C-9), 54.58

(C-14), 50.16 (C-26), 48.85 (C-13), 48.72 (C-22), 48.66 (C-15), 47.91 (C-21), 46.85 (C-4), 46.61 (C-23), 42.66 (C-7), 41.06 (C-1), 39.85 (C-3), 39.49 (C-8), 38.78 (C-10), 37.51 (C-12), 28.79 (C-25), 28.35 (C-20), 22.89 (C-6), 20.58 (C-11), 20.17 (C-17), 20.03 (C-2), 16.22 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 401 (100%, [M+H]⁺); analysis calcd for C₂₅H₄₀N₂O₂ (400.61): C 74.96, H 10.06, N 6.99; found: C 74.70, H 10.24, N 6.72.

3.29. 18-Morpholin-4-yl-18-oxostachan-16-one (26)

Following GPA from **1** (270 mg, 0.85 mmol), morpholine (0.44 mL, 5.1 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **26** (247 mg, 75%) was obtained as a colorless solid; R_f = 0.51 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 155 °C; $[\alpha]_D^{20}$ = −27.15° (c = 0.11, CHCl₃); IR (ATR): ν = 2922 m , 2850 m , 1733 s , 1644 m , 1452 m , 1389 m , 1373 m , 1264 m , 1224 m , 1174 m , 1111 s , 1068 w , 1027 m , 977 w , 928 w , 892 w , 844 w , 752 w , 694 w , 605 w , 508 w cm^{−1}; ¹H NMR (500 MHz, CDCl₃): δ = 3.75 – 3.48 (m , 8H, 21-H, 22-H, 23-H, 24-H), 2.73 (dd , J = 18.7, 3.8 Hz, 1H, 15-H), 2.29 (dt , J = 15.1, 3.5 Hz, 1H, 3-H), 2.10 (td , J = 13.9, 3.2 Hz, 1H, 6-H), 1.91 – 1.76 (m , 2H, 6-H, 15-H), 1.71 – 1.62 (m , 3H, 1-H, 7-H, 11-H), 1.62 – 1.56 (m , 2H, 2-H, 12-H), 1.53 (dd , J = 11.6, 2.8 Hz, 1H, 14-H), 1.49 – 1.41 (m , 2H, 2-H, 7-H), 1.41 – 1.32 (m , 2H, 12-H, 14-H), 1.28 (s , 3H, 20-H), 1.24 (s , 1H, 11-H), 1.21 – 1.15 (m , 2H, 3-H, 9-H), 1.01 (dd , J = 11.8, 1.9 Hz, 1H, 5-H), 0.96 (s , 3H, 17-H), 0.94 – 0.88 (m , 1H, 1-H), 0.84 (s , 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.80 (C-16), 176.63 (C-18), 67.11 (C-22, C-23), 61.96 (C-5), 56.23 (C-9), 54.59 (C-14), 48.85 (C-13), 48.68 (C-15), 46.84 (C-21, C-24), 46.27 (C-4), 42.61 (C-7), 40.93 (C-1), 39.82 (C-8), 39.76 (C-3), 38.73 (C-10), 37.50 (C-12), 28.15 (C-20), 22.67 (C-6), 20.56 (C-11), 20.02 (C-17), 19.98 (C-2), 16.17 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 388 (100%, [M+H]⁺); analysis calcd for C₂₄H₃₇NO₃ (387.56): C 74.77, H 9.79, N 3.49; found: C 74.50, H 9.97, N 3.28.

3.30. 18-(1,4-Oxazepan-4-yl)-18-oxostachan-16-one (27)

Following GPA from **1** (250 mg, 0.785 mmol), homomorpholine (166 mg, 1.21 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **27** (195 mg, 62%) was obtained as a colorless solid; R_f = 0.65 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 229 °C; $[\alpha]_D^{20}$ = −11.26° (c = 0.091, CHCl₃); IR (ATR): ν = 2922 m , 2845 m , 1736 s , 1626 s , 1459 m , 1404 w , 1363 w , 1252 w , 1219 w , 1183 w , 1123 s , 1013 w , 976 w , 963 w , 931 w , 849 w , 751 w , 696 w , 574 w , 507 w cm^{−1}; ¹H NMR (500 MHz, CDCl₃): δ = 3.85 – 3.56 (m , 8H, 21-H, 22-H, 23-H, 25-H), 2.71 (dd , J = 18.7, 3.8 Hz, 1H, 15-H), 2.49 – 2.41 (m , 1H, 3-H), 2.34 (ddt , J = 14.4, 4.1, 2.1 Hz, 1H, 6-H), 2.23 – 2.10 (m , 1H, 24-H), 1.99 – 1.88 (m , 1H, 24-H), 1.87 – 1.81 (m , 1H, 6-H), 1.78 (d , J = 18.8 Hz, 1H, 15-H), 1.73 – 1.56 (m , 5H, 1-H, 2-H, 7-H, 11-H, 12-H), 1.53 (dd , J = 11.7, 2.6 Hz, 1H, 14-H), 1.50 – 1.42 (m , 1H, 2-H), 1.42 – 1.31 (m , 3H, 7-H, 12-H, 14-H), 1.29 (s , 3H, 20-H), 1.26 (s , 1H, 11-H), 1.21 – 1.09 (m , 2H, 3-H, 9-H), 1.01 (dd , J = 11.8, 1.9 Hz, 1H, 5-H), 0.96 (s , 3H, 17-H), 0.94 (s , 1H, 1-H), 0.84 (s , 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.87 (C-16), 176.64 (C-18), 71.05 (C-23), 69.63 (C-22), 62.68 (C-5), 56.26 (C-9), 54.61 (C-14), 51.89 (C-25), 48.85 (C-13), 48.67 (C-15), 47.80 (C-21), 46.85 (C-4), 42.69 (C-7), 41.14 (C-1), 39.86 (C-8), 39.59 (C-3), 38.80 (C-10), 37.53 (C-12), 30.51 (C-24), 28.39 (C-20), 22.93 (C-6), 20.59 (C-11), 20.15 (C-2), 20.04 (C-17), 19.90, 16.22 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): m/z (%) = 402 (85%, [M+H]⁺), 425 (80%, [M+Na]⁺), 826 (100%, [2M+H]⁺); analysis calcd for C₂₅H₃₉NO₃ (401.59): C 74.77, H 9.79, N 3.49; found: C 74.47, H 9.97, N 3.29.

