

8 Host-Microbial Interactions Announcement

Draft genome sequence of a *Wolbachia* endosymbiont from *Syringophilopsis turdi* (Fritsch, 1958) (Acari, Syringophilidae)

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ABSTRACT We present the draft genome of a *Wolbachia* endosymbiont from quill mites. This is the first representative of a recently discovered distinct *Wolbachia* lineage (supergroup P). We hope the genome will be a useful resource for comparative evolutionary and genomic studies across the globally distributed symbiont *Wolbachia*.

KEYWORDS Wolbachia, symbiosis, evolution, mites (Acari)

W olbachia is a very common intracellular, maternally transmitted symbiont of arthropods and nematodes (1). The bacteria have profound effects on host reproduction and physiology, as well as on host-pathogen interactions (2). Due to the difficulty of culturing *Wolbachia* in cell-free media, genetically distinct *Wolbachia* lineages are designated as "supergroups" rather than species, and to date, 21 of such supergroups have been identified (3). However, many of these supergroups lack genomic representation, which hinders comparative evolutionary studies. Quill mites (Acari: Syringophilidae) are parasitic mites found in feather quills of many bird species (4). Previously, two *Wolbachia* supergroups not currently known from other hosts were detected in several species of quill mite (5, 6). We here present the first draft genome for one of these lineages, supergroup P, which is most closely related to *Wolbachia* supergroups C and F (5).

Mite material was originally sampled from feathers of Song thrush Turdus philomelos Brehm (April 2009, Kopań, Poland) and stored in 96% ethanol. DNA was extracted from a single adult individual of Syringophilopsis turdi using the DNeasy Blood & Tissue Kit (Qiagen, Hilden, Germany) by incubating the specimen in lysis buffer at 56°C overnight and digesting it using Proteinase K for 72 h as commonly done for quill mites (7). The intact exoskeleton was then removed from the buffer to be mounted on a microscopic slide and DNA extraction continued using the manufacturer's protocol. Identification was done using a microscope and the morphological characters described in reference (8). The DNA was stored in elution buffer until sequencing. The library for Illumina sequencing was prepared using a protocol for double-indexed libraries (9, 10). First, DNA was sheared using a Covaris E210 to insert sizes of 300 bp, followed by a blunt end repair using T4 DNA Polymerase, adapter ligation with T4 DNA ligase, and the Illumina adapters P5 and P7. Between each of these steps, the library was purified using SPRI beads. Library preparation success was determined by qPCR, and indices were added in an indexing PCR. The library was sequenced as 96 bp paired-end run on an Illumina HiSeq 2000. We performed base calling with freelbis using default parameters (11), trimmed adapters, and discarded all reads with more than five bases below a quality threshold of 15. Sequencing resulted in a total of 22.44 M read pairs, which were assembled using metaSPAdes version 3.15.3 (12) under default parameters. Wolbachia contigs were identified using BLAST+ searches (13) against the NCBI nt database and subsequent contig filtering by taxonomy (genus: *Wolbachia*), coverage (\geq 50×), and GC (quanine-cytosine) content (≥ 0.2 and ≤ 0.4) with Blobtools version 1.1.1. (14). Out of

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119,320 metagenomic contigs, 163 remained after this initial filtering. Paired-end reads aligning to these 163 contigs were then used for a second round of assembly using SPAdes version 3.15.3. (15) and k-mer sizes 21, 33, 55, 77, and 91, and contigs shorter than 500 bp were discarded.

The *w*Stur draft genome contains 14 contigs with an average GC content of 34% and a total length of 1,054,444 bp (N50 = 249,347 bp). The final coverage was 106×. NCBI's Prokaryotic Genome Annotation Pipeline version 6.5 (16) predicted 1,056 CDS (coding sequences), 34 of which were pseudogenes. The average CDS length is 761 bp, and the genome-wide coding density is 79%. Using the Rickettsiales-specific marker genes, we estimated the genome to be 100% complete with 0.32% contamination by CheckM version 1.1.3. (17). Using BLAST+ searches, we confirmed that the partial 16S sequence of *w*Stur. Five CDS had similarities to the *Wolbachia* insertion sequences ISWosp8, ISWen3, or ISWpi15 as determined with ISFinder (18) BLAST+ searches (e-value 1e-6). Using the PHASTER web server (19), we were unable to detect any prophage sequences in the draft genome. Overall, these characteristics indicate that *w*Stur is a typical *Wolbachia* genome that has lost most of its mobile genetic elements. The draft genome sequence will be useful for studying *Wolbachia* genome evolution and *Wolbachia*-quill mite interactions.

