

The efficacy of vitamin D supplementation and fish consumption to optimize the vitamin D status in healthy volunteers



Dissertation

zur Erlangung des Doktorgrades
der Ernährungswissenschaften (Dr. troph.)
der

**Naturwissenschaftliche Fakultät III
Institut für Agrar- und Ernährungswissenschaften
der Martin-Luther-Universität Halle-Wittenberg**

vorgelegt von

Diplom-Ernährungswissenschaftlerin Ulrike Spielau (geb. Lehmann)
geboren am 25. Juni 1986 in Bad Saarow

Gutachterinnen: Prof. Dr. Gabriele I. Stangl
Prof. Dr. Jutta Dierkes
Prof. Dr. Cornelia Weikert

Verteidigung: 3. Mai 2021

Contents

Abbreviations	I
List of tables	III
List of Figures	IV
1. Introduction	1
1.1. Absorption, degradation, metabolism and excretion	1
1.1.1. Sun exposure and skin synthesis.....	1
1.1.2. Absorption and transport of dietary Vitamin D	2
1.1.3. Metabolism	2
1.1.4. Degradation and Excretion.....	4
1.2. Effects of Vitamin D	4
1.2.1. Biochemical functions	4
1.2.2. Molecular mechanisms	5
1.3. Status assessment, recommendations and toxicity	6
1.3.1. Vitamin D status assessment.....	6
1.3.2. Dietary recommendations	8
1.3.3. Non-traditional outcomes	11
1.3.4. Toxicity	12
1.4. Dietary Vitamin D	13
1.4.1. Dietary sources.....	13
1.4.2. Vitamin D in foods.....	13
1.4.3. 25(OH)D ₃ -content of food.....	15
1.5. Vitamin D-fortified foods	16
1.5.1. Bio-fortification	16

1.6. Supplements	17
1.6.1. Supplements containing 25(OH)D ₃	18
1.7. Habitual vitamin D intake (<i>Germany and Europe</i>)	18
1.7.1. Fish consumption	19
2. Aims	21
2.1. General aims of the thesis	21
3. Studies	23
3.1. Study 1	23
3.2. Study 2	31
3.3. Study 3	40
3.3.1. Study Design	40
3.3.2. Subjects	40
3.3.3. Production of bio-fortified fish	41
3.3.4. Methods	41
3.3.5. Analysis of cholecalciferol concentration in fillet and skin of rainbow trout	42
3.3.6. Statistical analysis	42
3.3.7. Results	42
3.4. Study 4	46
4. Discussion	58
4.1. Efficacy of vitamin D supplementation to optimize vitamin D status in humans	58
4.2. Efficacy of fish and food consumption to optimize vitamin D status in humans	63
4.3. Effects of vitamin D supplementation on vitamin D metabolites and cardiovascular risk factors	66
4.4. Health and adverse effects of vitamin D supplementation and fish consumption	67

4.5. Limitations of vitamin D supplementation and food enrichment	68
4.6. Suggestions for future research.....	70
4.7. Conclusion	71
5. Zusammenfassung	72
6. Summary	74
References	V
Danksagung	XXXIII
Curriculum Vitae	XXXIV
Eidesstattliche Erklärung	XXXVI

Abbreviations

1,23,25(OH) ₃ D ₃	1,23,25-trihydroxyvitamin D ₃
1,24,25(OH) ₃ D ₃	1,24,25-trihydroxyvitamin D ₃
1,25(OH) ₂ D	1,25-dihydroxyvitamin D (calcitriol)
1,25(OH) ₂ D ₂	1,25-dihydroxyvitamin D ₂
1,25(OH) ₂ D ₃	1,25-dihydroxyvitamin D ₃
1,25(OH) ₂ D ₃ -26,23-lactone	1,25-dihydroxyvitamin D ₃ -26,23-lactone
7-DHC	7-dehydrocholesterol
24,25(OH) ₂ D ₃	24,25-dihydroxyvitamin D ₃
25(OH)D	25-hydroxyvitamin D (calcidiol)
25(OH)D ₂	25-hydroxyvitamin D ₂
25(OH)D ₃	25-hydroxyvitamin D ₃
AI	Adequate Intake
BMI	Body mass Index
CYP2R1	Cytochrom P450 2R1 (25-hydroxylase)
CYP24A1	Cytochrom P450 24A1 (25-hydroxyvitamin D-24-hydroxylase)
CYP27B1	Cytochrom P450 27B1 (25-hydroxyvitamin D 1-alpha-hydroxylase)
DACH	Deutschland-Österreich-Schweiz
DBP	Vitamin D-binding Protein
DEGS1	German Health Interview and Examination Survey for Adults
DGE	German Society for Nutrition
DNA	Deoxyribonucleic acid
DRV	Dietary reference values
EDTA	Ethylenediaminetetraacetate
EFSA	European Food Safety Authority
ELISA	Enzyme Linked Immunosorbent Assay
EU	European Union
FAO	Food and Agriculture Organization of the United Nations
FFQ	Food Frequency Questionnaire
FGF-23	Fibroblast growth factor 23

HDL	High Density Lipoprotein
HPLC	High performance liquid chromatography
HDM	Health and Medicine Division (<i>former IOM</i>)
IOM	Institute of Medicine
IU	International units
LC-MS/MS	Liquid chromatography with tandem mass spectrometry
LDL	Low Density Lipoprotein
µg	Microgram
mW/m ²	Milliwatts per square meter
nm	Nanometer
NHANES	The National Health and Nutrition Examination Survey
NNR	Nordic Nutrition Recommendations
NOAEL	No observed effect level
Npt2a/c	Sodium-dependent phosphate co-transporters
NVH II	German Nutrition Health Survey II
PKA	Protein Kinase A
PKC	Protein Kinase C
PTH	Parathyroid hormone
RCT	Randomized controlled trials
RDA	Recommended Daily Allowance
SACN	British Scientific Advisory Committee on Nutrition
SD	Standard deviation
UK	United Kingdom
UL	Tolerable upper intake level
US	United States
UV	Ultraviolet
UVB	Ultraviolet B
VDR	Vitamin D-receptor
VDR _e	Vitamin D-receptor response elements
VDSP	Vitamin D standardization program
WHO	World Health Organization

List of tables

Table 1: Differentiation limits of adult vitamin D status set by leading advisory organizations	7
Table 2: Overview of global vitamin D recommendations	9
Table 3: Vitamin D content in foods in the German food composition database	14
Table 4: Vitamin D content in fish in different food composition databases.....	15
Table 5: Baseline characteristics of the study population	43

List of Figures

Figure 1: Chemical structure of vitamin D₂ (ergocalciferol) and vitamin D₃ (cholecalciferol). 13

Figure 2: 25(OH)D₃ concentrations [nmol/l] in the control and intervention group at baseline and after four weeks. 44

Figure 3: Forest plot on the effect of daily supplementation with either vitamin D₂ or vitamin D₃ on plasma total 25(OH)D concentrations. 59

1. Introduction

As early as in the 18th century scientists recognized the positive effects of sun and cod liver oil for the prevention and treatment of rickets [Park, 1940; Stamp, 1975; Chick, 1976; DeLuca, 2014] and osteomalacia [Compston et al., 1979] which were widespread then in northern European countries [Holick, 2010]. However, it was not until a century later that US-scientist McCollum identified the active substance responsible for this medical effect [McCollum et al., 1922]: Vitamin D – a compound which can be found in high amounts in cod liver oil and is further produced in the human skin upon exposure to UVB-radiation. Research into the effects and the optimal dose of vitamin D for human health is still ongoing and has received attention in the 21st century.

1.1. Absorption, degradation, metabolism and excretion

1.1.1. Sun exposure and skin synthesis

Vitamin D is formed when the skin is exposed to UVB-radiation, which comprises not more than 2 – 12% of all UV-radiation and consists of intermediate wavelengths (UVB) between 290 and 315 nm [Holick et al., 1981]. Furthermore, UVB light does not reach the earth's surface at areas beyond 40 degrees latitude - which correspond an imaginary line drawn through Boston – from November to February [Webb et al., 1988].

UVB wavelengths cause the photoconversion of 7-dehydrocholesterol (7-DHC) into vitamin D₃ mainly in the stratum basale and stratum spinosum of the epidermis [Havinga, 1973; Holick et al., 2007]. The double-bonds of 7-DHC absorb UVB-rays leading to the opening of the B-ring at C9 as well as C10 and, ultimately, the conversion to previtamin D₃ (also named precalciferol) [Holick et al., 1981]. Previtamin D₃ is an unstable compound that is thermally isomerized, i.e. the three double bonds are re-arranged, to vitamin D₃ (cholecalciferol) [Havinga, 1973]. It consists of its characteristic structure of four combined rings (ring A-D) [Holick, 2011] with an open B-ring at C9-C10 [Jäpelt et al., 2013]. The vitamin is transported from the skin into the blood circulation where it is majorly bound to vitamin D-binding protein (DBP) [Haddad et al.,

1993] and to albumin to a lesser extent [Haddad, 1995]. With the blood stream, it is primarily carried to liver and kidney for hydroxylation.

1.1.2. Absorption and transport of dietary Vitamin D

It is estimated that the main source of vitamin D in humans is the cutaneous synthesis of cholecalciferol which accounts up to 90% of all vitamin D supply [Lehmann and Meurer, 2010], but is dependent on UVB radiation which is absent in long periods of the year in areas distant from the equator [Webb et al., 1988].

In these areas, humans depend on dietary vitamin D either from food or from supplements. Vitamin D is the overarching term for a number of substances differing in their side chains, and the most abundant forms are vitamin D₂ and vitamin D₃. Vitamin D₃ is derived from animal products and human skin [Holick et al., 2007; Schmid and Walther, 2013], while vitamin D₂ is derived from ergosterol in plants [Keegan et al., 2013]. There are little data on differences in absorption between vitamin D₂ and D₃, but it is assumed that both forms are absorbed to the same extent [Biancuzzo et al., 2010].

As with other fat soluble vitamins, dietary vitamin D is absorbed in the proximal small intestine [Dueland et al., 1983]. Dietary vitamin D reaches the enterocytes through passive diffusion. In the enterocytes, vitamin D is incorporated in chylomicrons that consist of triglycerides, cholesterol and other lipids which are released into the lymphatic system and then into the blood for transport to the liver and non-hepatic tissues [Holick, 2012]. Data on the amount of vitamin D absorption vary between 62-91% [Thompson et al., 1966]. Furthermore absorption can be affected in case of chronic malabsorption e.g. Crohn's disease [Driscoll et al., 1982].

1.1.3. Metabolism

Vitamin D is taken up by the liver for hydroxylation or – if not needed – stored in adipose tissue [Heaney et al., 2009] and skeletal muscle [Chen et al., 2007]. After hepatic uptake both forms of vitamin D go through the same process of hydroxylation. In the liver, 25-hydroxylase (*CYP2R1*) hydroxylates vitamin D₃ to 25-hydroxyvitamin D₃ (25(OH)D₃), also referred to as calcidiol, at C25 [Henry, 2011]. The compound is then

transported to the kidney, where the second hydroxylation step is performed. Due its lipophilic characteristic it has to be bound to proteins like albumin, lipoproteins, and specific DBPs in order to be transported via blood stream [Bikle et al., 1985; Chun et al., 2014]. At the kidney, the 25(OH)D₃-DBP complex enters the organ via receptor-mediated endocytosis [Nykjaer et al., 2001]. The complex directly binds to megalin and cubilin, two endocytic receptors which are mainly expressed by the epithelial cells of the proximal tubulus [Rowling et al., 2006]. In the next step, the complex is transferred into the lysosome of the proximal tubular cells [Willnow and Nykjaer, 2002], where the 25(OH)D₃ is cleaved via peptidase from DBP and is released to the cytosol. The 25(OH)D₃ is hydroxylated at C1 to 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃), also known as calcitriol, via 25-hydroxyvitamin D 1-alpha-hydroxylase (*CYP27B1*) in the renal mitochondria [Prentice et al., 2008; Bikle, 2009] and bound at DBP within the intestinal fluid [Willnow and Nykjaer, 2002].

The synthesis of 1,25(OH)₂D₃ is precisely regulated by processes within the body and influenced by a variety of endocrine factors. The 1-alpha-hydroxylation depends on healthy proximal tubular cells – a prerequisite usually not given in case of kidney disease [Gallagher et al., 2007]. Under normal conditions, low concentrations of 1,25(OH)₂D₃, high parathyroid hormone (PTH) [Henry, 2011] or low phosphate, calcium and fibroblast growth factor 23 (FGF-23) concentrations [Shimada et al., 2004; Perwad et al., 2007] induce the activation of 1-alpha-hydroxylase.

Although the kidney is the main organ containing 1-alpha-hydroxylase, other tissues or cells such as macrophages [Adams and Gacad, 1985; Crowle et al., 1987], keratinocytes, monocytes, osteoblasts, breast and colon cells also express *CYP27B1* mRNA [Bikle et al., 1994; Lehmann et al., 1999; Norman et al., 2002; Holick, 2007b]. 1,25(OH)₂D₃ mediates its effects as the active form via the vitamin D receptor (VDR) [McDonnell et al., 1987]. Vitamin D receptors are present in many tissues in the organism and mediate various physiological effects [Bikle, 2009]. The main target tissues include those involved in mineral homeostasis as bones, intestine, kidney and parathyroidea [DeLuca, 2004]. Besides, VDR is also expressed in e.g. the skin [Reichrath et al., 1994], immune cells [Provvedini et al., 1983] or placenta [Pike et al., 1980].

1.1.4. Degradation and Excretion

While degradation of vitamin D metabolites, $25(\text{OH})\text{D}_3$ and $1,25(\text{OH})_2\text{D}_3$, can be mediated through the C24 oxidation pathway [Ohnuma and Norman, 1982; Napoli and Horst, 1983]. Additionally, $1,25(\text{OH})_2\text{D}_3$ can be catabolized via the C23 lactone pathway [Ohnuma and Norman, 1982].

The 24-alpha-hydroxylase (*CYP24A1*), a mitochondrial inner membrane cytochrome P-450 enzyme, limits body concentrations of $25(\text{OH})\text{D}_3$ and $1,25(\text{OH})_2\text{D}_3$ by formation of inactive vitamin D metabolites, either 24,25-dihydroxyvitamin D_3 ($24,25(\text{OH})_2\text{D}_3$) or 1,24,25-trihydroxyvitamin D_3 ($1,24,25(\text{OH})_3\text{D}_3$) [Norman, 2008]. In the kidney, $25(\text{OH})\text{D}_3$ can be converted by *CYP24A1* at C24 to $24,25(\text{OH})_2\text{D}_3$. Additionally, high $1,25(\text{OH})_2\text{D}_3$ -concentrations and low PTH concentrations activate the C24 oxidation pathway [Kleiner-Bossaller and DeLuca, 1974; Tanaka et al., 1977] which catalyses the conversion into $1,24,25(\text{OH})_3\text{D}_3$. Within the C24 oxidation pathway, the enzyme further catalyses the formation of calcitroic acid which is water-soluble and mainly secreted into bile [Jones, 1999]. Following it is excreted with faeces and, to a lesser extent, excreted with urine [Avioli et al., 1967].