3.31. 18-Oxo-18-thiomorpholin-4-yl-stachan-16-one (28)

Following GPA from **1** (270 mg, 0.85 mmol), thiomorpholine (0.52 mL, 5.1 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **28** (198 mg, 58%) was obtained as a colorless solid; R_f = 0.86 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 137 °C; $[\alpha]_D^{20}$ = −31.96° (c = 0.148, CHCl₃); IR (ATR): ν = 2921 m , 2844 m , 1736 s , 1639 s , 1459 m , 1393 m , 1359 m , 1274 w , 1243 w , 1215 w , 1160 s , 1112 w , 1025 w , 959 s , 848 w , 758 w , 695 w , 604 w , 496 w cm^{−1}; ¹H NMR (500 MHz, CDCl₃): δ = 3.89 – 3.75 (m , 4H, 21-H, 24-H), 2.72 (dd , J = 18.7, 3.8 Hz, 1H, 15-H), 2.68 – 2.50 (m , 4H, 22-H, 23-H), 2.28 (dt , J = 14.0, 3.0 Hz, 1H, 3-H), 2.19 – 2.06 (m , 1H, 6-H), 1.79 (d , J = 18.8 Hz, 1H, 15-H), 1.78 (s , 1H, 6-H), 1.71 (dt , J = 18.8, 2.3 Hz, 1H, 1-H), 1.68 – 1.63 (m ,

2H, 7-H, 11-H), 1.63 – 1.56 (*m*, 1H, 12-H), 1.53 (*dd*, *J* = 11.5, 2.8 Hz, 1H, 14-H), 1.54 – 1.44 (*m*, 1H, 2-H), 1.47 – 1.38 (*m*, 2H, 7-H, 14-H), 1.37 – 1.32 (*m*, 1H, 12-H), 1.29 (*s*, 3H, 20-H), 1.28 – 1.23 (*m*, 1H, 11-H), 1.17 (*dd*, *J* = 11.5, 3.5 Hz, 2H, 9-H), 1.13 (*d*, *J* = 3.2 Hz, 1H, 3-H), 1.00 (*dd*, *J* = 11.7, 1.6 Hz, 1H, 5-H), 0.96 (*s*, 3H, 17-H), 0.92 (*td*, *J* = 13.1, 4.3 Hz, 1H, 1-H), 0.82 (*s*, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.84 (C-16), 176.56 (C-18), 62.37 (C-5), 56.31 (C-9), 54.60 (C-14), 48.92 (C-21, C-24), 48.86 (C-13), 48.68 (C-15), 46.46 (C-4), 42.68 (C-7), 41.02 (C-1), 39.85 (C-8), 39.63 (C-3), 38.76 (C-10), 37.51 (C-12), 28.23 (C-20), 27.67 (C-22, C-23), 22.71 (C-6), 20.56 (C-11), 20.07 (C-2), 20.03 (C-17), 16.31 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) = 404 (100%, [M+H]⁺); analysis calcd for C₂₄H₃₇NSO₂ (403.63): C 71.42, H 9.24, N 3.47; found: C 71.25, H 9.40, N 3.28.

3.32. 18-Oxo-18-(1,4-thiazepan-4-yl)-stachan-16-one (29)

Following GPA from **1** (270 mg, 0.85 mmol), homothiomorpholine (194 mg, 1.27 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **29** (245 mg, 69%) was obtained as a colorless solid; R_f = 0.5 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 198 °C; [α]_D²⁰ = −18.43° (*c* = 0.1, CHCl₃); IR (ATR): ν = 2920*m*, 2843*m*, 1737*s*, 1624*s*, 1462*m*, 1401*m*, 1358*m*, 1274*w*, 1162*m*, 1111*w*, 975*w*, 881*m*, 753*w*, 489*w* cm^{−1}; ¹H NMR (500 MHz, CDCl₃): δ = 4.00 (*ddd*, *J* = 14.3, 5.5, 4.1 Hz, 1H, 25-H), 3.81 (*dt*, *J* = 14.4, 5.3 Hz, 1H, 21-H), 3.65 – 3.52 (*m*, 2H, 21-H, 25-H), 2.86 – 2.67 (*m*, 4H, 15-H, 22-H, 24-H), 2.61 (*ddd*, *J* = 14.7, 9.4, 5.1 Hz, 1H, 22-H), 2.32 (*dt*, *J* = 12.8, 2.7 Hz, 1H, 3-H), 2.22 – 2.12 (*m*, 1H, 6-H), 2.11 – 2.04 (*m*, 1H, 23-H), 2.00 – 1.90 (*m*, 1H, 23-H), 1.84 (*dt*, *J* = 13.7, 2.9 Hz, 1H, 6-H), 1.79 (*d*, *J* = 18.7 Hz, 1H, 15-H), 1.76 – 1.69 (*m*, 1H, 1-H), 1.69 – 1.55 (*m*, 4H, 2-H, 7-H, 11-H, 12-H), 1.53 (*dd*, *J* = 11.6, 2.8 Hz, 1H, 14-H), 1.50 – 1.46 (*m*, 1H, 2-H), 1.45–1.34 (*m*, 2H, 7-H, 14-H), 1.36 – 1.21 (*m*, 2H, 11-H, 12-H), 1.29 (*s*, 3H, 20-H), 1.21 – 1.13 (*m*, 2H, 3-H, 9-H), 1.00 (*dd*, *J* = 11.8, 1.9 Hz, 1H, 5-H), 0.96 (*s*, 3H, 17-H), 0.93 (*d*, *J* = 4.2 Hz, 1H, 1-H), 0.86 – 0.83 (*m*, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.91 (C-16), 176.43 (C-18), 62.82 (C-5), 56.25 (C-9), 54.61 (C-14), 53.95 (C-25), 49.50 (C-21), 48.88 (C-15), 48.68 (C-13), 47.01 (C-4), 42.73 (C-7), 41.16 (C-1), 39.89 (C-8), 39.27 (C-3), 38.82 (C-10), 37.55 (C-12), 34.32 (C-24), 31.88 (C-22), 30.37 (C-23), 28.55 (C-20), 22.98 (C-6), 20.62 (C-11), 20.45 (C-2), 20.06 (C-17), 16.23 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) = 419 (100%, [M+H]⁺), 859 (67%, [2M+Na]⁺); analysis calcd for C₂₅H₃₉NSO₂ (417.65): C 71.90, H 9.41, N 3.35; found: C 71.70, H 9.65, N 3.16.

3.33. N-(2-Aminoethyl)-16-oxostachan-18-amide (30)

Following GPA from **1** (270 mg, 0.85 mmol), ethylenediamine (0.34 mL, 5.1 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **30** (270 mg, 88%) was obtained as a colorless solid; R_f = 0.25 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 105 °C; [α]_D²⁰ = −71.24° (*c* = 0.121, CHCl₃); IR (ATR): ν = 2924*s*, 1735*s*, 1637*s*, 1516*s*, 1453*s* cm^{−1}; ¹H NMR (500 MHz, CDCl₃): δ = 8.21 – 8.16 (*m*, 1H, NH), 6.75 – 6.70 (*m*, 2H, NH₂), 3.57 – 3.27 (*m*, 2H, 22-H), 3.23 (*s*, 2H, 21-H), 2.66 – 2.55 (*m*, 1H, 15-H), 2.18 – 2.09 (*m*, 1H, 3-H), 2.05 (*d*, *J* = 16.2 Hz, 1H, 6-H), 1.80 – 1.64 (*m*, 5H, 1-H, 2-H, 7-H, 11-H, 12-H), 1.46 – 1.45 (*dd*, 1H, 14-H), 1.36 – 1.22 (*m*, 4H, 2-H, 7-H, 11-H, 12-H), 1.17 (*s*, 3H, 20-H), 1.15 (*m*, 1H, 9-H, 11-H), 1.12 – 1.06 (*m*, 1H, 5-H), 0.96 (*d*, *J* = 1.4 Hz, 3H, 17-H), 0.91 – 0.82 (*m*, 1H, 1-H), 0.80 (*s*, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.57 (C-16), 178.63 (C-18), 139.92 (C-22), 106.67 (C-23), 57.48 (C-5), 55.57 (C-9), 54.76 (C-14), 50.06 (C-13), 48.70 (C-15), 48.47 (C-4), 43.84 (C-7), 40.16 (C-1), 40.09 (C-3), 39.51 (C-8), 38.17 (C-10), 37.29 (C-12), 29.96 (C-20), 22.12 (C-6), 20.33 (C-11), 19.83 (C-17), 19.13 (C-2), 13.54 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) = 359 (100%, [M-H][−]); analysis calcd for C₂₂H₃₆N₂O₂ (360.54): C 73.29, H 10.06, N 7.77; found: C 72.96, H 10.30, N 7.49.