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AUTHOR CONTRIBUTIONS

Eliza Głowska, Conceptualization, Data curation, Methodology, Resources, Writing – review and editing | Michael Gerth, Conceptualization, Data curation, Formal analysis, Investigation, Validation, Writing – original draft, Writing – review and editing

DATA AVAILABILITY

This Whole Genome Shotgun project has been deposited in GenBank under the accession no. JAUCRB000000000. The version described in this article is the first version, JAUCRB000000000.1. Illumina reads are available under NCBI SRA accession no. SRR24987314.

REFERENCES

- Werren JH, Baldo L, Clark ME. 2008. Wolbachia: master manipulators of invertebrate biology. Nat Rev Microbiol 6:741–751. https://doi.org/10. 1038/nrmicro1969
- Kaur R, Shropshire JD, Cross KL, Leigh B, Mansueto AJ, Stewart V, Bordenstein SR, Bordenstein SR. 2021. Living in the endosymbiotic world of *Wolbachia*: a centennial review. Cell Host & Microbe 29:879–893. https://doi.org/10.1016/j.chom.2021.03.006
- Beliavskaia A, Tan K-K, Sinha A, Husin NA, Lim FS, Loong SK, Bell-Sakyi L, Carlow CKS, AbuBakar S, Darby AC, Makepeace BL, Khoo JJ. 2023. Metagenomics of culture isolates and insect tissue illuminate the evolution of *Wolbachia*, *Rickettsia* and *Bartonella* symbionts in *Ctenocephalides* spp. fleas. Microb Genom 9:001045. https://doi.org/10. 1099/mgen.0.001045
- 4. Skoracki M. 2011. Quill mites (Acari: Syringophilidae) of the palaearctic region. Zootaxa 2840:1. https://doi.org/10.11646/zootaxa.2840.1.1
- Glowska E, Dragun-Damian A, Dabert M, Gerth M. 2015. New Wolbachia supergroups detected in quill mites (Acari: Syringophilidae). Infect Genet Evol 30:140–146. https://doi.org/10.1016/j.meegid.2014.12.019
- Glowska E, Filutowska ZK, Dabert M, Gerth M. 2020. Microbial composition of enigmatic bird parasites: *Wolbachia* and *Spiroplasma* are the most important bacterial associates of quill mites. Microbiologyopen 9:e964. https://doi.org/10.1002/mbo3.964
- Dabert J, Ehrnsberger R, Dabert M. 2008. *Glaucalges tytonis* sp. n. (Analgoidea, Xolalgidae) from the barn owl *Tyto alba* (Strigiformes, Tytonidae): compiling morphology with DNA barcode data for taxon descriptions in mites (Acari) Zootaxa 1719:41-52.
- Skoracki M. 2011. Quill mites (Acari: Syringophilidae) of the palaearctic region. Zootaxa 2840:1. https://doi.org/10.11646/zootaxa.2840.1.1
- Gerth M, Gansauge M-T, Weigert A, Bleidorn C. 2014. Phylogenomic analyses uncover origin and spread of the *Wolbachia* pandemic. Nat Commun 5:5117. https://doi.org/10.1038/ncomms6117
- Meyer M, Kircher M. 2010. Illumina sequencing library preparation for highly multiplexed target capture and sequencing. Cold Spring Harb Protoc 2010:pdb.prot5448. https://doi.org/10.1101/pdb.prot5448

- Renaud G, Kircher M, Stenzel U, Kelso J. 2013. freelbis: an efficient basecaller with calibrated quality scores for Illumina sequencers. Bioinformatics 29:1208–1209. https://doi.org/10.1093/bioinformatics/ btt117
- Nurk S, Meleshko D, Korobeynikov A, Pevzner PA. 2017. metaSPAdes: a new versatile metagenomic assembler. Genome Res 27:824–834. https:// doi.org/10.1101/gr.213959.116
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421. https://doi.org/10.1186/1471-2105-10-421
- Laetsch DR, Blaxter ML. 2017. BlobTools: interrogation of genome assemblies. F1000Res 6:1287. https://doi.org/10.12688/f1000research. 12232.1
- Prjibelski A, Antipov D, Meleshko D, Lapidus A, Korobeynikov A. 2020. Using spades de novo assembler. Curr Protoc Bioinformatics 70:e102. https://doi.org/10.1002/cpbi.102
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44:6614– 6624. https://doi.org/10.1093/nar/gkw569
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. Checkm: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043–1055. https://doi.org/10.1101/gr.186072.114
- Siguier P, Perochon J, Lestrade L, Mahillon J, Chandler M. 2006. ISfinder: the reference centre for bacterial insertion sequences. Nucleic Acids Res 34:D32–6. https://doi.org/10.1093/nar/gkj014
- Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. Nucleic Acids Res 44:W16–21. https://doi.org/10.1093/nar/gkw387