1.2. Effects of Vitamin D

1.2.1. Biochemical functions

Vitamin D is primarily responsible for the regulation of bone metabolism and mineralization which, in turn, is regulated by the skeleton's main components: calcium and phosphate [Holick, 1996]. Actions related to calcium homeostasis, i.e. bone metabolism and mineralization, are called *calcaemic effects* of vitamin D. They primarily help to ensure stabilisation of plasma calcium and phosphate concentrations and provide sufficient calcium for bone mineralization [Holick, 1996]. The active metabolite, $1,25(\text{OH})_2\text{D}_3$, has four so-called *traditional target tissues*: bone, kidney, intestine as well as parathyroid gland. One of the main functions is the homeostasis of the calcium concentration in blood within narrow bounds. The active metabolite, $1,25(\text{OH})_2\text{D}_3$ directly suppresses the synthesis of PTH in the parathyroid gland [Delmez et al., 1989]. Concentrations of PTH are largely determined by levels of $1,25(\text{OH})_2\text{D}_3$, calcium and phosphate [Lombardi et al., 2020]: A sufficient concentration of $1,25(\text{OH})_2\text{D}_3$ promotes the provision of calcium and phosphate (e.g. either through

intestinal absorption from food or re-absorption from urine in the kidney) and reduces the production of PTH. If there is an insufficient amount of $1,25(\text{OH})_2\text{D}_3$ available, PTH is produced to a larger extent. This will, if needed, trigger an increased release of calcium and phosphate from the skeleton. In the kidney, $1,25(\text{OH})_2\text{D}_3$ affects phosphate balance via regulation of bone-derived FGF-23 and its co-receptor α -Klotho [Hu et al., 2013]. Fibroblast growth factor 23 represses the reabsorption of phosphate in the kidney via inhibition of sodium-dependent phosphate co-transporter (Npt2a/c) [Gattineni et al., 2009]. Additionally, FGF-23 and α -Klotho are regulators of vitamin D metabolism [Kurosu et al., 2006]. They inhibit the activity of *CYP27B1* which converts $25(\text{OH})\text{D}_3$ into $1,25(\text{OH})_2\text{D}_3$ [Hu et al., 2013].

1.2.2. Molecular mechanisms

The biological active form, $1,25(\text{OH})_2\text{D}_3$, assumes important regulatory functions in cellular growth, differentiation and apoptosis throughout the body, e.g. bones, pancreatic tissue [Pike et al., 1980], immune system [Provvedini et al., 1983], vascular [Cardus et al., 2009] and coronary muscle cells [Wu-Wong et al., 2007], muscle [Simpson et al., 1985; Costa et al., 1986] and nerve tissues [Smolders et al., 2013]. In order to mediate its functions, $1,25(\text{OH})_2\text{D}_3$ binds to its specific vitamin D receptor (VDR), a ligand-activated transcription factor belonging to the superfamily of nuclear steroid receptors [Owen and Zelent, 2000; Yang et al., 2012]. VDR has been discovered in almost all tissues and cells in the human organism [MacDonald, 1999; Holick, 2004; Norman, 2008]. The VDR gene is located on chromosome 12, consists of nine exons [Li et al., 2009].

Ligand activation of VDR elicits the genomic pathway: The Vitamin D-VDR-complex interacts with the retinoid X receptor to form a heterodimer. The heterodimer binds to specific VDR-response-elements (VDRE) of the promoter region of $1,25(\text{OH})_2\text{D}_3$ regulated target genes and modulates transcriptions of mRNAs which encodes for different proteins. VDR can bind either co-activators or co-repressors which results in activation or repression of pathways for e.g. cell proliferation or differentiation [Evans, 1988].

It has been shown that vitamin D metabolites differ in their affinity for VDR: 1,25(OH)₂D₃ binds with high affinity to the VDR [Brumbaugh and Haussler, 1975], whereas other forms of vitamin D are significantly less potent for the receptor. For instance, the affinity of 25(OH)D₃ for VDR is 100 to 1000 times lower than of 1,25(OH)₂D₃ [Wecksler et al., 1978].

1,25(OH)₂D₃ acts also through an additional pathway: The non-genomic pathway. This pathway is responsible for rapid (seconds to minutes) cellular response effects and are mediated by activation of signal transduction pathways [Huhtakangas et al., 2004; Deeb et al., 2007].

Despite great efforts, the biochemistry of non-genomic actions and their effects on target cells is not fully understood. However it has been demonstrated that 1,25(OH)₂D₃ affected the ion-channel activity via enzymes of signal transduction as protein kinase A and C (PKA and PKC) [Duval et al., 1983]. As a result influx of calcium and phosphate may mediate vitamin D-related functions [Fleet, 2004].

1.3. Status assessment, recommendations and toxicity

1.3.1. Vitamin D status assessment

Vitamin D status is defined by serum concentrations of 25(OH)D [Dawson-Hughes et al., 2005; Holick, 2009] which are more stable and reflect body stores more tightly than the highly regulated active form 1,25(OH)₂D. The 25(OH)D has a half-time of two to three weeks and a high binding affinity to the DBP [Cooke and Haddad, 1989]. There is no consensus on the definition of normal vitamin D status [Lips, 2004; Holick, 2009] which is usually defined as deficient, inadequate or insufficient, adequate or sufficient and excessive. However, different cut-offs have been applied to define these stages. An overview on the different limits used in adults is provided in Table 1.

Table 1: Differentiation limits of adult vitamin D status set by leading advisory organizations

25(OH)D-concentration [nmol/l]	DACH	NORDEN	IOM	EFSA	Endocrine Society	SACN
<25 / 30	Deficient	Deficient	Deficient	Deficient	Deficient	Deficient
25 - 50	Insufficient	Insufficient	Uncertain ^a	Deficient	Deficient	
50 - 75	Sufficient	Sufficient	Sufficient	Sufficient	Insufficient	
> 75					Sufficient	

modified from [Lips et al., 2019]; ^a 30-50 nmol/l were adequate

DACH = [German Nutrition Society, 2012], NORDEN = [Nordic Council of Ministers, 2012], IOM = [Ross et al., 2011], EFSA = [EFSA, 2016], Endocrine Society = [Holick et al., 2011], SACN = [Scientific Advisory Committee on Nutrition, 2016]

Prolonged vitamin D deficiency promotes risk for low bone mineral density and consequently may lead to osteoporosis [Holick, 1996]. Severe vitamin D deficiency with high probability of development of clinical deficiency symptoms as rickets (in children) and osteomalacia (in adults) is considered usually at less than 12 nmol/l [Lips, 2004].

The reference used (50 or 75 nmol/l, Table 1) has huge implications on the prevalence of vitamin D deficiency, as the majority of European adults have 25(OH)D-concentrations in the range of 40 – 70 nmol/l [Burnand et al., 1992; Chapuy et al., 1997; Carnevale et al., 2001; Lamberg-Allardt et al., 2001; Gomez et al., 2004; Hintzpeter et al., 2008]. In Germany, the mean 25(OH)D-concentration was 45 nmol/l in 1998 [Hintzpeter et al., 2008] and 47 nmol/l in 2008-11 [Rabenberg et al., 2015]. Thus, prevalence of inadequacy of 57% (with 15% of the population having levels less than 25 nmol/l) have been reported in Germany when measured in 1998 [Hintzpeter et al., 2008] and about 60% when measured between 2008-2011 [Rabenberg et al., 2015]. In the latter study, only about 12% of the participants had serum 25(OH)D-concentrations exceeding 75 nmol/l.

However, there are several challenges related to the use of the limits shown. This refers first to documented differences among the assays for 25(OH)D determination [Brouwer-Brolsma et al., 2013; Cashman et al., 2013; EFSA, 2016]. The vitamin D standardization program (VDSP) surmounted a number of issues regarding these

differences [Cashman and Dowling et al., 2016], also among Liquid chromatography with tandem mass spectrometry (LC-MS/MS) methods, but in clinical practice, notable differences still occur. Furthermore, the serum 25(OH)D-concentrations in Europe undergoes a distinct seasonal variation with usually highest concentrations in late summer and lowest concentrations in late winter [Hyppönen and Power, 2007]. This variation follows a cosinor model with an amplitude of almost 16 nmol/l at a mean concentration of 60 nmol/l, as shown in a Norwegian cohort [Degerud et al., 2016]. This is usually not taken into account when status limits are defined.

1.3.2. Dietary recommendations

Dietary recommendations for vitamin D have been changed in many countries during the past decade when widespread deficiency was discovered and after a systematically update of the available literature (Table 2). Firstly, the Institute of Medicine (IOM), now the Health and Medicine Division (HDM), published new and increased vitamin D recommendations in 2011. The new intake recommendations were calculated to ensure a sufficient vitamin D status (>50 nmol/l) in 97.5% of the population [Ross et al., 2011]. Following the IOM, several European countries recalculated the vitamin D requirements and published new vitamin D recommendations [German Nutrition Society, 2012; Nordic Council of Ministers, 2012]. European recommendations usually refer to sunlight as the major source of vitamin D and are usually given for periods with limited access to UVB-radiation, e.g. in Germany from October to March [German Nutrition Society, 2012]. Indeed, most Europeans live in areas with limited UVB-radiation during winter when vitamin D supply is dependent on diet or supplements.

Table 2: Overview of global vitamin D recommendations

		DACH	NORDEN	IOM	EFSA	SACN
sufficient 25(OH)D-concentration [nmol/l]		50	50	50	50	25
Age group		Vitamin D intake in µg per day				
Infants	(1-12 months)	10	10	-	10 ^b	8.5-10
Children	(>2-18 years)	20	10	15	15	10
Females	(>18-69 years)	20	10	15	15	10
Males	(>18-69 years)	20	10	15	15	10
Elderly	(>70 years)	20	10 / 20 ^a	20	15	10
Pregnancy		20	10	15	15	10
Lactation		20	10	15	15	10

reproduced from [Pilz et al., 2018];

DACH = [German Nutrition Society, 2012], NORDEN = [Nordic Council of Ministers, 2012], IOM = [Ross et al., 2011], EFSA = [EFSA, 2016], Endocrine Society = [Holick et al., 2011], SACN = [Scientific Advisory Committee on Nutrition, 2016]

^a 20 µg for elderly > 75 years, ^b 10 µg for infants > 7 months

In the German speaking countries, the recommendations for vitamin D intake were updated in 2012, following a review of “Vitamin D and prevention of selected chronic diseases” [Linseisen et al., 2011]. This review showed a convincing evidence that vitamin D supplementation could prevent falls and fractures in old adults, and that there is a possible association of vitamin D supplementation with mobility and mortality among old adults. For other diseases, like many types of cancer, diabetes or cardiovascular diseases, either the evidence was missing, non-convincing or the database was regarded insufficient. Following this analysis, the recommendations were updated in 2012 and increased to 20 µg per day (µg/d) for the adult population [German Nutrition Society, 2012].

The IOM recognizes that dietary vitamin D is only one source of vitamin D, and that the role of dietary intake for achieving serum concentrations of 50 nmol/l is not clear. The report states that there is a lack of data on the association of vitamin D intake and health outcomes, and rather research on vitamin D status (which is the sum of diet and sun exposure) and health outcomes (p 363, [Ross et al., 2011]. The lack of a clear dose-response association of dietary vitamin D and serum concentrations of 25(OH)D

has also been highlighted by other authors [Lamberg-Allardt et al., 2013]. Indeed, the dose-response is also affected by sunshine exposure and also by the baseline concentrations of 25(OH)D.

Following the IOM approach, sufficient vitamin D status was defined as 25(OH)D-concentrations exceeding 50 nmol/l in almost all individuals by DACH [German Nutrition Society, 2012]. It was stated that the usual dietary vitamin D intake in Germany is low and not sufficient, and that either sun exposure to enhance endogenous synthesis or the intake of supplements is required to achieve a serum 25(OH)D-concentration of 50 nmol/l in 97% of the population.

Additionally other European countries changed their recommendations on vitamin D intake. For example, the Nordic countries sharing common recommendations called the Nordic Nutrition recommendations (NNR), published vitamin D recommendations in 2012 [Nordic Council of Ministers, 2012] as shown in Table 2. They also follow the approach of achieving 25(OH)D-concentrations of 50 nmol/l in the majority of the population [Lamberg-Allardt et al., 2013].

In 2016, the British Scientific Advisory Committee on Nutrition (SACN) published new recommendations to ensure a mean 25(OH)D-concentration of 25 nmol/l throughout the year in 97.5% of the population, also in periods when UVB radiation is limited [Scientific Advisory Committee on Nutrition, 2016], which are 8.5-10 µg/d vitamin D. In addition, supplements are recommended for pregnant and breastfeeding women, children <4 years, people with no or limited sun exposure, ethnic minorities and adults during wintertime (October – March).

Furthermore, the European Food Safety Authority (EFSA) changed the dietary reference values (DRV) for vitamin D in 2016 and set the adequate intake (AI) to 10 µg vitamin D in children <12 months or to 15 µg in children >12 months and adults. These DRVs apply under conditions of minimal cutaneous vitamin D synthesis, and it is concluded that in times of endogenous synthesis, less dietary vitamin D is recommended or even not required [EFSA, 2016].

The World Health Organisation (WHO) and the Food and Agriculture Organization of the United States (FAO) did not change their recommendations since 2004 [WHO & FAO, 2004].

In conclusion, discrepancies in both cut-offs of vitamin D status and for recommendations of vitamin D intake may be explained by the different health outcomes that are considered for sufficient vitamin D status. New recommendations may also take into account the association of low vitamin D status and chronic diseases [Linseisen et al., 2011] and provoke an adjustment of vitamin D recommendations in the USA. It has to be taken into account that the IOM recommendations are mainly based for achieving a sufficient vitamin D status in 50% of the population and on the importance of vitamin D status for bone health. However, even the outcome 'bone health' can be interpreted in different ways, e.g. maximising intestinal calcium absorption, bone mineral density, prevention of rickets and osteomalacia, or prevention of fractures and that these outcomes of bone health are associated with different limits and optimized 25(OH)D-concentrations [Ross et al., 2011].

Several other factors associated with 25(OH)D-status need to be considered. Among these, the most important are skin pigmentation [Holick, 2007a], age and BMI [Bischof et al., 2006; Lagunova et al., 2009]. Furthermore, lifestyle is important as it comes to time spend outside in the sun, use of sunscreens, veiling due to religious or other reasons, and smoking [Holick, 2007a]. In the German Health Interview and Examination Survey for Adults (DEGS1), as an example, obesity, low physical activity, non-use of vitamin D supplements, wintertime and media consumption were significantly associated with low 25(OH)D-concentrations in the German population [Rabenberg et al., 2015].