3.34. 18-(4-{2-[3,6-Bis(diethylamino)xanthenium-9-yl]-benzoyl}-piperazin-1-yl)-18-oxostachan-16-one Chloride (31)

Following GPC from **24** (400 mg, 1.03 mmol), oxalyl chloride (0.35 mL, 4.14 mmol), rhodamine B (744 mg, 1.55 mmol), NEt₃ (0.16 mL, 1.14 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **31** (410 mg, 47%) was obtained as a purple solid; R_f = 0.3 (SiO₂,

CHCl₃/MeOH, 9:1); m.p. = 193 °C; UV-Vis (MeOH): λ_{\max} (log ϵ) = 561.85 nm (1.74); IR (ATR): ν = 2924w, 1733w, 1632w, 1585s, 1528w, 1451m, 1411m, 1335s, 1272m, 1247m, 1179s, 1132m, 1072m, 1003m, 976w, 922w, 824w, 747m, 683m, 498w cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 7.75 – 7.27 (*m*, 6H, 28-H, 29-H, 30-H, 31-H, 35-H, 38-H), 7.12 – 6.98 (*m*, 2H, 37-H), 6.78 (*s*, 2H, 35-H), 3.88 – 3.14 (*m*, 16H, 21-H, 22-H, 23-H, 24-H, 39-H), 2.66 (*dd*, *J* = 18.6, 3.7 Hz, 1H, 15-H), 2.29 – 2.22 (*m*, 1H, 3-H), 2.07 – 1.96 (*m*, 1H, 6-H), 1.80 – 1.72 (*m*, 2H, 6-H, 15-H), 1.70 – 1.54 (*m*, 4H, 1-H, 7-H, 11-H, 12-H), 1.51 (*dd*, *J* = 11.7, 2.6 Hz, 1H, 14-H), 1.47 – 1.08 (*m*, 11H, 2-H, 3-H, 7-H, 9-H, 11-H, 12-H, 14-H, 20-H), 0.98 (*d*, *J* = 11.8 Hz, 1H, 5-H), 0.96 – 0.86 (*m*, 15H, 17-H, 40-H), 0.84 – 0.81 (*m*, 1H, 1-H), 0.75 (*s*, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.87 (C-16), 177.31 (C-18), 167.85 (C-25), 157.86 (C-36), 155.94 (C-32), 155.77 (C-34), 135.24 (C-27), 132.58 (C-26), 132.39 (C-28), 130.98 (C-38), 130.84 (C-33), 130.44 (C-29), 130.38 (C-31), 130.28 (C-30), 114.55 (C-37), 96.50 (C-35), 61.81 (C-5), 56.02 (C-9), 54.47 (C-14), 48.83 (C-13), 48.66 (C-15), 46.36 (C-21, C-24), 46.33 (C-22, C-23), 46.28 (C-4), 42.48 (C-7), 42.16 (C-39), 40.78 (C-1), 39.78 (C-8), 39.51 (C-3), 38.65 (C-10), 37.44 (C-12), 28.09 (C-20), 22.54 (C-6), 20.50 (C-11), 19.98 (C-17), 19.88 (C-2), 12.82 (C-40), 11.07 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) = 812 (78%, [M-Cl]⁺); analysis calcd for C₅₂H₆₇N₄O₄Cl (847.58): C 73.69, H 7.97, N 6.61; found: C 73.40, H 8.16, N 6.45.

3.35. 18-(4-{2-[3,6-Bis(diethylamino)xanthenium-9-yl]-benzoyl}-1,4-diazepan-1-yl)-18-oxostachan-16-one Chloride (32)

Following GPC from **25** (200 mg, 0.49 mmol), oxalyl chloride (0.17 mL, 2.0 mmol), rhodamine B (359.3 mg, 0.75 mmol), NEt₃ (0.1 mL, 0.05 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **32** (207 mg, 49%) was obtained as a purple solid; R_f = 0.2 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 185 °C; UV-Vis (MeOH): λ_{\max} (log ϵ) = 561.92 nm (2.55); IR (ATR): ν = 2925w, 1734w, 1586s, 1528w, 1466m, 1412w, 1336s, 1274m, 1246m, 1179s, 1132m, 1073m, 1011w, 976w, 920w, 822w, 771m, 682m, 619w, 577w, 497w cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ = 7.68 – 7.22 (*m*, 6H, 29-H, 30-H, 31-H, 32-H, 39-H), 6.98 – 6.74 (*m*, 4H, 38-H, 39-H), 3.98 – 3.11 (*m*, 16H, 21-H, 22-H, 23-H, 25-H, 40-H), 2.76 – 2.64 (*m*, 1H, 15-H), 2.31 (*d*, *J* = 14.4 Hz, 1H, 3-H), 2.19 – 1.97 (*m*, 1H, 6-H), 1.91 – 1.73 (*m*, 4H, 6-H, 15-H, 24-H), 1.72 – 1.54 (*m*, 4H, 1-H, 7-H, 11-H, 12-H), 1.55 – 1.05 (*m*, 24H, 2-H, 3-H, 7-H, 9-H, 11-H, 12-H, 14-H, 20-H, 41-H), 1.06 – 0.74 (*m*, 8H, 1-H, 5-H, 17-H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.94 (C-16), 176.96 (C-18), 168.99 (C-26), 168.49 (C-37), 157.93 (C-33), 155.78 (C-35), 140.56 (C-39), 136.52 (C-28), 136.30 (C-27), 132.65 (C-29), 132.43 (C-30), 129.57 (C-32), 126.87 (C-31), 114.09 (C-38), 113.73 (C-34), 96.90 (C-36), 62.25 (C-5), 56.21 (C-9), 54.56 (C-14), 50.49 (C-25), 49.83 (C-23), 48.86 (C-13), 48.65 (C-15), 46.77 (C-40), 46.55 (C-21), 46.40 (C-22), 44.07 (C-4), 42.60 (C-7), 40.94 (C-1), 39.82 (C-8), 39.43 (C-10), 39.27 (C-3), 37.50 (C-12), 29.31 (C-24), 27.87 (C-20), 22.77 (C-6), 20.56 (C-11), 20.27 (C-2), 20.02 (C-17), 15.36 (C-19), 12.87 (C-41) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) = 825 (92%, [M-Cl]⁺); analysis calcd for C₅₃H₆₉N₄O₄Cl (861.61): C 73.88, H 8.07, N 6.50; found: C 73.68, H 8.21, N 6.36.

3.36. 18-(4-[3-(2,3,6,7,12,13,16,17-Octahydro-1H,5H,11H,15H-pyrido[3,2,1-*ij*]-pyrido[1'',2'',3'':1',8']-quinolino[6',5':5,6]-pyrano[2,3-*fl*]-quinoline-4-ium-9-yl)-benzoyl]-piperazin-1-yl)-18-oxostachan-16-one Chloride (33)

Following GPC from **24** (500 mg, 1.29 mmol), oxalyl chloride (0.6 mL, 6.78 mmol), rhodamine 101 (423.2 mg, 0.86 mmol), NEt₃ (0.5 mL, 3.39 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **33** (416 mg, 36%) was obtained as a purple solid; R_f = 0.26 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 197 °C; UV-Vis (MeOH): λ_{\max} (log ϵ) = 582.71 nm (1.14); IR (ATR): ν = 2845w, 1732w, 1633w, 1594m, 1542w, 1493m, 1458w, 1360w, 1293s, 1180m, 1087s, 1024m, 1002m, 747w, 621m, 420m cm⁻¹; ¹H NMR (500 MHz, DMSO-d₆): δ = 7.78 – 7.53 (*m*, 6H, 28-H, 29-H, 30-H, 31-H, 35-H, 38-H), 7.40 (*dd*, *J* = 5.8, 2.7 Hz, 2H, 37-H), 6.66 (*d*, *J* = 2.8 Hz, 2H, 35-H), 3.50 (*dt*, *J* = 17.1, 5.6 Hz, 16H, 21-H, 22-H, 23-H, 24-H, 39-H), 3.04 – 2.91 (*m*, 10H, 40-H, 41-H), 2.66 (*t*, *J* = 6.5 Hz, 1H, 15-H), 2.26 – 2.15 (*m*, 1H, 3-H), 2.05 – 1.92 (*m*, 1H, 6-H), 1.85 (*q*, *J* = 9.3, 5.2 Hz, 2H, 6-H, 15-H), 1.76 – 1.54 (*m*, 4H,

1-H, 7-H, 11-H, 12-H), 1.54 – 1.29 (*m*, 12H, 2-H, 3-H, 7-H, 9-H, 11-H, 12-H, 14-H, 20-H), 1.23 (*d*, *J* = 2.9 Hz, 1H, 5-H), 1.18 (*s*, 3H, 20-H), 1.12–0.95 (*m*, 3H, 17-H), 0.87 (*s*, 1H, 1-H), 0.69 (*s*, 3H, 19-H); ¹³C NMR (126 MHz, CDCl₃): δ = 222.78 (C-16), 177.14 (C-18), 167.87 (C-25), 153.17 (C-36), 152.01 (C-32), 151.30 (C-34), 134.84 (C-27), 131.75 (C-26), 130.16 (C-28), 127.67 (C-38), 126.75 (C-33), 123.64 (C-30), 113.67 (C-37), 105.35 (C-35), 61.66 (C-5), 55.87 (C-9), 54.37 (C-14), 51.15 (C-13), 50.63 (C-15), 48.70 (C-21, C-24), 48.55 (C-22, C-23), 46.18 (C-4), 42.35 (C-39), 40.68 (C-7), 39.66 (C-1), 38.55 (C-8), 37.33 (C-3), 29.66 (C-10), 28.01 (C-12), 27.75 (C-20), 22.47 (C-6), 20.72 (C-41), 20.39 (C-40), 20.00 (C-11), 19.86 (C-17), 19.74 (C-2), 16.02 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) = 860 (90%, [M-Cl]⁺); analysis calcd for C₅₆H₆₇N₄O₄Cl (895.63): C 75.10, H 7.54, N 6.26; found: C 74.88, H 7.78, N 6.17.

3.37. 18-(4-[3-(2,3,6,7,12,13,16,17-Octahydro-1H,5H,11H,15H-pyrido[3,2,1-*ij*]-pyrido[1'',2'',3'':1',8']-quinolino[6',5':5,6]-pyranol[2,3-*fl*]-quinoline-4-ium-9-yl)-benzoyl]-1,4-diazepan-1-yl)-18-oxostachan-16-one chloride (34)

Following GPZ from **25** (636 mg, 1.58 mmol), oxalyl chloride (0.6 mL, 6.78 mmol), rhodamine 101 (408 mg, 0.83 mmol), NEt₃ (0.5 mL, 3.39 mmol), and chromatography (SiO₂, CHCl₃/MeOH, 95:5), **34** (532 mg, 37%) was obtained as a purple solid; R_f = 0.23 (SiO₂, CHCl₃/MeOH, 9:1); m.p. = 189 °C; UV-Vis (MeOH): λ_{max} (log ε) = 583.47 nm (1.79); IR (ATR): ν = 2924s, 1732w, 1594m, 1543w, 1493m, 1458w, 1361w, 1293s, 1181m, 1088s, 1025m, 746m, 621m, 421m cm⁻¹; ¹H NMR (500 MHz, DMSO-d₆): δ = 7.75 – 7.61 (*m*, 6H, 29-H, 30-H, 31-H, 32-H, 39-H), 6.73 – 6.60 (*m*, 4H, 38-H, 39-H), 3.65 – 3.38 (*m*, 16H, 21-H, 22-H, 23-H, 24-H, 40-H), 3.26 (*d*, *J* = 21.3 Hz, 4H, 41-H, 42-H), 3.01 – 2.83 (*m*, 4H, 41-H, 42-H), 2.74 – 2.57 (*m*, 1H, 15-H), 2.48 – 2.40 (*m*, 1H, 6-H), 2.21 (*m*, 1H, 3-H), 1.90 – 1.76 (*m*, 4H, 6-H, 15-H, 24-H), 1.61 (*m*, 4H, 1-H, 7-H, 11-H, 12-H), 1.35 (*ddd*, *J* = 21.8, 12.8, 11.1, 6.9 Hz, 3H, 11-H, 12-H, 14-H), 1.25 – 1.18 (*m*, 3H, 2-H, 3-H, 7-H), 1.18 – 0.96 (*m*, 3H, 5-H, 7-H, 9-H), 0.94 (*d*, *J* = 12.8 Hz, 3H, 20-H), 0.87 (*d*, *J* = 5.5 Hz, 3H, 17-H), 0.75 (*s*, 1H, 1-H), 0.66 (*s*, 3H, 19-H) ppm; ¹³C NMR (126 MHz, CDCl₃): δ = 222.81 (C-16), 176.39 (C-18), 168.42 (C-26), 163.05 (C-37), 152.01 (C-33), 152.94 (C-35), 151.35 (C-39), 131.33 (C-28), 130.69 (C-27), 130.15 (C-29), 129.76 (C-30), 129.21 (C-32), 127.75 (C-31), 123.84 (C-38), 123.43 (C-34), 113.27 (C-36), 62.49 (C-5), 62.09 (C-9), 56.07 (C-14), 54.44 (C-41), 50.94 (C-25), 49.97 (C-23), 48.72 (C-13), 48.67 (C-15), 47.14 (C-40), 46.73 (C-21), 46.58 (C-22), 46.37 (C-42), 44.92 (C-4), 42.57 (C-7), 40.98 (C-1), 40.81 (C-8), 39.71 (C-10), 38.64 (C-3), 37.34 (C-12), 28.51 (C-24), 27.54 (C-20), 22.64 (C-6), 20.61 (C-11), 19.90 (C-2), 19.67 (C-17), 16.27 (C-19) ppm; MS (ESI, MeOH/CHCl₃, 4:1): *m/z* (%) 874 (96%, [M-Cl]⁺); analysis calcd for C₅₇H₆₉N₄O₄Cl (909.64): C 75.26, H 7.65, N 6.16; found: C 75.01, H 7.83, N 5.97.