1.3.3. Non-traditional outcomes

In observational studies, low vitamin D status has been associated with increased risk for cardiovascular disease [Dobnig et al., 2008; Degerud et al., 2018], hypertension [Burgaz et al., 2011], diabetes [Pittas et al., 2007; Zhang et al., 2015], multiple sclerosis and other neurological [Evatt et al., 2008; Littlejohns et al., 2014] or autoimmune

diseases [Rossini et al., 2010; Wang et al., 2015] and various types of cancer [Garland et al., 1989; John et al., 1999; Ahonen et al., 2000] .

Randomized controlled trials (RCTs) with intermediate endpoints did not confirm the observational studies for cardiovascular risk factors [Jorde et al., 2010], insulin sensitivity in diabetic patients [Mousa et al., 2017]. In the meantime, several randomized clinical studies with clinical endpoints have been conducted and published, which, however, do not confirm the findings from observational studies for cardiovascular diseases [Manson et al., 2019], fractures and falls [Sanders et al., 2010; Khaw et al., 2017] and asthma [Castro et al., 2014].

1.3.4. Toxicity

High vitamin D intake may also exert toxic effects. The excess intake of vitamin D is described as “intoxication” or “hypervitaminosis D” and increases both calcium absorption in the intestines and calcium resorption from bone [Vieth, 1990; Selby et al., 1995]. This promotes both increased blood calcium levels (*hypercalcemia*) and increased calcium excretion in urine (*hypercalciuria*) [Rizzoli et al., 1994]. Furthermore, a deposition of calcium in soft tissues might cause kidney or cardiovascular damages [Zittermann and Koerfer, 2008].

In 2006, the EFSA Panel on Dietetic Products, Nutrition and Allergies evaluated the safety of vitamin D intake in all age groups and set hypercalcemia (serum calcium >2.75 nmol/l) as indicator of vitamin D toxicity [EFSA, 2016]. Taking into account studies with high vitamin D intake [Barger-Lux et al., 1998; Heaney et al., 2003], the ‘no observed effect level’ (NOAEL) was set at 250 µg/day [Hathcock et al., 2007]. When considering a safety factor of 2.5 for interindividual variation, the ‘tolerable upper intake level’ (UL) has been set to 100 µg/d for children >11 years and adults (EFSA 2012). This level is in line with the IOM that published in 2011 an UL of 100 µg/d for adults and children >9 years [Ross et al., 2011]. Indeed, it is discussed whether high serum levels of 25(OH)D exceeding 100 nmol/l, are associated with deleterious effects on the musculoskeletal system and probably on the cardiovascular system and mortality [Zittermann, 2017; Degerud et al., 2018].

1.4. Dietary Vitamin D

1.4.1. Dietary sources

What is commonly known as vitamin D is in fact a vitamin that has different chemical forms and consequently different sub-names. Therefore, various kinds of chemical vitamin D compounds with similar basic structures but different side chains exist. Vitamin D₂ (ergocalciferol) and D₃ (cholecalciferol) are the two kinds of vitamin D that are most relevant for human nutrition and vitamin D supply [Armas et al., 2004; Holick et al., 2008b; Biancuzzo et al., 2010]

In terms of chemical structure, vitamin D₂ and D₃, like all vitamin D forms, have a steroidal structure and are distinguished by different side chains (Figure 1) [Bikle, 2009]. Unlike vitamin D₃, vitamin D₂ contains one supplementary methyl-group at C24 and a double-bond at C22-C23 [Bikle, 2009; Keegan et al., 2013]. Vitamin D₂ and D₃ can be formed by UVB-radiation from their respective sterol precursors, ergosterol and 7-dehydrocholesterol. In the intestine, the absorption of vitamin D₂ and D₃ seems to be similar [Biancuzzo et al., 2010].

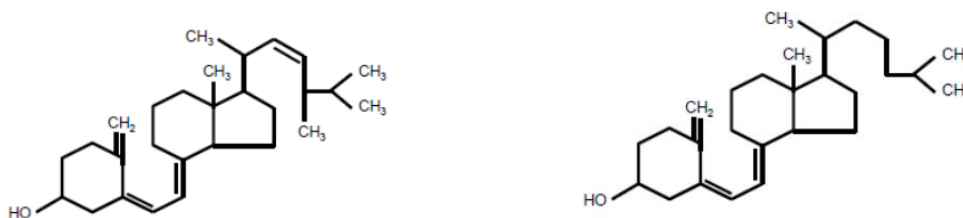


Figure 1: Chemical structure of vitamin D₂ (ergocalciferol) and vitamin D₃ (cholecalciferol). (adopted from [Norman, 2008])

1.4.2. Vitamin D in foods

Vitamin D can be obtained from food in form of vitamin D₂ and D₃ or as the hydroxylated 25(OH)D₃ [Schmid and Walther, 2013]. Fish, fish liver, egg yolk, milk and milk products are good sources of vitamin D and contain mainly vitamin D₃ [Lehmann and Meurer, 2010]. The data on the vitamin D content in foods varies between different sources,

which may be explained by differences in housing, feeding [Chick and Roscoe, 1926], seasons [Mattila et al., 2011] but also analytical methods for the determination of vitamin D [Ovesen et al., 2003; Lamberg-Allardt et al., 2013]. An overview on vitamin D content of several foods in different food composition tables is provided in Table 3.

Eggs are a valuable source of vitamin D: They contain, mainly in the egg yolk, both vitamin D₃ and 25(OH)D₃ (which has a higher bioavailability). Additionally, meat contains only small amounts of vitamin D₃ and 25(OH)D₃ [Mattila et al., 1995].

Plant sources of vitamin D are mushrooms and yeasts, which contain vitamin D₂ [Mattila et al., 1994; Hohman et al., 2011]. However, the vitamin D₂ content of plant foods is low and in the order of less than 1 µg/100 g product [Souci et al., 2016]. The low availability of plant vitamin D-rich foods leads to low intake of vitamin D in vegetarians and vegans [Outila et al., 2000].

Table 3: Vitamin D content in foods in the German food composition database

<i>Souci-Fachmann-Kraut</i>	
<i>Year</i>	2008
µg or ng/100g	
Milk	74 ng
Milk (1.5% fat)	28 ng
Yoghurt (3.5% fat)	62 ng
Yoghurt (1.5% fat)	28 ng
Cheedar cheese (50% fat)	340 ng
Edam cheese (40% fat)	290 ng
Chicken egg (total)	2.9 µg
Egg yolk	5.6 µg
Egg white	0
Mushrooms	1.9 µg
Edible boletus	3.1 µg
Butter	1.2 µg
Margarine	2.5 µg
Margarine (fat-reduced)	2.5 µg

adopted from Souci-Fachmann-Kraut [Souci et al., 2016]

Fish is the one of the major food sources of vitamin D in several European countries including France [ANSES – French Agency for Food, Environmental and Occupational Health & Safety, 2013], the United Kingdom [Henderson et al., 2003; Bates et al., 2014], Ireland [Irish Universities Nutrition Alliance, 2011], Norway [Calvo et al., 2005; Lamberg-Allardt et al., 2013], Spain [Serra-Majem et al., 2007] and Germany [Max Rubner-Institut, 2008]. Vitamin D is especially found in fatty fish like salmon, herring and mackerel [Mattila et al., 1995] and to a lower extent in lean fish like cod. Typical levels of vitamin D in fish are presented in Table 4. The data further indicate remarkable differences between food composition databases.

Table 4: Vitamin D content in fish in different food composition databases

Country	Denmark <i>Fødevare-databanken</i> (2009)	Germany <i>Souci-Fachmann-Kraut</i> (2008)	Netherlands <i>NEVO</i> (2013)	Norway <i>MVT</i> (2012)	Sweden <i>Livsmedels-databasen</i> (2014)	United Kingdom <i>McCance and Widdowson's</i> (2014)
Year	$\mu\text{g}/100\text{g}$					
Wild salmon <i>(Salmo salar L.)</i>	30.0	16.0	9.2	8.0	12.5	8.6
Farmed salmon				10.0	11.3	4.7
Atlantic herring <i>(Clupea harengus L.)</i>		25.0			8.19	
Baltic herring <i>(Clupea harengus membras L.)</i>	10.7	7.8	19.0	11.5	5.78	19.0
Farmed cod <i>(Gadus morhua L.)</i>	0.0	1.3	0.0	0.7	0.0	no data

Fødevare-databanken [Danish National Food Institute]; SFK – Souci-Fachmann-Kraut [Souci et al., 2016], NEVO [Dutch National Institute for Public Health and the Environment]; MVT – The Norwegian Food composition table [Norwegian Food Safety Authority], Livsmedels-databasen [Swedish National Food Agency]; McCance and Widdowson's [McCance and Widdowson, 2014]

1.4.3. 25(OH)D₃-content of food

Data on the content of 25(OH)D₃ in foods are limited. It is known, for instance, that human milk and eggs contain substantial amounts of the total vitamin D₃ content in the 25-hydroxylated form [Hollis et al., 1986; Mattila et al., 2011; Kühn et al., 2014].

Usually, the total vitamin D content is calculated by factorisation of vitamin D₃ by 1 and 25(OH)D₃ by 4-5 [Cashman, 2012]. This is considered in several food composition databases, e.g. Switzerland, Denmark and United Kingdom [ETH Zurich and BAG; Saxholt et al., 2008; McCance and Widdowson, 2014]. The reason for the higher factorisation of 25(OH)D₃ is that this metabolite leads to higher serum 25(OH)D-concentrations than vitamin D₃ at equimolar amounts [Cashman et al., 2012].

1.5. Vitamin D-fortified foods

There is a long tradition of fortifying foods with vitamin D in several countries, e.g. milk in the United States [Park et al., 2001], or dairy products in Scandinavian [Burgaz et al., 2007; O'Mahony et al., 2011] countries. Indeed, milk, dairy products and spreads are regarded as suitable vehicles for fortification, as they are consumed by a large proportion of the population and are widely available [O'Mahony et al., 2011].

The efficacy of vitamin D fortified foods on the vitamin D status was investigated in >25 RCTs of various duration and doses of vitamin D which have been summarized in three meta-analyses [O'Donnell et al., 2008; Black et al., 2012; Tangestani et al., 2019]. These meta-analyses concluded, despite substantial heterogeneity among studies, that consumption of food items enriched with vitamin D is effective to increase the serum 25(OH)D-concentration. However, these meta-analyses did not report whether vitamin D₃ or D₂ were added to the food items.

In the US, where food fortification has a long tradition, numerous foods i.e. orange juice, breakfast cereals, bread, cheese and milk have been enriched with vitamin D [Holden and Lemar, 2008]. The daily intake of vitamin D from fortified food accounted in the National Health and Nutrition Examination Survey (NHANES) 1999-2000 for 65-86% of total dietary vitamin D intake [Moore et al., 2005]. In the overall population, 58% of men and 39% of women used vitamin D-enriched milk, making this item to the most important single contributor for vitamin D intake [O'Mahony et al., 2011]. Data from Finland, where fortification of fluid milk products and fat spreads was introduced in 2003, suggest that this measure was effective to increase the 25(OH)D-concentration in the Finish population [Jääskeläinen et al., 2017].

In Germany, only a few products, like margarine and other mixed fat products are allowed to be fortified with vitamin D in strictly limited amounts of 25 µg/kg in margarines and mixed fat products [LMvitV].

1.5.1. Bio-fortification

Increasing the vitamin D content of animal products through increased vitamin D intake of the animal is limited in the EU as there are strict regulations for the in-feed vitamin

D content of livestock [EFSA, 2016]. Another and relatively new option is the enrichment of vitamin D in food items by exposing the food to UVB-radiation and thus inducing the synthesis of vitamin D from either ergosterol or 7-DHC. This approach has been shown in 2011 already in mushrooms [Urbain et al., 2011], and since then was followed also for fish and eggs [Kühn et al., 2014]. It seems that the amount of vitamin D can be increased substantially by this approach. Wild living animals and mushrooms growing outside which are exposed to UVB-radiation, may have higher vitamin D contents than cultivated varieties [Mattila et al., 1994; Müller-Belecke A. et al., 2014]. In this field, further research is required.

1.6. Supplements

Vitamin D supplements were originally used to treat and prevent rickets [McCollum et al., 1922]. Both vitamin D forms, D₂ and D₃, have been shown to be effective to cure rickets [Jones et al., 1998; Jurutka et al., 2001].

Vitamin D supplementation can be administered using different regimes and routes. In addition to oral supplementation, intramuscular injections of high bolus doses have been used [Romagnoli et al., 2008; Leventis and Kiely, 2009]. Bolus supplementation can also be given orally, and doses are up to 300 000 IU, although lower doses are regarded to be more physiological. The main advantage of bolus administration is to ensure compliance.

There are numerous vitamin D supplementation studies in the scientific literature, which have been summarized in several meta-analysis. For example, vitamin D₂ and D₃ supplementation was systematically evaluated by Tripkovic et al. [2012], and low-to-moderate doses of vitamin D₃ were summarized by Whiting et al. [2015], while dose-response associations by vitamin D₃ were summarized by Shab-Bidar et al. [2014]. To date, there is no systematic evaluation (e.g. as a meta-analysis) available elucidating whether bolus or continuous supplementation is more effective to increase the 25(OH)D-concentrations. Results from available literature can be summarized as follows:

- Vitamin D supplements are effective in raising 25(OH)D-concentrations, and higher doses are associated with higher increases. However, other factors affecting the increase in 25(OH)D-concentrations are the baseline 25(OH)D-concentration, duration of the study and age of participants [Shab-Bidar et al., 2014].
- There is large variation in the increase of 25(OH)D-concentrations, which is especially evident at low to moderate doses of 400-1000 IU vitamin D per day [Cranney et al., 2007; Whiting et al., 2015].
- Tripkovic [Tripkovic et al., 2012] did not observe a difference between vitamin D₂ and D₃ in their ability to raise 25(OH)D-concentrations when given as oral, continuous supplements (5 studies), but vitamin D₃ was superior to vitamin D₂ when given as bolus (3 studies).

1.6.1. Supplements containing 25(OH)D₃

Recently, 25(OH)D₃ supplements became commercially available and have been tested in comparison to vitamin D₃ in healthy volunteers [Cashman et al., 2012; Jetter et al., 2014; Minisola et al., 2017; Vaes et al., 2018]. Although the evidence is limited (as these studies included in total about 250 participants), these three studies reported superiority of 25(OH)D₃ compared to vitamin D₃ in increasing the serum 25(OH)D-concentrations. Doses tested were in the range between 5 and 20 µg/d 25(OH)D₃, and the increase in serum 25(OH)D-concentrations was about 2-3 times [Jetter et al., 2014] or up to five times [Cashman et al., 2012] the increase after similar amounts of vitamin D₃. It is of interest that doses of 15 µg [Vaes et al., 2018] and 20 µg 25(OH)D₃ µg/d [Cashman et al., 2012] increased the 25(OH)D-concentrations on average to concentrations exceeding 100 nmol/l, Further studies are needed on this topic.