4. Conclusions

Acid hydrolysis of stevioside resulted in a 63% yield of isosteviol (1), which was used as a starting material for the preparation of numerous amides. These amides were tested for cytotoxic activity; while almost all the amides were found to be non-cytotoxic in a panel of human tumor cell lines, only the combination of isosteviol, a (homo)piperazinyl spacer and rhodamine B or rhodamine 101 unit proved to be particularly suitable. These spaced rhodamine conjugates showed cytotoxic activity in the sub-micromolar concentration range. In this regard, the homopiperazinyl-spacer derivatives were found to be better than the piperazinyl-spacer compounds, and the rhodamine 101 conjugates were more cytotoxic than the rhodamine B.

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References

1. Gasmalla, M.A.A.; Yang, R.; Hua, X. Stevia rebaudiana Bertoni: An alternative Sugar Replacer and Its Application in Food Industry. *Food Eng. Rev.* **2014**, *6*, 150–162. [[CrossRef](#)]
2. Geuns, J.M.C. Safety of Stevia and stevioside. *Recent Res. Dev. Phytochem.* **2000**, *4*, 75–88.
3. Geuns, J.M.C. Stevioside. *Phytochemistry* **2003**, *64*, 913–921. [[CrossRef](#)]
4. Kinghorn, A.D.; Wu, C.D.; Soejarto, D.D. Stevioside. *Food Sci. Technol.* **2001**, *112*, 167–183.
5. Lemus-Mondaca, R.; Vega-Galvez, A.; Zura-Bravo, L.; Kong, A.-H. Stevia rebaudiana Bertoni, source of a high-potency natural sweetener: A comprehensive review on the biochemical, nutritional and functional aspects. *Food Chem.* **2012**, *132*, 1121–1132. [[CrossRef](#)] [[PubMed](#)]
6. Chatsudthipong, V.; Muanprasat, C. Stevioside and related compounds: Therapeutic benefits beyond sweetness. *Pharmacol. Ther.* **2009**, *121*, 41–54. [[CrossRef](#)] [[PubMed](#)]
7. Ferrazzano, G.F.; Cantile, T.; Alciadi, B.; Coda, M.; Ingenito, A.; Zarrelli, A.; Di Fabio, G.; Pollio, A. Is Stevia rebaudiana bertoni a non cariogenic sweetener? A review. *Molecules* **2016**, *21*, 38. [[CrossRef](#)]
8. Heerranz-Lopez, M.; Barrajon-Catalan, E.; Beltran-Debon, R.; Joven, J.; Micol, V. Stevia is a source for alternative sweeteners: Potential medicinal effects. *Agro Food Ind. Hi-Tech* **2010**, *21*, 38–42.
9. Iatridis, N.; Kougioumtzi, A.; Vlataki, K.; Papadaki, S.; Magklara, A. Anti-Cancer Properties of Stevia rebaudiana; More than a Sweetener. *Molecules* **2022**, *27*, 1362. [[CrossRef](#)]
10. Momtazi-Borojeni, A.A.; Esmaeili, S.-A.; Abdollahi, E.; Sahebkar, A. A Review on the Pharmacology and Toxicology of Steviol Glycosides Extracted from Stevia rebaudiana. *Curr. Pharm. Des.* **2017**, *23*, 1616–1622. [[CrossRef](#)]
11. Wang, Y.; Luo, X.; Chen, L.; Mustapha, A.T.; Yu, X.; Zhou, C.; Okonkwo, C.E. Natural and low-caloric rebaudioside A as a substitute for dietary sugars: A comprehensive review. *Compr. Rev. Food Sci. Food Saf.* **2023**, *22*, 615–642. [[CrossRef](#)] [[PubMed](#)]
12. Renwick, A.G.; Tarka, S.M. Microbial hydrolysis of steviol glycosides. *Food Chem. Toxicol.* **2008**, *46*, S70. [[CrossRef](#)] [[PubMed](#)]
13. Wang, M.; Li, H.; Xu, F.; Gao, X.; Li, J.; Xu, S.; Zhang, D.; Wu, X.; Xu, J.; Hua, H.; et al. Diterpenoid lead stevioside and its hydrolysis products steviol and isosteviol: Biological activity and structural modification. *Eur. J. Med. Chem.* **2018**, *156*, 885–906. [[CrossRef](#)] [[PubMed](#)]
14. Lohoeelter, C.; Weckbecker, M.; Waldvogel, S.R. (–)-Isosteviol as a Versatile Ex-Chiral-Pool Building Block for Organic Chemistry. *Eur. J. Org. Chem.* **2013**, *2013*, 5539–5554. [[CrossRef](#)]
15. Moons, N.; De Borggraeve, W.; Dehaen, W. Isosteviol as a starting material in organic synthesis. *Curr. Org. Chem.* **2011**, *15*, 2731–2741. [[CrossRef](#)]
16. Moons, N.; De Borggraeve, W.; Dehaen, W. Stevioside and steviol as starting materials in organic synthesis. *Curr. Org. Chem.* **2012**, *16*, 1986–1995. [[CrossRef](#)]
17. Ullah, A.; Munir, S.; Mabkhot, Y.; Badshah, S.L. Bioactivity profile of the diterpene isosteviol and its derivatives. *Molecules* **2019**, *24*, 678. [[CrossRef](#)]
18. Mizushina, Y.; Akihisa, T.; Ukiya, M.; Hamasaki, Y.; Murakami-Nakai, C.; Kuriyama, I.; Takeuchi, T.; Sugawara, F.; Yoshida, H. Structural analysis of isosteviol and related compounds as DNA polymerase and DNA topoisomerase inhibitors. *Life Sci.* **2005**, *77*, 2127–2140. [[CrossRef](#)]
19. Malki, A.; El-Sharkawy, A.; Mohamed, M.; Bergmeier, S. Antitumor activities of the novel isosteviol derivative 10C against liver cancer. *Anticancer. Res.* **2017**, *37*, 1591–1601. [[CrossRef](#)]
20. Zhang, H.; Zhong, K.; Lu, M.; Mei, Y.; Tan, E.; Sun, X.; Tan, W. Neuroprotective effects of isosteviol sodium through increasing CYLD by the downregulation of miRNA-181b. *Brain Res. Bull.* **2018**, *140*, 392–401. [[CrossRef](#)]
21. Andreeva, O.V.; Garifullin, B.F.; Sharipova, R.R.; Strobykina, I.Y.; Sapunova, A.S.; Voloshina, A.D.; Belenok, M.G.; Dobrynin, A.B.; Khabibulina, L.R.; Kataev, V.E. Glycosides and Glycoconjugates of the Diterpenoid Isosteviol with a 1,2,3-Triazolyl Moiety: Synthesis and Cytotoxicity Evaluation. *J. Nat. Prod.* **2020**, *83*, 2367–2380. [[CrossRef](#)] [[PubMed](#)]
22. Garifullin, B.F.; Strobykina, I.Y.; Khabibulina, L.R.; Sapunova, A.S.; Voloshina, A.D.; Sharipova, R.R.; Khairutdinov, B.I.; Zuev, Y.F.; Kataev, V.E. Synthesis and cytotoxicity of the conjugates of diterpenoid isosteviol and N-acetyl-D-glucosamine. *Nat. Prod. Res.* **2021**, *35*, 1372–1378. [[CrossRef](#)] [[PubMed](#)]
23. Li, J.; Zhang, D.; Wu, X. Synthesis and biological evaluation of novel exo-methylene cyclopentanone tetracyclic diterpenoids as antitumor agents. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 130–132. [[CrossRef](#)]