1.7. Habitual vitamin D intake (*Germany and Europe*)

Dietary intake of vitamin D is usually low and the average or median population intake is below the recommended dietary intake, even if older recommendations are used. In most countries, the mean vitamin D intake is about 2-5 µg/d. The Scandinavian countries report higher intakes. Integrating additionally the intake from supplements

into the considerations, the dietary recommendations are met exclusively in Norway and Finland [Elmadfa, 2009; Flynn et al., 2009].

Vitamin D intake in the German population was reported in 2008 (data obtained in the German Nutrition Health survey (NVS II) [Max Rubner-Institut, 2008], and were at median in men and women 2.9 and 2.2 µg/d. Thus, 82% of men and 91% of women did not meet the recommendations (that were 5 µg/d at that time) [Max Rubner-Institut, 2008]. In the DEGSI from 2008-2011, vitamin D intake was assessed by a semi-quantitative food-frequency questionnaire (FFQ) allowing the estimation of low intake (<1.65 µg/d), intermediate (1.65–2.81 µg/d) or high (>2.81 µg/d), which was measured in 40, 33 and 27% of the women and in 30, 31 and 38% of the men, respectively [Rabenberg et al., 2015]. Vitamin D supplements were taken by 6% of the women and 1% of men in the DEGSI survey [Rabenberg et al., 2015].

In the NVS II, the main sources of dietary vitamin D were fish and fish dishes (47%), butter and fats (11%), eggs and egg products (11%), and dairy (10%) [Max Rubner-Institut, 2008]. Other countries also report these food types as main sources. In Norway, even though the absolute intake of vitamin D is much higher, similar contributions of these foods groups to the vitamin D intake are reported [Totland et al., 2012].

1.7.1. Fish consumption

The average annual per capita fish consumption in Germany is 13.7 kg [BLE, 2018]. Most important fish species are pollock (19.2%), salmon (17.3%), tuna (12.4%) and herring (8.9%) [FIZ, 2019]. However, fish consumption is unevenly distributed with about 50% of the population who do not consume fish at all [Max Rubner-Institut, 2008].

With such a low dietary intake, it is not surprising that large parts of the population are regarded as vulnerable groups for vitamin D deficiency. Among these are:

- pregnant and breastfeeding women [Mulligan et al., 2010; Gellert et al., 2017]
- teenagers and young women [González-Gross et al., 2012]
- old adults over 65 years [Mosekilde, 2005]
- people who are affected by low exposure to sunlight [Webb et al., 1988]
- people who have high pigmentation of skin [Holick, 2006]
- people with bowel or kidney disease [Holick, 2007a]
- overweight or obese people [Alemzadeh et al., 2008]
- vegetarians or vegans [Lamberg-Allardt et al., 1993]

In conclusion, virtually all population groups are affected by vitamin D deficiency, which is also evident from the average 25(OH)D-concentrations which are in most population groups less than 50 nmol/l.

2. Aims

2.1. General aims of the thesis

This thesis aimed to investigate whether a sufficient vitamin D status can be achieved by supplements or fish consumption. It investigates the effect of different types and doses of vitamin D supplements and the effect of regular bio-fortified fish consumption on vitamin D status in apparently healthy volunteers in three separate studies. All studies were blinded, randomized and controlled. Further, the effect of regular fish consumption on 25(OH)D-concentrations was summarized in a meta-analysis of additional nine randomized controlled studies.

Specific aims of the individual studies:

Study 1 - Bioavailability of vitamin D₂ and vitamin D₃: As there was a debate on the efficacy of vitamin D₂ compared to vitamin D₃, it was the aim to compare the efficacy of oral 50 µg/d either vitamin D₂ or D₃ to increase the total 25(OH)D-, 25(OH)D₂- and 25(OH)D₃-concentrations in healthy volunteers over a period of 8 weeks during wintertime.

Study 2 - Effect of vitamin D₃ supplementation according to the new recommendations: In 2012, the dietary recommendations (RDA) for vitamin D in the German speaking countries have been increased from 5 to 20 µg per day [German Nutrition Society, 2012]. This amount should be able to increase the serum 25(OH)D₃-concentrations above the target level of >50 nmol/l (equivalent to 20 ng/ml) in the majority of the population. Therefore, it was the aim to study the efficacy of 20 µg/d vitamin D₃ to increase 25(OH)D₃-concentrations >50 nmol/l in healthy volunteers during wintertime when endogenous vitamin D production is absent. Furthermore, the study wanted to investigate the effect on relevant vitamin D metabolites and cardiovascular risk factors.

Study 3 - Efficacy of vitamin D-fortified rainbow trout on vitamin D status: The main source of vitamin D₃ in the diet is fish, especially fatty fish. However, it was unclear to which extent serum 25(OH)D₃-concentrations can be increased due to fish consumption. Therefore, the effect on 25(OH)D₃-concentrations after six times/week

consumption of 100 g rainbow trout that was fortified with vitamin D through UVB treatment was compared to ordinary rainbow trout in healthy volunteers over a period of four weeks.

Study 4 - Efficacy of fish consumption on vitamin D status: However, effects on the 25(OH)D-concentrations from other fish consumption studies were so far not systematically investigated. Therefore, it was the aim to conduct a systematic review and a meta-analysis on the effect on 25(OH)D-concentrations from randomized controlled fish consumption studies and derive an estimate of fish intake that ensures sufficient vitamin D status.

3. Studies

3.1. Study 1

Lehmann, U., Hirche, F., Stangl, G.I., Hinz, K., Westphal, S., & Dierkes, J. (2013). Bioavailability of vitamin D2 and D3 in healthy volunteers, a randomized placebo-controlled trial. *The Journal of Clinical Endocrinology & Metabolism*, 98(11), 4339-4345.

Bioavailability of Vitamin D₂ and D₃ in Healthy Volunteers, a Randomized Placebo-Controlled Trial

Ulrike Lehmann, Frank Hirche, Gabriele I. Stangl, Katja Hinz, Sabine Westphal, and Jutta Dierkes

Institute of Agricultural and Nutritional Sciences (U.L., F.H., G.I.S.), Martin-Luther University Halle-Wittenberg, 06110 Halle, Germany; Institute of Clinical Chemistry and Biochemistry (K.H., S.W.), Otto-von-Guericke University Magdeburg, 39106 Magdeburg, Germany; and Department of Clinical Medicine (J.D.), University of Bergen, N-5020 Bergen, Norway

Background: The bioequivalence of the different forms of vitamin D, ergocalciferol (vitamin D₂) and cholecalciferol (vitamin D₃), has been questioned. Earlier studies have suggested that vitamin D₂ is less biologically active than vitamin D₃.

Objective and Design: In a parallel study, we tested the effects of supplementation with 50- μ g/d doses of vitamin D₂ or D₃ or a placebo over a period of 8 weeks on 25(OH)D₂, 25(OH)D₃, their sum 25(OH)D (primary outcome variables), and PTH in healthy volunteers applying a double-blind, randomized study design. The study was conducted during the winter of 2012 in Halle (Saale), Germany, at latitude 51°47N, when UVB irradiation is virtually absent. Blood samples for the determinations of vitamin D status and PTH were collected at baseline and after 4 and 8 weeks of supplementation.

Results: In the placebo group (n = 19), 25(OH)D₃ decreased from 39.4 \pm 14.2 to 31.1 \pm 12.4 nmol/L after 8 weeks (P < .01). In the vitamin D₃ group (n = 42), the concentrations of 25(OH)D₃ increased from 41.5 \pm 22.8 nmol/L at baseline to 88.0 \pm 22.1 nmol/L after 8 weeks (P < .01). In the group receiving vitamin D₂ (n = 46), the 25(OH)D₂ concentrations increased significantly, whereas the 25(OH)D₃ concentration fell from 36.4 \pm 13.3 nmol/L at baseline to 16.6 \pm 6.3 nmol/L after 8 weeks (P < .01). The total 25(OH)D was not different between the groups at baseline but differed significantly between the groups after 4 and 8 weeks (P < .001).

Conclusions: Vitamin D₃ increases the total 25(OH)D concentration more than vitamin D₂. Vitamin D₂ supplementation was associated with a decrease in 25(OH)D₃, which can explain the different effect on total 25(OH)D. (*J Clin Endocrinol Metab* 98: 4339–4345, 2013)

Vitamin D exists in two different forms: ergocalciferol (vitamin D₂), which occurs in plants, mainly in mushrooms; and cholecalciferol (vitamin D₃), which occurs in animals and is also produced in human skin. Vitamins D₂ and D₃ differ only in their side chains. The best dietary sources of vitamin D are fatty fish and products fortified with vitamin D (1, 2). It has been estimated that most of the vitamin D₃ in humans is derived from endogenous synthesis in the epidermis, which contains 7-dehydrocholesterol as a precursor for vitamin D₃, after irradi-

ation with UVB light at wavelengths of 290–330 nm (3). Although vitamin D₂ is less frequently used in Europe, it is the standard form of fortification and supplementation outside Europe.

Thus, both forms can be found in human blood, as well as the hydroxylated forms 25(OH)D₂ and 25(OH)D₃.

It has been debated for many years whether the two forms are bioequivalent. A number of studies have shown that vitamin D₂ does not increase the serum total 25(OH)D concentrations to the same extent as vitamin D₃

(4–6), but this finding has also been questioned by other investigators (7, 8). Because fortification or supplementation with vitamin D is currently the subject of much discussion owing to the widespread occurrence of vitamin D deficiency in nearly all populations investigated (9–19), it is important to know which form is more effective in supplementation and fortification. Although some studies have already shown that serum 25(OH)D₃ is lowered after the administration of vitamin D₂, either these studies lack sufficient statistical power (5, 20) and a control group (21) and they measured only total 25(OH)D (6, 7), or they were conducted in specific population groups (eg, elderly) (21, 22). Furthermore, it seems that the route of administration (bolus vs daily) may affect the comparison of both vitamin D forms. A recent meta-analysis showed that there was no significant difference in total 25(OH)D after daily administration of either vitamin D₂ or vitamin D₃ (1). In this meta-analysis, studies using 1000–1600 IU of vitamin D₂ or vitamin D₃ were included, but it was also estimated that larger, more robust trials are required that further address this issue.

We therefore conducted a bioavailability study in healthy volunteers who received a placebo—50 μg/d of vitamin D₂ or 50 μg/d of vitamin D₃ (2000 IU/d). The aim was to investigate the effects of this high dose on the serum levels of the hydroxylated forms 25(OH)D₂ and 25(OH)D₃ and on their sum total 25(OH)D. In addition, we investigated PTH concentrations, which are regarded as a functional parameter of vitamin D status (23). The measurability of 25(OH)D₃ serum or plasma levels is superior to that of 1,25(OH)₂D₃, owing to the much lower concentrations of 1,25(OH)₂D₃ and its shorter half-life compared with 25(OH)D₃ (24). The PTH concentrations are higher in the presence of vitamin D deficiency and decline upon supplementation with vitamin D; they can therefore be used as a functional parameter of vitamin D metabolism.

Furthermore, due to the inclusion of a placebo group, we were able to monitor the decrease of 25(OH)D₃ and total 25(OH)D in healthy subjects during wintertime at latitude 51°North.

Subjects and Methods

Design

The trial was conducted as a double-blind, randomized study in parallel groups during January, February, and March 2012, when virtually no UVB irradiation is measurable in Halle and the surrounding region. Study visits were scheduled at baseline and after 4 and 8 weeks. The subjects were randomized (stratified for body mass index [BMI] as determined during the screening visit) to receive vitamin D₂ (50 μg/d; n = 46), vitamin D₃ (50 μg/d; n = 42), or placebo (n = 19).

The supplements were manufactured commercially (Zein-Pharma) and were outwardly indistinguishable from one another. The tablets were tested for their vitamin D content after the study by a liquid chromatography, tandem mass spectrometry method in four separate runs, and the content was found to be 54 ± 12 μg for vitamin D₂ and 48 ± 6 μg for vitamin D₃ per tablet.

The participants were issued containers of tablets at baseline and after 4 weeks and were instructed to take one tablet orally per day and to return any remaining tablets at 4 and 8 weeks. The containers were numbered by an investigator with no involvement in the trial. All investigators were unaware of the order of numbering. The participants were enrolled by the physician involved in the trial but were assigned to the intervention by another investigator. Compliance, which was checked by counting the returned tablets, was 97%. During each study visit, a venous blood sample was collected for determination of 25(OH)D₂, 25(OH)D₃, their sum 25(OH)D, PTH, and serum calcium. The samples were frozen at –80°C until the time of analysis. The study protocol had been evaluated and approved by the Ethics Committee of the Medical Faculty at the Martin-Luther-University Halle-Wittenberg, and each participant gave his or her written, informed consent before the start of the study. The study was registered at clinicaltrials.gov (NCT01503216).

Subjects

Participants were recruited through newspaper advertisements, personal contacts, and information in public institutions. During a screening in the autumn (about 2 mo before the start of the study), the participants answered a self-administered questionnaire on their medical history, weight, height, lifestyle (smoking, use of sun blocker-containing cosmetics), and dietary habits relating to food rich in vitamin D. The exclusion criteria were: use of vitamin D and calcium supplements, history of chronic illness and elevated serum creatinine (in females, ≥1.1 mg/dL; in males, ≥1.3 mg/dL), elevated serum calcium, pregnancy or lactation, and vacations in areas with abundant UVB irradiation in the course of the study.

A total of 119 subjects had been recruited for the intervention study (age range, 19–67 y), were finally included in the study, and were randomized by a computer-generated randomization list to the intervention groups with the BMI as the stratification criterion. Participants were randomized into three groups according to their BMI: normal weight (BMI below 25 kg/m²), overweight (25–30 kg/m²), and obese (above 30 kg/m²). Before the start of the intervention, seven subjects (placebo group, n = 1; vitamin D₂ group, n = 3; vitamin D₃ group, n = 3) dropped out. During the study period, five subjects (vitamin D₂ group, n = 1; vitamin D₃ group, n = 4) dropped out for personal reasons. During each visit, the participants were interviewed about any adverse effect. In addition, the calcium concentration in serum was measured in serum obtained at each visit.

After completion of the study, all subjects, including those in the control group, were informed about their vitamin D status and offered vitamin D supplements.