24. Lin, L.-H.; Lee, L.-W.; Sheu, S.-Y.; Lin, P.-Y. Study on the stevioside analogues of steviolbioside, steviol, and isosteviol 19-alkyl amide dimers: Synthesis and cytotoxic and antibacterial activity. *Chem. Pharm. Bull.* **2004**, *52*, 1117–1122. [[CrossRef](#)] [[PubMed](#)]
25. Liu, C.-J.; Liu, Y.-P.; Yu, S.-L.; Dai, X.-J.; Zhang, T.; Tao, J.-C. Syntheses, cytotoxic activity evaluation and HQSAR study of 1,2,3-triazole-linked isosteviol derivatives as potential anticancer agents. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 5455–5461. [[CrossRef](#)] [[PubMed](#)]
26. Liu, C.-J.; Wang, Y.-F.; Yao, J.-H.; Liu, Y.-P.; Jiang, Q.-J.; Liu, P.-P. Cytotoxic Activities and QSAR Studies of Diterpenoid Isosteviol Derivatives as Anti-Esophageal Agents. *Russ. J. Bioorg. Chem.* **2021**, *47*, 288–298.
27. Liu, C.-J.; Yu, S.-L.; Liu, Y.-P.; Dai, X.-J.; Wu, Y.; Li, R.-J.; Tao, J.-C. Synthesis, cytotoxic activity evaluation and HQSAR study of novel isosteviol derivatives as potential anticancer agents. *Eur. J. Med. Chem.* **2016**, *115*, 26–40. [[CrossRef](#)]
28. Liu, C.-J.; Zhang, T.; Yu, S.-L.; Dai, X.-J.; Wu, Y.; Tao, J.-C. Synthesis, cytotoxic activity, and 2D- and 3D-QSAR studies of 19-carboxyl-modified novel isosteviol derivatives as potential anticancer agents. *Chem. Biol. Drug Des.* **2017**, *89*, 870–887. [[CrossRef](#)]
29. Liu, J.; Li, L.; Li, X.; Wang, X.; Zhao, X.; Qiao, Y.; Xu, Y.; Sun, Y.; Qian, L.; Liu, Z.; et al. Discovery of lysosome-targeted covalent anticancer agents based on isosteviol skeleton. *Eur. J. Med. Chem.* **2021**, *209*, 112896. [[CrossRef](#)]
30. Malki, A.; Laha, R.; Bergmeier, S.C. Synthesis and cytotoxic activity of MOM-ether analogs of isosteviol. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 1184–1187. [[CrossRef](#)]
31. Murillo, J.A.; Echeverri, F.; Quinones, W.; Torres, F.; Isaza, L.; Robledo, S.M.; Pineda, T.; Olivo, H.F.; Escobar, G.A. Synthesis, Cytotoxicity, and Leishmanicidal Evaluation of Ent-beyerene and Ent-kaurene Derivatives. *Eur. J. Org. Chem.* **2021**, *2021*, 3386–3397. [[CrossRef](#)]
32. Sharipova, R.R.; Belenok, M.G.; Garifullin, B.F.; Sapunova, A.S.; Voloshina, A.D.; Andreeva, O.V.; Strobykina, I.Y.; Skvortsova, P.V.; Zuev, Y.F.; Kataev, V.E. Synthesis and anti-cancer activities of glycosides and glycoconjugates of diterpenoid isosteviol. *MedChemComm* **2019**, *10*, 1488–1498. [[CrossRef](#)]
33. Strobykina, I.Y.; Nemtarev, A.V.; Garifullin, B.F.; Voloshina, A.D.; Sapunova, A.S.; Kataev, V.E. Synthesis and Biological Activity of Alkane-1,1-diylbis(phosphonates) of Diterpenoid Isosteviol. *Russ. J. Org. Chem.* **2019**, *55*, 17–24. [[CrossRef](#)]
34. Ukiya, M.; Sawada, S.; Kikuchi, T.; Kushi, Y.; Fukatsu, M.; Akihisa, T. Cytotoxic and Apoptosis-Inducing Activities of Steviol and Isosteviol Derivatives against Human Cancer Cell Lines. *Chem. Biodivers.* **2013**, *10*, 177–188. [[CrossRef](#)]
35. Voloshina, A.D.; Sapunova, A.S.; Kulik, N.V.; Belenok, M.G.; Strobykina, I.Y.; Lyubina, A.P.; Gumerova, S.K.; Kataev, V.E. Antimicrobial and cytotoxic effects of ammonium derivatives of diterpenoids steviol and isosteviol. *Bioorg. Med. Chem.* **2021**, *32*, 115974. [[CrossRef](#)] [[PubMed](#)]
36. Zhang, T.; Lu, L.-H.; Liu, H.; Wang, J.-W.; Wang, R.-X.; Zhang, Y.-X.; Tao, J.-C. D-ring modified novel isosteviol derivatives: Design, synthesis and cytotoxic activity evaluation. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 5827–5832. [[CrossRef](#)] [[PubMed](#)]
37. Zhu, S.-L.; Wu, Y.; Liu, C.-J.; Wei, C.-Y.; Tao, J.-C.; Liu, H.-M. Design and stereoselective synthesis of novel isosteviol-fused pyrazolines and pyrazoles as potential anticancer agents. *Eur. J. Med. Chem.* **2013**, *65*, 70–82. [[CrossRef](#)] [[PubMed](#)]
38. Zhu, S.-L.; Wu, Y.; Liu, C.-J.; Wei, C.-Y.; Tao, J.-C.; Liu, H.-M. Synthesis and in vitro cytotoxic activity evaluation of novel heterocycle bridged carbthioamide type isosteviol derivatives as antitumor agents. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 1343–1346. [[CrossRef](#)]
39. Brandes, B.; Hoenke, S.; Starke, N.; Serbian, I.; Deigner, H.-P.; Al-Harrasi, A.; Csuk, R. Synthesis and cytotoxicity of apoptosis-inducing N-heterocyclic triterpene amides. *Eur. J. Med. Chem. Rep.* **2022**, *6*, 100085. [[CrossRef](#)]
40. Macasoi, I.; Pavel, I.Z.; Moaca, A.E.; Avram, S.; David, V.L.; Coricovac, D.; Mioc, A.; Spandidos, D.A.; Tsatsakis, A.; Soica, C.; et al. Mechanistic investigations of antitumor activity of a Rhodamine B-oleanolic acid derivative bioconjugate. *Oncol. Rep.* **2020**, *44*, 1169–1183. [[CrossRef](#)]
41. Denner, T.C.; Hoenke, S.; Kraft, O.; Deigner, H.-P.; Al-Harrasi, A.; Csuk, R. Hydroxyethylamide substituted triterpenic acids hold good cytotoxicity for human tumor cells. *Results Chem.* **2022**, *4*, 100371. [[CrossRef](#)]
42. Kroškins, V.; Lugiņina, J.; Mishnev, A.; Turks, M. Synthesis of 8-Aminoquinoline Amides of Ursolic and Oleanolic Acid. *Molbank* **2022**, *2022*, M1361. [[CrossRef](#)]
43. Heise, N.V.; Heisig, J.; Hoehlich, L.; Hoenke, S.; Csuk, R. Synthesis and cytotoxicity of diastereomeric benzylamides derived from maslinic acid, augustic acid and bredemolic acid. *Results Chem.* **2023**, *5*, 100805. [[CrossRef](#)]
44. Heise, N.V.; Hoenke, S.; Serbian, I.; Csuk, R. An improved partial synthesis of corosolic acid and its conversion to highly cytotoxic mitocans. *Eur. J. Med. Chem. Rep.* **2022**, *6*, 100073. [[CrossRef](#)]
45. Tian, T.; Liu, X.; Lee, E.-S.; Sun, J.; Feng, Z.; Zhao, L.; Zhao, C. Synthesis of novel oleanolic acid and ursolic acid in C-28 position derivatives as potential anticancer agents. *Arch. Pharm. Res.* **2017**, *40*, 458–468. [[CrossRef](#)] [[PubMed](#)]
46. Hoenke, S.; Serbian, I.; Deigner, H.-P.; Csuk, R. Mitocanic Di- and triterpenoid rhodamine B conjugates. *Molecules* **2020**, *25*, 5443. [[CrossRef](#)]
47. Kahnt, M.; Wiemann, J.; Fischer, L.; Sommerwerk, S.; Csuk, R. Transformation of asiatic acid into a mitocanic, bimodal-acting rhodamine B conjugate of nanomolar cytotoxicity. *Eur. J. Med. Chem.* **2018**, *159*, 143–148. [[CrossRef](#)]
48. Kozubek, M.; Denner, T.C.; Eckert, M.; Hoenke, S.; Csuk, R. On the influence of the rhodamine substituents onto the cytotoxicity of mitocanic maslinic acid rhodamine conjugates. *Results Chem.* **2023**, *5*, 100708. [[CrossRef](#)]
49. Kozubek, M.; Hoenke, S.; Deigner, H.-P.; Csuk, R. Betulinic acid and glycyrrhetic acid derived piperazinyl spaced rhodamine B conjugates are highly cytotoxic and necrotic. *Results Chem.* **2022**, *4*, 100429. [[CrossRef](#)]