Analytical methods

Serum concentrations of total 25(OH)D, 25(OH)D₃, and 25(OH)D₂ were determined by liquid chromatography coupled with mass spectrometry (MassChrom 25-OH Vitamin D₃/D₂ reagent kit for liquid chromatography, tandem mass spectrom-

Table 1. Characteristics of Study Participants at Baseline

	Vitamin D ₂ Group	Vitamin D ₃ Group	Placebo Group	P (ANOVA)
n	46	42	19	
Age, y	33.2 ± 12.4	35.6 ± 13.5	31.6 ± 9.3	.445
No. of males/females	15/31	16/26	8/11	.745
BMI, kg/m ²	23.7 ± 3.8	24.0 ± 4.2	23.7 ± 4.9	.928
Systolic blood pressure, mm Hg	121 ± 14	120 ± 15	115 ± 8	.201
Diastolic blood pressure, mm Hg	76 ± 8	76 ± 10	75 ± 6	.894
Creatinine at screening, mg/dL	0.80 ± 0.22	0.86 ± 0.23	0.88 ± 0.24	.298

Data are expressed as mean ± SD.

etry analysis; Chromsystems Instruments and Chemicals GmbH) on an API 2000 system (Applied Biosystems). The coefficient of variation for the determination of 25(OH)D₂ was 3.1% at a concentration of 44.8 nmol/L; for 25(OH)D₃, it was 5.3% at a concentration of 42.8 nmol/L. Total 25(OH)D was calculated as the sum of 25(OH)D₂ and 25(OH)D₃. The detection limit for both 25(OH)D₂ and 25(OH)D₃ was 2.5 nmol/L, and the limit of quantification was 7.5 nmol/L. However, the measured levels were used for the calculation of total 25(OH)D as the sum of 25(OH)D₂ and 25(OH)D₃, even in subjects with 25(OH)D₂ levels below the limit of quantification.

Intact PTH was measured in the serum by an ELISA (Biomerica Inc). Serum creatinine was determined spectrophotometrically (DiaSys Diagnostic Systems GmbH).

Statistical analyses

Statistical analyses were performed using PASW version 18.0 (SPSS Inc). All data are expressed in the form of means ± SD, with $P < .05$ as the significance threshold. The primary outcome variables were the 25(OH)D₂, 25(OH)D₃, and total 25(OH)D concentrations. These variables and PTH concentrations are presented in Table 2. Because changes in total 25(OH)D and PTH tend to depend on the baseline level, we used repeated measure analysis to analyze changes upon supplementation. We used the generalized linear models repeated measures procedure in PASW for this analysis. Total 25(OH)D and 25(OH)D₃ at baseline and at 4 and 8 weeks were used as the within-subjects factor, and the supplementation group was used as the between-subjects factor. In addition, post hoc analyses by Scheffé were used to detect differences between single groups. PTH was highly skewed and was therefore analyzed by the nonparametric Kruskal-Wallis test.

In addition, we calculated the absolute change and the percentage change in total 25(OH)D, 25(OH)D₃, and PTH (8 wk – baseline) and compared these changes among groups by ANOVA (Table 3).

According to a power calculation, 50 subjects per group would be required to show a difference of 10 nmol/L in the mean total 25(OH)D concentration after 8 weeks of supplementation between the vitamin D₂ and D₃ groups (at an assumed standard variation of 15 nmol/L for each group, at a power of 80%, and a significance level of 0.05). Because it was the main aim to compare vitamin D₂ with D₃, the size of the placebo group was only about half that of the vitamin D groups. Only subjects who finished the study according to protocol were included into the analyses.

Results

The characteristics of the subjects are presented in Table 1. The average total 25(OH)D concentration at baseline in January was 40.2 ± 18.0 nmol/L, indicating a high degree of suboptimal vitamin D status in these healthy, young volunteers, with no significant differences between the groups. The total 25(OH)D concentration increased significantly throughout the study in the groups supplemented with vitamin D₂ or vitamin D₃ and decreased significantly to 33.1 ± 13.9 nmol/L after 4 weeks and to 32.1 ± 12.8 nmol/L after 8 weeks in the placebo group. After 4 and 8 weeks, the 25(OH)D concentrations differed significantly between the groups (Table 2).

At baseline, the 25(OH)D₂ concentration was below the limit of quantification (7.5 nmol/L) in all but two participants. In neither the vitamin D₃ group nor the placebo group did the average 25(OH)D₂ rise above the limit of quantification in the course of the study. In the vitamin D₂ group, 25(OH)D₂ increased significantly to 39.6 ± 11.7 nmol/L at 4 weeks and to 51.2 ± 18.5 nmol/L at 8 weeks (Table 2).

At baseline, there was no difference in the 25(OH)D₃ concentration between the groups. Although in the vitamin D₃ group 25(OH)D₃ increased significantly after 4 and 8 weeks, it decreased significantly in the vitamin D₂ and placebo groups. The decrease was more pronounced in the vitamin D₂ group, and the difference from the placebo group was significant at both 4 and 8 weeks (Table 2).

The increases (4-wk baseline, 8-wk baseline) in the specific hydroxylated forms of vitamin D [either 25(OH)D₂ or 25(OH)D₃] were as follows: in the case of 25(OH)D₂ in the vitamin D₂ group, 38.4 ± 11.0 nmol/L after 4 weeks and 50.0 ± 18.0 nmol/L after 8 weeks; in the case of 25(OH)D₃ in the vitamin D₃ group, 34.2 ± 17.2 nmol/L after 4 weeks and 46.7 ± 21 nmol/L after 8 weeks. The increase was calculated from the baseline value in this group, without taking the decrease in 25(OH)D₃ in the placebo group into account. The increase was not significantly different at either 4 or 8 weeks.

Table 2. Vitamin D Metabolites in Healthy Volunteers Receiving Supplementation With Vitamin D₂, Vitamin D₃, or Placebo for 8 Weeks

	Vitamin D ₂ Group	Vitamin D ₃ Group	Placebo Group	P (ANOVA)
n	46	42	19	
Total 25(OH)D				
Baseline, nmol/L	37.6 ± 13.3	43.7 ± 23.3	40.7 ± 14.5	.292
4 wk, nmol/L	59.9 ± 15.2 ^a	77.1 ± 23.5 ^b	33.1 ± 13.9	.001
8 wk, nmol/L	67.8 ± 20.1 ^a	89.2 ± 22.1 ^b	32.1 ± 12.8	.001
Repeated measure analysis				<.001
25(OH)D ₃				
Baseline, nmol/L	36.4 ± 13.3	41.5 ± 22.8	39.4 ± 14.2	.409
4 wk, nmol/L	20.3 ± 8.1 ^a	75.7 ± 23.2 ^b	31.1 ± 13.9	.001
8 wk, nmol/L	16.6 ± 6.3 ^a	88.0 ± 22.1 ^b	31.1 ± 12.4	.001
Repeated measure analysis				.001
25(OH)D ₂				
Baseline, nmol/L	<7.5 ^c	<7.5	<7.5	.110
4 wk, nmol/L	39.6 ± 11.7 ^a	<7.5	<7.5	.001
8 wk, nmol/L	51.2 ± 18.5 ^a	<7.5	<7.5	.001
Repeated measure analysis				.001
PTH				
Baseline, ng/mL	69.8 ± 45.2	59.3 ± 22.6	79.4 ± 49.2	.334
4 wk, ng/mL	63.0 ± 33.2	49.1 ± 19.5	65.0 ± 40.0	.086
8 wk, ng/mL	56.8 ± 26.5	40.3 ± 19.5	60.8 ± 38.1	.007
Repeated measure analysis				.651

Data are shown as mean ± SD. Differences between the groups at the various time points of the study were tested by one-way ANOVA with post hoc Scheffé comparison. The overall effect of supplementation was tested by an ANOVA with repeated measurement (PASW procedure GLM repeated measure). Due to the high degree of skewness, the Kruskal-Wallis test was used for testing differences in PTH between groups.

^a Significantly different at $P < .01$ from vitamin D₃ group and placebo.

^b Significantly different at $P < .01$ from vitamin D₂ group and placebo.

^c Values for 25(OH)D₂ at baseline and in the vitamin D₃ and placebo groups in the course of the study are only provided for those levels exceeding the limit of detection (>2.5 nmol/L).

The PTH concentrations were not significantly different between the groups at baseline or after 4 and 8 weeks (Table 2). PTH concentrations decreased significantly during the course of the study in all groups.

The absolute and percentage differences in total 25(OH)D, 25(OH)D₃, and 25(OH)D₂ between baseline and 8 weeks were significant among the supplementation groups. Absolute or percentage differences in PTH

concentrations were not significant among the groups (Table 3).

No adverse effects were reported by the participants. Serum calcium did not exceed the normal range in any of the participants (data not shown). The analysis for total 25(OH)D, the primary outcome variable, was repeated with all randomized subjects included (intention-to-treat analysis). This did not change the results (data not shown).

Table 3. Absolute and Percentage Changes in Total 25(OH)D, 25(OH)D₃, 25(OH)D₂ (Absolute Change Only), and PTH at 8 Weeks Compared to Baseline

	Vitamin D ₂ Group	Vitamin D ₃ Group	Placebo Group	P (ANOVA)
n	46	42	19	
Δ Total 25(OH)D at 8 wk (to baseline), nmol/L	+30.2 ± 20.1 ^c	+45.5 ± 21.7 ^a	-8.6 ± 7.3	.001
% Total 25(OH)D at 8 wk (of baseline)	200 ± 97% ^a	259 ± 149% ^a	79 ± 16%	.001
Δ 25(OH)D ₃ at 8 wk (to baseline), nmol/L	-19.8 ± 9.6 ^c	+46.5 ± 21.3 ^b	-8.3 ± 6.1	.001
% 25(OH)D ₃ at 8 wk (of baseline)	47 ± 14%	280 ± 183% ^b	79 ± 15%	.001
Δ 25(OH)D ₂ at 8 wk (to baseline), nmol/L	+43.7 ± 18.5 ^d	<7.5	<7.5	.001
Δ PTH at 8 wk (to baseline), ng/mL	-13.0 ± 35.4	-19.0 ± 29.4	-18.6 ± 35.1	.658
% PTH at 8 wk (of baseline)	95 ± 47%	80 ± 58%	82 ± 38%	.354

Data are shown as mean ± SD. Significance was tested by ANOVA, followed by a post hoc Scheffé comparison.

^a Significantly different from placebo group.

^b Significantly different from vitamin D₂ and placebo groups.

^c Significantly different from vitamin D₃ and placebo groups.

^d Significantly different from vitamin D₂ and vitamin D₃ groups.

Discussion

Our major finding is that vitamin D₃ increased 25(OH)D more effectively than vitamin D₂. By measuring the specific hydroxylated forms, we have been able to show that the underlying reason for this difference is a substantial decrease in 25(OH)D₃ in subjects receiving vitamin D₂. This had not been demonstrated earlier with sufficient statistical power. We have also been able to show that hydroxylation of vitamin D₂ was similar to hydroxylation of vitamin D₃ because the increase in the specific hydroxylated forms [25(OH)D₂ and 25(OH)D₃] was similar in the two groups (compare the absolute differences in Table 3).

Vitamins D₂ and D₃ have been compared earlier in a number of studies that differed in their design, supplement dosage, frequency of supplementation, use of the delivery method, and selection of participants and also in their conclusion regarding the bioequivalence of the two forms of the vitamin (4–8, 20–22, 25, 26). A recent meta-analysis that included seven of these studies (4–8, 21, 22) concluded that the change in 25(OH)D was significantly greater after supplementation with vitamin D₃ than after one with vitamin D₂, although the effect was largely due to the studies that used a bolus dose; it was not significant in studies with daily supplementation (1). However, for the latter analysis, only six studies (6–8, 21, 22) with a total number of 248 participants were available. Our study with 42 and 46 participants in the vitamin D₃ and D₂ groups, respectively, would have changed the result of this analysis, yielding a significant effect in favor of vitamin D₃ compared to vitamin D₂ also with daily supplementation (the analysis using present data in addition to those of Tripkovic et al [1] was made using Review Manager 5.2; data not shown).

The most interesting result of our study, however, is the decrease in 25(OH)D₃ after supplementation with vitamin D₂. This was already evident after 4 weeks, and the decrease was significantly different from the seasonal decrease observed in the placebo group. A decrease in 25(OH)D₃ after supplementation with vitamin D₂ was reported earlier by Glendenning et al (22) in elderly hip fracture patients receiving 1000 IU/d for a period of 3 months, and also by Armas et al (26), who studied single doses of 50 000 IU of D₂ and D₃ in healthy men with a follow-up period of 28 days. Interestingly, both groups of authors did not discuss these findings specifically. This was also observed by Binkley et al (21) after administration of 1600 IU daily for a period of 12 months. It is surprising that this effect was observed in only a few studies, although it should be pointed out that only studies using methods capable of distinguish-

ing between 25(OH)D₂ and 25(OH)D₃ would be able to show this effect. The use of immunoassays will therefore not make it possible to observe the effect. The biological reason behind this finding remains to be elucidated.

It has been suggested that an increased catabolism of 25(OH)D takes place due to supplementation with vitamin D₂ (5). Heaney et al (5) studied, groups of 16 and 17 subjects who received 50 000 IU once weekly for 12 weeks, and a significantly higher AUC_{25(OH)D} was observed after 84 days for vitamin D₃. Interestingly, vitamins D₃ and D₂ were also measured in the fat tissue of two participants, and a decrease in vitamin D₃ in fat tissue after supplementation with vitamin D₂ was observed. Because the authors measured vitamin D₂ in fat biopsies from only two participants, however, this finding did not reach statistical significance.

It has also been suggested that one reason for the lower increase in 25(OH)D after vitamin D₂ in comparison with supplementation with D₃ was due to impaired hydroxylation at C25 (atom of the vitamin D molecule) (27). We have shown that at least the increases in the specific hydroxylation products [either 25(OH)D₂ or 25(OH)D₃] were similar. However, we cannot exclude the possibility that vitamin D₂ impairs hydroxylation of vitamin D₃, which is also present in the circulation. Because the decrease in 25(OH)D₃ exceeded the observed decrease in the placebo group, this is a likely explanation. The problem should be investigated further.

Other explanations include an increased catabolism of the 25(OH)D₂ molecule due to a lower degree of binding to the vitamin D binding protein (28). Our data do not support an increased catabolism of 25(OH)D₂, although they cannot exclude it.

Because we did not measure any other metabolite [24,25(OH)₂D metabolites, 1,24,25(OH)₃D metabolites], we can only speculate about differences in the 24-hydroxylation step between 25(OH)D₂ and 25(OH)D₃. Further studies should include these metabolites to obtain a deeper insight into the competitive nature of the two forms of vitamin D.

Our study has several strengths and also some limitations. The strengths of the present study include its large sample size, which allowed us to detect small differences between vitamin D₂ and D₃ treatments that earlier studies had been unable to show. Another important strength is the measurement of both 25(OH)D₂ and 25(OH)D₃ in this study. Measurements of the specific hydroxylated forms of vitamin D enabled us to show the effect of vitamin D₂ on the 25(OH)D₃ levels. In addition, due to the inclusion of the placebo group, we were able to monitor the decrease in total 25(OH)D concentrations within healthy subjects living at the approximate latitude 51°North. We

observed a strong decrease from January to February and no further decrease from February to March.