50. Kozubek, M.; Hoenke, S.; Schmidt, T.; Stroehl, D.; Csuk, R. Platanic acid derived amides are more cytotoxic than their corresponding oximes. *Med. Chem. Res.* **2022**, *31*, 1049–1059. [[CrossRef](#)]
51. Kraft, O.; Hartmann, A.-K.; Brandt, S.; Hoenke, S.; Heise, N.V.; Csuk, R.; Mueller, T. Asiatic acid as a leading structure for derivatives combining sub-nanomolar cytotoxicity, high selectivity, and the ability to overcome drug resistance in human preclinical tumor models. *Eur. J. Med. Chem.* **2023**, *250*, 115189. [[CrossRef](#)] [[PubMed](#)]
52. Kraft, O.; Hartmann, A.-K.; Hoenke, S.; Serbian, I.; Csuk, R. Madecassic Acid-A New Scaffold for Highly Cytotoxic Agents. *Int. J. Mol. Sci.* **2022**, *23*, 4362. [[CrossRef](#)] [[PubMed](#)]
53. Kraft, O.; Hoenke, S.; Csuk, R. A tormentic acid-homopiperazine-rhodamine B conjugate of single-digit nanomolar cytotoxicity and high selectivity for several human tumor cell lines. *Eur. J. Med. Chem. Rep.* **2022**, *5*, 100043. [[CrossRef](#)]
54. Petrenko, M.; Guettler, A.; Pflueger, E.; Serbian, I.; Kahnt, M.; Eiselt, Y.; Kessler, J.; Funtan, A.; Paschke, R.; Csuk, R.; et al. MSBA-S-A pentacyclic sulfamate as a new option for radiotherapy of human breast cancer cells. *Eur. J. Med. Chem.* **2021**, *224*, 113721. [[CrossRef](#)]
55. Sommerwerk, S.; Heller, L.; Kerzig, C.; Kramell, A.E.; Csuk, R. Rhodamine B conjugates of triterpenoic acids are cytotoxic mitocans even at nanomolar concentrations. *Eur. J. Med. Chem.* **2017**, *127*, 1–9. [[CrossRef](#)]
56. Sommerwerk, S.; Heller, L.; Kuhfs, J.; Csuk, R. Selective killing of cancer cells with triterpenoic acid amides—The substantial role of an aromatic moiety alignment. *Eur. J. Med. Chem.* **2016**, *122*, 452–464. [[CrossRef](#)]
57. Shao, J.-W.; Dai, Y.-C.; Xue, J.-P.; Wang, J.-C.; Lin, F.-P.; Guo, Y.-H. In vitro and in vivo anticancer activity evaluation of ursolic acid derivatives. *Eur. J. Med. Chem.* **2011**, *46*, 2652–2661. [[CrossRef](#)]
58. Wolfram, R.K.; Heller, L.; Csuk, R. Targeting mitochondria: Esters of rhodamine B with triterpenoids are mitocanic triggers of apoptosis. *Eur. J. Med. Chem.* **2018**, *152*, 21–30. [[CrossRef](#)]
59. Siewert, B.; Pianowski, E.; Csuk, R. Esters and amides of maslinic acid trigger apoptosis in human tumor cells and alter their mode of action with respect to the substitution pattern at C-28. *Eur. J. Med. Chem.* **2013**, *70*, 259–272. [[CrossRef](#)]
60. Siewert, B.; Pianowski, E.; Obernauer, A.; Csuk, R. Towards cytotoxic and selective derivatives of maslinic acid. *Bioorg. Med. Chem.* **2014**, *22*, 594–615. [[CrossRef](#)]
61. Guzman-Villanueva, D.; Weissig, V. Mitochondria-targeted agents: Mitochondriotropics, mitochondriotoxics, and mitocans. *Handb. Exp. Pharmacol.* **2017**, *240*, 423–438. [[PubMed](#)]
62. Huang, M.; Myers, C.R.; Wang, Y.; You, M. Mitochondria as a novel target for cancer chemoprevention: Emergence of mitochondrial targeting agents. *Cancer Prev. Res.* **2021**, *14*, 285–306. [[CrossRef](#)] [[PubMed](#)]
63. Modica-Napolitano, J.S.; Aprile, J.R. Delocalized lipophilic cations selectively target the mitochondria of carcinoma cells. *Adv. Drug Deliv. Rev.* **2001**, *49*, 63–70. [[CrossRef](#)]
64. Murphy, M.P. Development of lipophilic cations as therapies for disorders due to mitochondrial dysfunction. *Expert. Opin. Biol. Ther.* **2001**, *1*, 753–764. [[CrossRef](#)] [[PubMed](#)]
65. Murphy, M.P. Targeting lipophilic cations to mitochondria. *Biochim. Biophys. Acta Bioenerg.* **2008**, *1777*, 1028–1031. [[CrossRef](#)]
66. Spivak, A.Y.; Nedopekina, D.A.; Gubaidullin, R.R.; Dubinin, M.V.; Belosludtsev, K.N. Conjugation of Natural Triterpenic Acids with Delocalized Lipophilic Cations: Selective Targeting Cancer Cell Mitochondria. *J. Pers. Med.* **2021**, *11*, 470. [[CrossRef](#)]
67. Zielonka, J.; Joseph, J.; Sikora, A.; Hardy, M.; Ouari, O.; Vasquez-Vivar, J.; Cheng, G.; Lopez, M.; Kalyanaraman, B. Mitochondria-Targeted Triphenylphosphonium-Based Compounds: Syntheses, Mechanisms of Action, and Therapeutic and Diagnostic Applications. *Chem. Rev.* **2017**, *117*, 10043–10120. [[CrossRef](#)]
68. Zinovkin, R.A.; Zamyatnin, A.A. Mitochondria-Targeted Drugs. *Curr. Mol. Pharmacol.* **2019**, *12*, 202–214. [[CrossRef](#)]
69. Wiemann, J.; Al-Harrasi, A.; Csuk, R. Cytotoxic Dehydroabietylamine Derived Compounds. *Anti-Cancer Agents Med. Chem.* **2020**, *20*, 1756–1767. [[CrossRef](#)]
70. Wiemann, J.; Fischer, L.; Kessler, J.; Stroehl, D.; Csuk, R. Ugi multicomponent-reaction: Syntheses of cytotoxic dehydroabietylamine derivatives. *Bioorg. Chem.* **2018**, *81*, 567–576. [[CrossRef](#)]
71. Wiemann, J.; Fischer, L.; Rohmer, M.; Csuk, R. Syntheses of C-ring modified dehydroabietylamine derivatives and their cytotoxic activity. *Eur. J. Med. Chem.* **2018**, *156*, 861–870. [[CrossRef](#)]
72. Kuzu, O.F.; Gowda, R.; Sharma, A.; Robertson, G.P. Leelamine Mediates Cancer Cell Death through Inhibition of Intracellular Cholesterol Transport. *Mol. Cancer Ther.* **2014**, *13*, 1690–1703. [[CrossRef](#)] [[PubMed](#)]
73. Kraft, O.; Kozubek, M.; Hoenke, S.; Serbian, I.; Major, D.; Csuk, R. Cytotoxic triterpenoid-safirinium conjugates target the endoplasmic reticulum. *Eur. J. Med. Chem.* **2021**, *209*, 112920. [[CrossRef](#)] [[PubMed](#)]
74. Kamkaew, A.; Thavornpradit, S.; Puangsamlee, T.; Xin, D.; Wanichacheva, N.; Burgess, K. Oligoethylene glycol-substituted aza-BODIPY dyes as red emitting ER-probes. *Org. Biomol. Chem.* **2015**, *13*, 8271–8276. [[CrossRef](#)]
75. Chen, P.; Zhang, D.; Li, M.; Wu, Q.; Lam, Y.P.Y.; Guo, Y.; Chen, C.; Bai, N.; Malhotra, S.; Li, W.; et al. Discovery of novel, potent, isosteviol-based antithrombotic agents. *Eur. J. Med. Chem.* **2019**, *183*, 111722. [[CrossRef](#)] [[PubMed](#)]
76. Hsu, F.-L.; Hou, C.-C.; Yang, L.-M.; Cheng, J.-T.; Chi, T.-C.; Liu, P.-C.; Lin, S.-J. Microbial transformations of isosteviol. *J. Nat. Prod.* **2002**, *65*, 273–277. [[CrossRef](#)] [[PubMed](#)]
77. Korochkina, M.; Fontanella, M.; Casnati, A.; Arduini, A.; Sansone, F.; Ungaro, R.; Latypov, S.; Kataev, V.; Alfonsov, V. Synthesis and spectroscopic studies of isosteviol-calix[4]arene and -calix[6]arene conjugates. *Tetrahedron* **2005**, *61*, 5457–5463. [[CrossRef](#)]
78. Buddrus, J.; Bauer, H. New analytical methods. Part (32). Determination of the carbon skeleton of organic compounds by double quantum coherent carbon-13 NMR spectroscopy, the INADEQUATE pulse sequence. *Angew. Chem.* **1987**, *99*, 642. [[CrossRef](#)]