One limitation of our study was that we did not measure the active forms, 1,25(OH)₂D₂ and 1,25(OH)₂D₃, or other metabolites. In addition, we did not obtain a dose-response curve after a single dose, and we did not determine the catabolic products 24,25(OH)₂D, 24,25(OH)₂D₃, or 24,25(OH)₂D₂. Measurement of these metabolites would provide valuable insights into the metabolism of vitamin D₃ in the presence of vitamin D₂. We also studied only one dose, and the level of 50 μg/d is beyond current recommendations and fortification levels.

In future studies, the effect of lower doses of vitamin D that are closer to the recommended daily amounts should be investigated. In light of the decrease in 25(OH)D₃ by vitamin D₂, the effect of vitamin D₂ supplementation on disease outcomes, eg, bone health and fractures, should be carefully analyzed. Indeed, the effect of vitamin D₂ on falls was found to be lower than that of vitamin D₃ in recent meta-analyses (29, 30).

PTH and vitamin D are both involved in bone metabolism (31) and show an inverse correlation. PTH secretion is directly modulated (23) and suppressed by 25(OH)D concentrations (31). Leventis and Kiely (32) demonstrated that vitamin D₃ affected PTH concentration more than vitamin D₂, a finding that is not supported by our data. However, our study was not designed to demonstrate an effect of vitamin D supplementation on PTH concentrations as the primary outcome. To demonstrate such an effect, we had to include even more subjects due to the large variation in PTH concentrations. Therefore, we may have missed an effect of vitamin D supplementation on PTH concentrations. This is in line with a number of other studies (8, 21, 22).

In conclusion, we have shown that vitamin D₃ is more effective in raising the vitamin D status than vitamin D₂ and that vitamin D₂ supplementation causes a decrease in 25(OH)D₃. These findings question the usefulness of vitamin D₂ supplements. Instead, vitamin D₃ should be used for supplementation and fortification purposes.

Acknowledgments

Address all correspondence and requests for reprints to: Jutta Dierkes, PhD, Department of Clinical Medicine, P.O. Box 7804, N-5020 Bergen, Norway. E-mail: jutta.dierkes@med.uib.no.

This work was supported by Grant 0315668A from the German Ministry of Education and Research.

Author contributions: J.D. and G.I.S. designed the research. U.L., K.H., S.W., and F.H. conducted the study. J.D. and U.L. analyzed the data and wrote the paper. J.D. has primary responsibility for the final content. All authors read and approved the final manuscript. None of the authors declared any financial or

personal relationships with other persons or organizations that could have an inappropriate influence on this work. The funding agency had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Registered at clinicaltrials.gov with identifier NCT01503216.

Disclosure Summary: The authors have nothing to disclose.

References

1. Tripkovic L, Lambert H, Hart K, et al. Comparison of vitamin D₂ and vitamin D₃ supplementation in raising serum 25-hydroxyvitamin D status: a systematic review and meta-analysis. *Am J Clin Nutr*. 2012;95:1357–1364.
2. Pramyothin P, Holick MF. Vitamin D supplementation: guidelines and evidence for subclinical deficiency. *Curr Opin Gastroenterol*. 2012;28:139–150.
3. Holick MF. The cutaneous photosynthesis of previtamin D₃: a unique photoendocrine system. *J Invest Dermatol*. 1981;77:51–58.
4. Romagnoli E, Mascia ML, Cipriani C, et al. Short and long-term variations in serum calcitropic hormones after a single very large dose of ergocalciferol (vitamin D₂) or cholecalciferol (vitamin D₃) in the elderly. *J Clin Endocrinol Metab*. 2008;93:3015–3020.
5. Heaney RP, Recker RR, Grote J, Horst RL, Armas LA. Vitamin D(3) is more potent than vitamin D(2) in humans. *J Clin Endocrinol Metab*. 2011;96:E447–E452.
6. Trang HM, Cole DE, Rubin LA, Pierratos A, Siu S, Vieth R. Evidence that vitamin D₃ increases serum 25-hydroxyvitamin D more efficiently than does vitamin D₂. *Am J Clin Nutr*. 1998;68:854–858.
7. Holick MF, Biancuzzo RM, Chen TC, et al. Vitamin D₂ is as effective as vitamin D₃ in maintaining circulating concentrations of 25-hydroxyvitamin D. *J Clin Endocrinol Metab*. 2008;93:677–681.
8. Biancuzzo RM, Young A, Bibuld D, et al. Fortification of orange juice with vitamin D(2) or vitamin D(3) is as effective as an oral supplement in maintaining vitamin D status in adults. *Am J Clin Nutr*. 2010;91:1621–1626.
9. Hintzpeter B, Scheidt-Nave C, Müller MJ, Schenk L, Mensink GB. Higher prevalence of vitamin D deficiency is associated with immigrant background among children and adolescents in Germany. *J Nutr*. 2008;138:1482–1490.
10. Scharla SH. Prevalence of subclinical vitamin D deficiency in different European countries. *Osteoporos Int*. 1998;8(suppl 2): S7–S12.
11. Scragg R, Sowers M, Bell C. Serum 25-hydroxyvitamin D, ethnicity, and blood pressure in the Third National Health and Nutrition Examination Survey. *Am J Hypertens*. 2007;20:713–719.
12. Shivane VK, Sarathi V, Bandgar T, Menon P, Shah NS. High prevalence of hypovitaminosis D in young healthy adults from the western part of India. *Postgrad Med J*. 2011;87(1030):514–518.
13. Hyppönen E, Turner S, Cumberland P, Power C, Gibb I. Serum 25-hydroxyvitamin D measurement in a large population survey with statistical harmonization of assay variation to an international standard. *J Clin Endocrinol Metab*. 2007;92:4615–4622.
14. Lamberg-Allardt C, Ala-Houhala M, Ahola M, Parviainen MT, Räsänen L, Viakorpi J. Vitamin D status of children and adolescents in Finland. *Ann Nutr Metab*. 1986;30:267–272.
15. Nair R, Maseeh A. Vitamin D: The “sunshine” vitamin. *J Pharmacol Pharmacother*. 2012;3:118–126.
16. Brot C, Vestergaard P, Kolthoff N, Gram J, Hermann AP, Sorensen OH. Vitamin D status and its adequacy in healthy Danish perimenopausal women: relationships to dietary intake, sun exposure and serum parathyroid hormone. *Br J Nutr*. 2001;86(suppl 1):S97–S103.
17. Davies JH, Shaw NJ. Preventable but no strategy: vitamin D deficiency in the UK. *Arch Dis Child*. 2011;96:614–615.
18. Harinarayan CV, Joshi SR. Vitamin D status in India—its implica-

- tions and remedial measures. *J Assoc Physicians India*. 2009;57:40–48.
19. Gillie O. Sunlight robbery: a critique of public health policy on vitamin D in the UK. *Mol Nutr Food Res*. 2010;54:1148–1163.
 20. Tjellesen L, Hummer L, Christiansen C, Rødbro P. Serum concentration of vitamin D metabolites during treatment with vitamin D2 and D3 in normal premenopausal women. *Bone Miner*. 1986;1:407–413.
 21. Binkley N, Gemar D, Engelke J, et al. Evaluation of ergocalciferol or cholecalciferol dosing 1,600 IU daily or 50,000 IU monthly in older adults. *J Clin Endocrinol Metab*. 2011;96:981–988.
 22. Glendenning P, Chew GT, Seymour HM, et al. Serum 25-hydroxyvitamin D levels in vitamin D-insufficient hip fracture patients after supplementation with ergocalciferol and cholecalciferol. *Bone*. 2009;45:870–875.
 23. Pepe J, Romagnoli E, Nofroni I, et al. Vitamin D status as the major factor determining the circulating levels of parathyroid hormone: a study in normal subjects. *Osteoporos Int*. 2005;16:805–812.
 24. Holick MF. Vitamin D status: measurement, interpretation, and clinical application. *Ann Epidemiol*. 2009;19:73–78.
 25. Hartwell D, Tjellesen L, Christiansen C, Rødbro P. Metabolism of vitamin D2 and vitamin D3 in patients on anticonvulsant therapy. *Acta Neurol Scand*. 1989;79:487–492.
 26. Armas LA, Hollis BW, Heaney RP. Vitamin D2 is much less effective than vitamin D3 in humans. *J Clin Endocrinol Metab*. 2004;89:5387–5391.
 27. Holmberg I, Berlin T, Ewerth S, Björkhem I. 25-Hydroxylase activity in subcellular fractions from human liver. Evidence for different rates of mitochondrial hydroxylation of vitamin D2 and D3. *Scand J Clin Lab Invest*. 1986;46:785–790.
 28. Hollis BW. Comparison of equilibrium and disequilibrium assay conditions for ergocalciferol, cholecalciferol and their major metabolites. *J Steroid Biochem*. 1984;21:81–86.
 29. Bischoff-Ferrari HA, Willet WC, Wong JB, et al. Prevention of non-vertebral fractures with oral vitamin D and dose dependency: a meta-analysis of randomized controlled trials. *Arch Intern Med*. 2009;169:551–561.
 30. Bischoff-Ferrari HA, Shao A, Dawson-Hughes B, Hathcock J, Giovannucci E, Willet WC. Benefit-risk assessment of vitamin D supplementation. *Osteoporos Int*. 2010;21:1121–1132.
 31. Nigwekar SU, Bhan I, Thadhani R. Ergocalciferol and cholecalciferol in CKD. *Am J Kidney Dis*. 2012;60:139–156.
 32. Leventis P, Kiely PD. The tolerability and biochemical effects of high-dose bolus vitamin D2 and D3 supplementation in patients with vitamin D deficiency. *Scand J Rheumatol*. 2009;38:149–153.



THE
ENDOCRINE
SOCIETY®



Members receive free electronic delivery of
FDA drug safety alerts from the PDR Network.

www.endocrine.org/FDA

3.2. Study 2

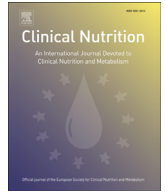
Lehmann, U., Riedel, A., Hirche, F., Brandsch, C., Girndt, M., Ulrich, C., Seibert E., Henning C., Glomb M.A., Dierkes J., & Stangl, G.I. (2016). Vitamin D3 supplementation: response and predictors of vitamin D3 metabolites—a randomized controlled trial. *Clinical nutrition*, 35(2), 351-358.



ELSEVIER

Contents lists available at ScienceDirect

Clinical Nutrition

journal homepage: <http://www.elsevier.com/locate/clnu>

Randomized control trials

Vitamin D₃ supplementation: Response and predictors of vitamin D₃ metabolites – A randomized controlled trial

Ulrike Lehmann^a, Annett Riedel^a, Frank Hirche^a, Corinna Brandsch^a, Matthias Girndt^b, Christof Ulrich^b, Eric Seibert^b, Christian Henning^c, Marcus A. Glomb^c, Jutta Dierkes^d, Gabriele I. Stangl^{a,*}

^a Institute of Agricultural and Nutritional Sciences, Martin Luther University Halle-Wittenberg, Germany

^b Department of Internal Medicine II, Martin Luther University Halle-Wittenberg, Germany

^c Institute of Chemistry, Food Chemistry, Martin Luther University Halle-Wittenberg, Germany

^d Department of Clinical Medicine, University of Bergen, Norway

ARTICLE INFO

Article history:

Received 6 October 2014

Accepted 30 April 2015

Keywords:

Vitamin D

25(OH)D₃

24,25(OH)₂D₃

Supplementation

Healthy subjects

Randomized controlled trial

SUMMARY

Background & aims: Large parts of the population are insufficiently supplied with vitamin D, in particular when endogenous synthesis is absent. Therefore many health care providers recommend the use of vitamin D supplements. The current study aimed to investigate the efficacy of an once-daily oral dose of 20 µg vitamin D₃ to improve the vitamin D status and to evaluate predictors of response.

Methods: The study was conducted as a double-blind, randomized, placebo-controlled parallel trial from January till April 2013. In total, 105 subjects (20–71 years) were allocated to receive either a vitamin D₃ supplement (20 µg/d) or a placebo for 12 weeks. Circulating levels of vitamin D₃ metabolites such as the 25(OH)D₃ and the 24,25(OH)₂D₃, and biomarkers of calcium and phosphate metabolism were quantified. **Results:** The 25(OH)D₃ serum concentrations in the placebo group decreased from 38 ± 15 nmol/L at baseline to 32 ± 14 nmol/L and 32 ± 13 nmol/L at weeks 8 and 12 of the study, respectively ($p < 0.01$). In the vitamin D₃ group, the serum 25(OH)D₃ concentration increased from 38 ± 14 nmol/L at baseline to 70 ± 15 nmol/L and 73 ± 16 nmol/L at weeks 8 and 12 of vitamin D₃ supplementation ($p < 0.001$), respectively. As a result, 94% of the vitamin D₃-supplemented participants reached 25(OH)D₃ concentrations of ≥50 nmol/L and thereof 46% attained 25(OH)D₃ levels of ≥75 nmol/L until the end of the study. The extent of the 25(OH)D₃ increase upon vitamin D₃ supplementation depended on 25(OH)D₃ baseline levels, age, body weight and circulating levels of triglycerides. In contrast to 25(OH)D₃, the response of 24,25(OH)₂D₃ to the vitamin D₃ treatment was affected only by baseline levels of 24,25(OH)₂D₃ and age.

Conclusions: The average improvement of 25(OH)D₃ levels in individuals who received 20 µg vitamin D₃ per day during the winter months was 41 nmol/L compared to individuals without supplementation. As a result almost all participants with the vitamin D₃ supplementation attained 25(OH)D₃ concentrations of 50 nmol/L and higher. The suitability of 24,25(OH)₂D₃ as a marker of vitamin D status needs further investigation.

Clinical trial registration number at clinicaltrials.gov: NCT01711905.

© 2015 Elsevier Ltd and European Society for Clinical Nutrition and Metabolism. All rights reserved.

Abbreviations: BIA, bioelectrical impedance analysis; BMI, body mass index; FGF-23, fibroblast growth factor 23; LC–MS/MS, liquid chromatography coupled with mass spectrometry; LLOQ, lower limit of quantification; 1,25(OH)₂D, 1,25-dihydroxyvitamin D; 24,25(OH)₂D₃, 24,25-dihydroxyvitamin D₃; 25(OH)D₃, 25-hydroxyvitamin D₃.

* Corresponding author. Institute of Agricultural and Nutritional Sciences, Von-Danckelmann-Platz 2, D-06120 Halle/Saale, Germany. Tel.: +49 345 55 22707; fax: +49 345 55 27124.

E-mail address: gabriele.stangl@landw.uni-halle.de (G.I. Stangl).

1. Introduction

There is an ongoing debate on the necessity of vitamin D supplementation in the healthy population. Assessment of vitamin D status is currently based on measuring circulating 25-hydroxyvitamin D₃ (25(OH)D₃), which is considered as reliable biomarker of the vitamin D status. The classification of 25(OH)D concentrations into deficient, insufficient and adequate is mainly

based on the onset of bone ossification disorders [1]. Although it is generally agreed that serum 25(OH)D concentrations below 30 nmol/L are deficient, the optimum level of 25(OH)D remains controversial. The Institute of Medicine (IOM) classified 25(OH)D concentrations of 50 nmol/L as sufficient to prevent bone disorders [1]. Studies that considered further vitamin D associated diseases such as cancer and cardiovascular diseases, recommend 25(OH)D levels of at least 75 nmol/L [2,3]. Heaney proposed that the vitamin D status to ensure normal cell function had to be higher than that necessary to prevent diseases [4].