79. Buddrus, J.; Lambert, J. Connectivities in molecules by INADEQUATE: Recent developments. *Magn. Reson. Chem.* **2002**, *40*, 3–23. [[CrossRef](#)]
80. Ismail, F.M.D.; Nahar, L.; Sarker, S.D. Application of INADEQUATE NMR techniques for directly tracing out the carbon skeleton of a natural product. *Phytochem. Anal.* **2021**, *32*, 7–23. [[CrossRef](#)]
81. Li, D.; Owen, N.L. Structure determination using the NMR “inadequate” technique. *Adv. Mol. Struct. Res.* **1996**, *2*, 191–211.
82. Cohen, R.D.; Wang, X.; Sherer, E.C.; Martin, G.E. Application of 1,1-ADEQUATE and DFT to correct ¹³C misassignments of carbonyl chemical shifts for carbapenem antibiotics. *Magn. Reson. Chem.* **2022**, *60*, 963–969. [[CrossRef](#)]
83. Martin, G.E. Using 1,1- and 1,n-ADEQUATE 2D NMR data in structure elucidation protocols. *Annu. Rep. NMR Spectrosc.* **2011**, *74*, 215–291.
84. Roginkin, M.S.; Ndukwe, I.E.; Craft, D.L.; Williamson, R.T.; Reibarkh, M.; Martin, G.E.; Rovnyak, D. Developing nonuniform sampling strategies to improve sensitivity and resolution in 1,1-ADEQUATE experiments. *Magn. Reson. Chem.* **2020**, *58*, 625–640. [[CrossRef](#)] [[PubMed](#)]
85. Roy, A.; Roberts, F.G.; Wilderman, P.R.; Zhou, K.; Peters, R.J.; Coates, R.M. 16-Aza-ent-beyerane and 16-Aza-ent-trachylobane: Potent Mechanism-Based Inhibitors of Recombinant ent-Kaurene Synthase from *Arabidopsis thaliana*. *J. Am. Chem. Soc.* **2007**, *129*, 12453–12460. [[CrossRef](#)] [[PubMed](#)]

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