Recent data from a random sub-cohort of the German arm of the European Prospective Investigation into Cancer and Nutrition (EPIC) demonstrated pronounced seasonal variations in 25(OH)D₃ concentrations and showed that, from January to March, 82.2% of the subjects had 25(OH)D₃ serum concentrations <50 nmol/L and 34.1% were even below 30 nmol/L [5]. With regard to the high prevalence of vitamin D insufficiency, the German Nutrition Society recommends a daily vitamin D intake of 20 µg for healthy adults in periods of limited endogenous vitamin D synthesis, to achieve serum 25(OH)D₃ concentrations of at least 50 nmol/L [6]. Previous work of Cashman and co-workers clearly showed that the amount of dietary vitamin D necessary to maintain the 25(OH)D concentrations during wintertime depends on the extent of summer sunshine exposure, diet intake of vitamin D and the desired levels of circulating 25(OH)D [7]. Data of that study pointed out that there is a substantial heterogeneity in the individual response of 25(OH)D to vitamin D supplementation [7], although other factors that may explain that phenomenon have not been entirely clarified.

The baseline 25(OH)D level is assumed to be an important determinant (e.g. 8–10), but also genetic variances (e.g. 9, 10), body fat and/or body mass index (BMI) (e.g. 8, 11) are suggested to influence the response. In vitamin D studies, little attention has been paid to age, since most investigations were dedicated to a specific age group, e.g. premenopausal women (19–35 years) [12], postmenopausal women [11] or elderly (≥60 years) [8,10].

The present study aimed: (i) to investigate the efficacy of a daily oral dosage of 20 µg vitamin D₃ to increase the 25(OH)D₃ serum concentration of healthy volunteers to at least 50 nmol/L during the winter months, (ii) to identify factors that may modify the response of 25(OH)D₃ to vitamin D₃ supplementation, and (iii) to validate the suitability of 24,25-dihydroxyvitamin D₃ (24,25(OH)₂D₃) as biomarker for vitamin D supplementation. To this end, females and males (≥18 years of age) without restrictions regarding BMI were recruited to participate in the study and serum concentrations of 25(OH)D₃, 1,25-dihydroxyvitamin D (1,25(OH)₂D), 24,25(OH)₂D₃ and biomarkers of calcium and phosphate homeostasis were quantified.

2. Materials and methods

2.1. Study design and study population

This study was conducted as a double-blind, randomized, controlled human intervention trial in parallel groups. The aims of the study were to assess the effect of an 8 and 12 week recommendation-based supplementation of 20 µg/d vitamin D₃ versus a placebo on the serum concentrations of 25(OH)D₃ and 24,25(OH)₂D₃, and to identify factors that modify the response of these metabolites upon vitamin D₃ supplementation. The study was performed from January 2013 until the beginning of April 2013 in Halle/Saale, Germany (51° of Northern latitude), when endogenous vitamin D synthesis via UVB irradiation was absent. The study was approved by the local ethics committee of the Medical Faculty at the Martin Luther University Halle-Wittenberg. Each participant

gave written informed consent prior to the beginning of the study. The study was registered at clinicaltrials.gov (NCT01711905).

Sample size was calculated using G*Power analysis program [13] to find statistically significant differences in the serum vitamin D metabolite response between predictor-based subgroups. Considering one-way ANOVA as statistical test, a mean 25(OH)D₃ difference of 10 nmol/L between subgroups and a standard deviation of 14 nmol/L, a power of 95% and a significance level of 0.05, a total sample size of 51 subjects (n = 17 in each subgroup) was required.

Apparently healthy subjects of both sexes from a minimum age of 18 years were recruited in cooperation with the blood donation center of the Department of Transfusion Medicine (University Hospital Halle, Germany), through newspapers advertisements, personal contacts and public information events. All volunteers underwent a screening visit in autumn 2012. They had to fill in a questionnaire on medical history, body weight and height, lifestyle behaviors (e.g. smoking, usage of sun blocker containing cosmetics) and dietary habits especially focusing on the intake of foods rich in vitamin D. Blood samples were drawn to estimate the serum 25(OH)D₃ concentrations. The overall health status was assessed by questionnaires on disease history, clinical measurements (e.g. blood pressure) and clinical chemical analysis of markers of renal function, thyroid function and calcium metabolism. Exclusion criteria were use of dietary supplements, vacations in regions with abundant UVB irradiation 8 or less weeks prior to the study begin or during the study period, serum 25(OH)D₃ concentrations >75 nmol/L, pregnancy or lactation, participation in other clinical studies, intake of prescription medication, deviations in blood count or metabolic disorders.

A total of 106 subjects (age range: 20–71 years) were included in the study. They were randomly assigned into the two groups (placebo group, n = 52, vitamin D₃ group, n = 54) by block randomization using a computer-generated randomization schedule with serum 25(OH)D₃ concentration, BMI and sex as stratification criteria. One female participant from the placebo group dropped out because of personal reasons. Therefore, 105 participants completed the study and were included into the statistical analysis.

Study visits were scheduled at baseline, and after 8 and 12 weeks of intervention. At each study visit anthropometric data (height, body weight, waist circumference, body fat mass), blood pressure and heart frequency were recorded, fasting blood samples were drawn and participants were asked for adverse effects in response to the treatment. Body fat mass was determined by bioelectrical impedance analysis (BIA, Data Input, Darmstadt, Germany). After 5 min of rest, blood pressure and heart rate were measured in triplicate at the left arm at heart level with a one minute interval in between (BpTRU Medical Devices, Coquitlam, Canada).

Prior to the start of the study, the participants received their pre-packed and numbered (according to the randomization schedule) 12-week ration of either the placebo or the vitamin D₃ supplement. The participants were instructed to take one tablet per day for a total of 12 weeks. The time of consumption was not specified but it was recommended to take the tablet together with a meal. Compliance was ensured by counting the remaining tablets, and achieved 99%. The supplements were manufactured by Vital Products GmbH (Waldsassen, Germany). Vitamin D₃ was received from DSM Nutritional Products Ltd. (Basel, Switzerland). Placebo and vitamin D₃ tablets were outwardly indistinguishable in appearance and taste. Cellulose was used as the placebo. The vitamin D₃ content per tablet was quantified by liquid chromatography coupled with mass spectrometry (LC–MS/MS) in four separate runs [14]. The analyzed vitamin D₃ content of a single vitamin D₃ tablet amounted to 19.6 ± 1.5 µg. An independent

investigator numbered the vitamin D₃ tablet containers, so that all researchers and staff who conducted the study and analyzed the samples and all study participants were unaware of the group assignment. Treatment codes were generated in blocks of two by using a computer-generated randomization schedule. Subjects were enrolled by a physician involved in the trial, but assigned to the intervention by another investigator.

The study was completed as scheduled. After completion of the study, information on the individual vitamin D status and strategies for a vitamin D status improvement were provided to all subjects, and individuals of the placebo group were offered vitamin D supplements to improve the vitamin D status.

2.2. Analytical methods

For biochemical analyses venous blood was collected and centrifuged at $2000 \times g$ for 10 min to obtain serum and EDTA-plasma samples. For the quantification of glucose, fluoride-coated tubes were used. Samples were aliquoted, frozen and stored at -80°C until analyses.

Serum 25(OH)D₃ concentrations at baseline and after 8 and 12 weeks of intervention were quantified by means of a MassChrom[®] 25-OH Vitamin D₃ reagent kit (Chromsystems GmbH, Munich, Germany) for LC–MS/MS using an API 2000™ system (Applied Biosystems, Darmstadt, Germany) as described elsewhere [15]. The lower limit of quantification (LLOQ) was 10.7 nmol/L and the coefficients of variance were 8.3% ($n = 4$, inter assay) and 5.0% ($n = 3$, intra assay) at 39.9 nmol/L. Serum 1,25(OH)₂D concentrations were determined using a commercially available ELISA kit (Immunodiagnostic Systems GmbH, Frankfurt am Main, Germany) at baseline and after 12 weeks of intervention.

Serum 24,25(OH)₂D₃ concentration at baseline and week 12 was determined by LC–MS/MS (PU-2080 Plus, LG-2080-02, LG-2080-04, Jetstream II, AS-2057 Plus, all Jasco, Gross-Umstadt, Germany; 4000 QTrap system, Applied Biosystems) using a Hypersil ODS column, $150 \times 2 \text{ mm}^2$, $5 \mu\text{m}$ (VDS optilab, Berlin, Germany) at 40°C and 0.576 mL/min. The mobile phase consisted of (A) 5 mM ammonium formate, 0.1% formic acid in water/acetonitrile (9 + 1, v/v), and (B) acetonitrile (time table: 0–1.9 min 50% B; 5.5 min 56% B; 5.7 min 100% B; 11.8 min 100% B; 12–15 min 50% B). Calculations were based on (m/z) 574.6/298.4 for 24,25(OH)₂D₃ and 582.5/298.4 for 25(OH)D₃-d₆ (Chemaphor Inc., Ottawa, Canada) which was used as internal standard. The coefficients of variance were 11.3% ($n = 6$, inter assay) and 7.5% ($n = 3$, intra assay) at 2.11 nmol/L. The LLOQ was 0.26 nmol/L.

Calcium and inorganic phosphate serum concentrations were determined using spectrophotometric assays (Analyticon Biotechnologies AG, Lichtenfels, Germany). Commercial ELISA kits were used to quantify serum concentrations of intact parathyroid hormone (PTH, Biomerica Inc., Irvine, USA) and soluble α -Klotho (IBL Immuno-Biological Laboratories Co., Ltd., Japan) as well as plasma fibroblast growth factor 23 (FGF-23; C-term, Immotopics Inc., San Clemente, USA). Plasma glucose and serum triglycerides concentrations were measured using enzymatic assays (DiaSys Diagnostic Systems GmbH, Holzheim, Germany). All analyses were run in duplicate following the manufacturer's instructions.

2.3. Statistical analyses

Statistical analyses were performed using SPSS Version 22.0 (IBM, Chicago, USA) and SigmaPlot (Systat Software, Inc., San Jose, USA). Data are presented as mean \pm SD. The significance level was set at 5%. Normal distribution of the data was verified by the Kolmogorov–Smirnov test, skewed variables (24,25(OH)₂D₃, PTH, FGF-23, Klotho) were logarithmically transformed. Differences between

groups at baseline were analyzed by a two sample t-test. To assess the effects of the factors time (within-subjects factor) and treatment (between-subjects factor) as well as their interaction a two-way ANOVA with Bonferroni post hoc test was conducted. The number of volunteers was $n = 51$ in the placebo group, and $n = 54$ in the vitamin D₃ group.

To identify correlations between plasma vitamin D metabolite concentrations Spearman's correlation coefficients were calculated. Therefore, coefficients between circulating concentrations of 25(OH)D₃, 1,25(OH)₂D and 24,25(OH)₂D₃ were calculated by considering data from all subjects at baseline and after 12 weeks. To elucidate the impact of an improved vitamin D status on health parameters, correlations between changes (baseline to week 12) of vitamin D metabolites and anthropometric and clinical markers were assessed.

In order to compile a prediction model of the vitamin D₃ supplementation response, a regression analysis was performed. Best subset regression based on the adjusted R² and subsequent multivariate linear regression analyses were accomplished to identify predictors for the changes (baseline to week 12) of serum 25(OH)D₃ and 24,25(OH)₂D₃ upon vitamin D₃ supplementation representing the dependent variables. For those parameters that significantly affected changes in serum 25(OH)D₃ and 24,25(OH)₂D₃, a subgroup analysis was conducted. The subjects were divided into tertiles based on the baseline values of the predictors and a one-way ANOVA with Bonferroni post hoc test was performed to estimate differences between the subgroups.

3. Results

3.1. Baseline characteristics of the subjects and vitamin D status

Characteristics of the subjects at study entry are presented in Table 1. The study collective comprised of 67% females and 33% males. The mean age and BMI at baseline were 39 years and 24.0 kg/m², respectively. Anthropometric data, blood pressure, heart rate and concentrations of glucose and triglycerides at baseline did not differ between the two groups. None of the participants were vegan or vegetarian.

Serum concentrations of the vitamin D metabolites at the beginning of the study were also not different between the two groups (Table 2). At baseline, 32% of the individuals had 25(OH)D₃ concentrations lower than 30 nmol/L, 48% had concentrations between 30 and 50 nmol/L, and 20% had concentrations that ranged between 50 and 75 nmol/L. During the study, the serum level of 25(OH)D₃ from individuals of the placebo group decreased significantly from baseline to week 8 ($p < 0.01$), but remained unchanged from week 8 to week 12 (Table 2). In the placebo group the decrease of 25(OH)D₃ within the 12 weeks during winter was on average 6 nmol/L. In the vitamin D₃ group, the serum levels of 25(OH)D₃ increased from baseline to week 8 ($p < 0.001$), without showing any further increase from week 8 to week 12. The 25(OH)D₃ levels could be improved on average by about 35 nmol/L during the 12 week intervention. After 8 and 12 weeks of treatment, individuals from the vitamin D₃ group showed 2.2- and 2.3-fold higher 25(OH)D₃ levels than those of the placebo group. After 12 weeks, 94% of the participants that received vitamin D₃ reached 25(OH)D₃ concentrations higher than 50 nmol/L and 46% even reached concentrations higher than 75 nmol/L. In the placebo group, 53% had 25(OH)D₃ concentrations below 30 nmol/L, 33% ranged between 30 and 50 nmol/L and 14% had concentrations above 50 nmol/L. None of them showed 25(OH)D₃ concentrations higher than 75 nmol/L.

Due to the non-significant changes of 25(OH)D₃ between week 8 and 12 in both groups, the serum levels of 1,25(OH)₂D and 24,25(OH)₂D were analyzed only at baseline and after 12 weeks of

Table 1
Characteristics of study participants at baseline.

	Placebo group (n = 51)	Vitamin D ₃ group (n = 54)	Range (n = 105)
Sex [male/female]	17/34	18/36	
Age [years]	39 ± 14	39 ± 14	20–71
Body weight [kg]	70 ± 14	72 ± 11	49–107
Body mass index [kg/m ²]	24 ± 3	24 ± 3	18–31
Body fat mass (BIA) [%]	19 ± 6	20 ± 7	10–43
Waist circumference [cm]	80 ± 11	83 ± 10	62–106
Blood pressure [mmHg]			
Systolic	114 ± 13	113 ± 13	93–148
Diastolic	75 ± 9	74 ± 9	54–102
Heart rate [beats/min]	69 ± 11	69 ± 9	38–94
Glucose [mmol/L]	4.9 ± 0.6	4.8 ± 0.8	3.4–8.3
Triglycerides [mmol/L]	1.0 ± 0.6	0.9 ± 0.5	0.3–3.4

Values are given as mean ± SD.

intervention. Analysis revealed that the serum levels of 1,25(OH)₂D were altered by the treatment but to a lesser extent than 25(OH)D₃ (Table 2). Serum concentrations of 1,25(OH)₂D slightly decreased from baseline to week 12 in the placebo group ($p < 0.05$), whereas no changes were observed in the vitamin D₃ group. After 12 weeks, subjects of the vitamin D₃ group had higher serum levels of 1,25(OH)₂D than subjects of the placebo group ($p < 0.001$).

As observed for 25(OH)D₃, the 24,25(OH)₂D₃ serum concentration was significantly influenced by treatment and time (Table 2). In the placebo group, the serum concentration of 24,25(OH)₂D₃ decreased from baseline to week 12 ($p < 0.001$), whereas it increased in the vitamin D₃ group ($p < 0.001$). After 12 weeks of treatment, subjects of the vitamin D₃ group had nearly 3-fold

higher 24,25(OH)₂D₃ concentrations than those of the placebo group.

3.2. Biomarkers of calcium and phosphate metabolism

Data demonstrated no differences in baseline concentrations of calcium, phosphate, PTH, klotho and FGF-23 between the two groups (Table 2). Serum concentration of calcium was influenced by time, but not by the treatment; there was a decline from baseline to week 12 in both groups (Table 2). The circulating concentration of inorganic phosphate was neither affected by treatment nor by time. The serum concentration of PTH increased from baseline to week 12 in the placebo group ($p < 0.05$), whereas it remained unchanged

Table 2
Concentrations of vitamin D metabolites and parameters of calcium and phosphate homeostasis at baseline as well as after 8 and/or 12 weeks of intervention.

	Placebo group	Vitamin D ₃ group	Two-way ANOVA (p -value)		
			Time	Treatment	Time × Treatment
25(OH)D₃ [nmol/L]					
Baseline	38 ± 15 ^a	38 ± 14 ^a	<0.001	<0.001	<0.001
8. week	32 ± 14 ^b	70 ± 15 ^{b*}			
12. week	32 ± 13 ^b	73 ± 16 ^{b*}			
1,25(OH)₂D [pmol/L]					
Baseline	110 ± 37 ^a	119 ± 44	0.66	<0.005	<0.001
12. week	96 ± 34 ^b	130 ± 35 [*]			
24,25(OH)₂D₃ [nmol/L]					
Baseline	1.9 ± 1.1 ^a	1.8 ± 0.9 ^a	<0.05	<0.001	<0.001
12. week	1.2 ± 0.8 ^b	3.4 ± 1.2 ^{b*}			
Calcium [mmol/L]					
Baseline	2.4 ± 0.1 ^a	2.4 ± 0.1 ^a	<0.001	0.20	<0.05
8. week	2.3 ± 0.1 ^a	2.3 ± 0.1 ^b			
12. week	2.2 ± 0.1 ^b	2.3 ± 0.1 ^b			
Inorganic phosphate [mmol/L]					
Baseline	1.2 ± 0.2	1.2 ± 0.2	0.35	0.17	<0.05
8. week	1.2 ± 0.2	1.2 ± 0.2			
12. week	1.2 ± 0.2	1.2 ± 0.2			
PTH [pmol/L]					
Baseline	6.5 ± 2.1 ^a	6.4 ± 2.5	<0.05	0.08	<0.001
8. week	7.0 ± 2.3 ^{a,b}	6.2 ± 1.9			
12. week	7.4 ± 2.6 ^b	6.1 ± 2.3			
Klotho [pg/mL]					
Baseline	821 ± 738	753 ± 459	0.08	0.82	0.93
8. week	845 ± 795	738 ± 419			
12. week	807 ± 795	716 ± 414			
FGF-23 [RU/mL]					
Baseline	95 ± 53	96 ± 53	0.16	0.91	0.96
8. week	93 ± 58	85 ± 35			
12. week	100 ± 72	96 ± 61			

Data are given as mean ± SD. Bonferroni post hoc test was applied.

^{a,b}Significant differences between time points within a group ($p < 0.05$).

*Significantly different from placebo group at a given time ($p < 0.05$).

in the vitamin D₃ group. The circulating concentrations of klotho and FGF-23 were not affected by treatment and time, respectively.

3.3. Data from correlation analyses

Correlation analysis of vitamin D metabolite serum concentrations from all subjects at baseline and after 12 weeks of treatment are presented in Fig. 1. Data revealed a strong positive association between the serum concentrations of 25(OH)D₃ and 24,25(OH)₂D₃ (Fig. 1A). The correlation between 25(OH)D₃ and 1,25(OH)₂D₃ was weaker than that between 25(OH)D₃ and 24,25(OH)₂D₃ (Fig. 1B). Similar correlation data were observed when taking the changes (from baseline to week 12) of the vitamin D metabolites as a basis (Fig. 1C and D).

Correlation analysis between changes of circulating vitamin D metabolites and changes of the other analyzed parameter is presented in Table 3. Data showed a weak positive correlation between Δ 24,25(OH)₂D₃ and Δ waist circumference and a weak positive correlation between Δ 25(OH)D₃ and Δ calcium. In contrast, all vitamin D metabolites changes were inversely correlated with Δ PTH, in the order of magnitude from the strongest to weakest correlation coefficients: 25(OH)D₃ > 24,25(OH)₂D₃ > 1,25(OH)₂D₃.

3.4. Predictors of serum 25(OH)D₃ and 24,25(OH)₂D₃ response to vitamin D₃ supplementation

Best subset regression analyses were performed to identify factors that modify the response of 25(OH)D₃ and 24,25(OH)₂D₃ serum concentrations to vitamin D₃ supplementation (Table 4). Choice of predictors was based on the adjusted R² that considers

the number of independent variables and prevents an overfitting in regression-type models.

The variance in 25(OH)D₃ changes explained by the regression model was 43% (overall *p*-value of the regression < 0.001, R² = 0.48, adjusted R² = 0.43). There was no multi-collinearity between the determinants. The prediction model revealed the baseline 25(OH)D₃ concentration as the strongest predictor for Δ 25(OH)D₃, followed by age, body weight and baseline serum triglycerides. For Δ 24,25(OH)₂D₃, about 17% of the overall variability were explained by the regression model (overall *p*-value of the regression < 0.001, R² = 0.20, adjusted R² = 0.17). Determinants of the 24,25(OH)₂D₃ response were baseline 24,25(OH)₂D₃ concentration and age.

Results from the subgroup analyses are shown in Table 5. The Δ 25(OH)D₃ decreased significantly with increasing levels of baseline 25(OH)D₃ and with increasing age. Although body weight and baseline triglyceride concentrations were shown to affect Δ 25(OH)D₃, significant differences between subgroups were not demonstrated. Analyses of Δ 24,25(OH)₂D₃ revealed only a trend toward decreasing changes with increasing levels of baseline 24,25(OH)₂D₃ and increasing age.

4. Discussion

This study aimed to investigate the efficacy of 20 μ g vitamin D₃ per day to improve the vitamin D status of individuals during the winter months by analyzing changes of serum 25(OH)D₃ concentrations. Taking the recommended 25(OH)D level of the IOM as a basis (\geq 50 nmol/L) [1], 32% of the study participants had a deficient and 48% had an insufficient vitamin D status at baseline. Virtually all participants who were treated with 20 μ g vitamin D₃ per day reached serum 25(OH)D levels of at least 50 nmol/L. When taking

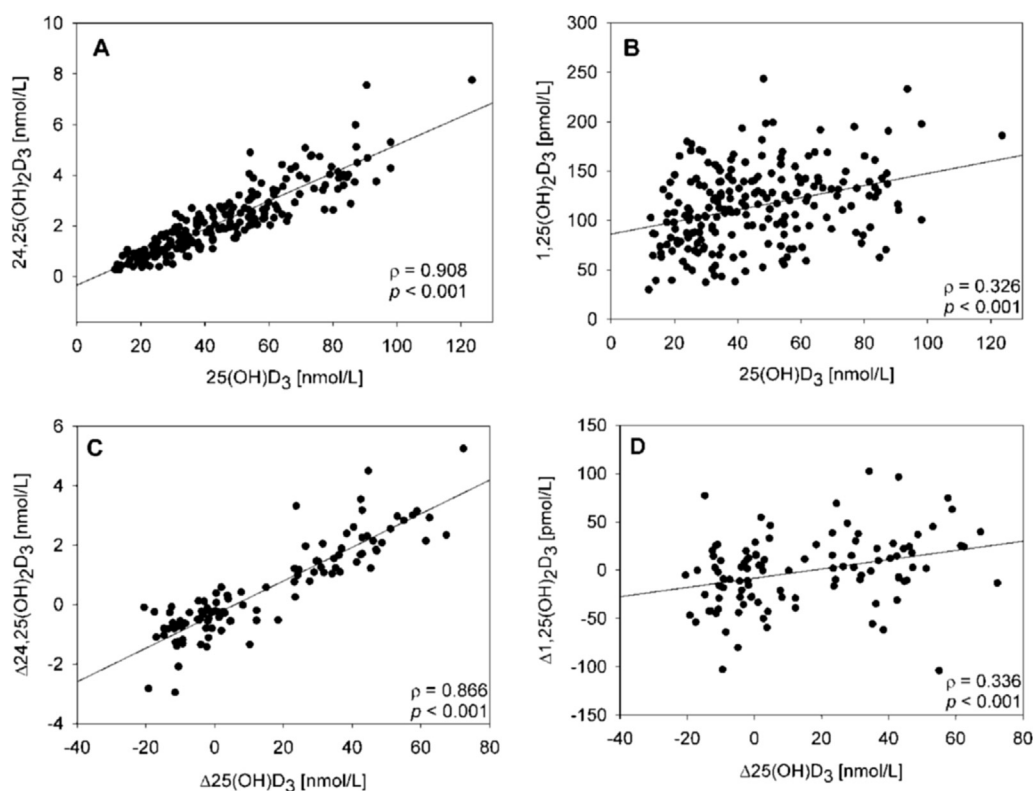


Fig. 1. Spearman's correlation between 25(OH)D₃ and (A) 24,25(OH)₂D₃ and (B) 1,25(OH)₂D₃ serum concentrations comprising data at baseline and after 12 weeks of treatment from both groups (vitamin D₃ and placebo) (n = 210); correlations between Δ 25(OH)D₃ and (C) Δ 24,25(OH)₂D₃ and (D) Δ 1,25(OH)₂D₃ including changes from baseline to week 12 of treatment from both groups (n = 105).

Table 3
Spearman's correlation analysis of changes in parameters in the vitamin D₃ group.

	Correlation coefficients		
	Δ25(OH)D ₃ [nmol/L]	Δ1,25(OH) ₂ D [pmol/L]	Δ24,25(OH) ₂ D ₃ [nmol/L]
ΔBody mass index [kg/m ²]	0.09	0.07	0.10
ΔBody fat mass (BIA)[%]	-0.12	-0.13	-0.07
ΔWaist circumference [cm]	0.12	-0.08	0.20*
ΔGlucose [mmol/L]	-0.06	-0.01	0.02
ΔTriglycerides [mmol/L]	-0.10	-0.14	-0.09
ΔCalcium [mmol/L]	0.21*	-0.04	0.18
ΔInorganic phosphate [mmol/L]	0.18	0.08	0.15
ΔParathyroid hormone [pmol/L]	-0.42***	-0.24*	-0.40***
ΔKlotho [pg/mL]	-0.01	0.07	-0.04
ΔFGF-23 [RU/mL]	-0.07	-0.06	-0.03

p* < 0.05, **p* < 0.001.**Table 4**
Identified predictors for supplementation response assessed by best subset regression analysis.

Predictors	Coefficient	<i>p</i> -value
Δ25(OH)D₃		
Constant	97 ± 14	<0.001
Baseline 25(OH)D ₃ [nmol/L]	-0.67 ± 0.14	<0.001
Age [years]	-0.38 ± 0.14	0.01
Body weight [kg]	-0.43 ± 0.18	<0.05
Baseline triglycerides [mmol/L]	8.8 ± 3.9	<0.05
Δ24,25(OH)₂D₃		
Constant	3.5 ± 0.6	<0.001
Baseline 24,25(OH) ₂ D ₃ [nmol/L]	-0.50 ± 0.17	<0.01
Age [years]	-0.02 ± 0.01	<0.05

Coefficients are given as mean ± standard error.

Δ25(OH)D₃: *p*-value of the regression < 0.001, R = 0.69, R² = 0.48, adjusted R² = 0.43; Δ24,25(OH)₂D₃: *p*-value of the regression < 0.001, R = 0.45, R² = 0.20, adjusted R² = 0.17.

75 nmol/L as recommended cut-off level [16], only half of the vitamin D₃ treated participants in our study attained the required 25(OH)D concentrations. Systematic review data demonstrated that the 25(OH)D concentrations to prevent falls, cancer and respiratory infection should be at least 95 nmol/L, 100 nmol/L, and 95 nmol/L, respectively [3]. We conclude that the currently used vitamin D₃ dosage is capable of attaining the recommendations of IOM and the German Nutrition Society but not to reach levels of ≥75 nmol/L. In line with previous findings [17], extended times of administered vitamin D₃ appear not to improve 25(OH)D₃ levels

since changes in serum 25(OH)D₃ concentrations between the weeks 8 and 12 were minimal and non-significant.

Since 25(OH)D₃ response to vitamin D₃ showed a great inter-individual variability (Δ25(OH)D ranging from -13 to 72 nmol/L), we tested whether 24,25(OH)₂D₃ could provide a more robust biomarker to assess vitamin D status. 24,25(OH)₂D₃ is formed from 25(OH)D₃ and 1,25(OH)₂D₃ by the action of CYP24A1 and is proposed to be an inactive vitamin D metabolite that is destined for excretion [18]. Most studies that aimed to investigate the vitamin D status did not analyze 24,25(OH)₂D₃. Cashman et al. were the first who demonstrated the response of 24,25(OH)₂D₃ to 20 μg vitamin D₃, although modifying factors of 24,25(OH)₂D₃ and associations to 25(OH)D were not analyzed [19]. Current data show that 24,25(OH)₂D₃ levels highly correlated with the 25(OH)D₃ concentration, and also the magnitude of 24,25(OH)₂D₃ increase in response to vitamin D₃ supplementation closely resembles to that of 25(OH)D₃. So far, an association between 24,25(OH)₂D₃ and 25(OH)D has only been demonstrated in studies that administered extremely high vitamin D doses of 600,000 IU as bolus [20] or 28,000 IU once per week over 8 weeks [21].

To identify determinants that explain the individual differences in the response to the vitamin D₃ supplementation, we applied a best subset regression model and conducted a subgroup analysis. Among the comprised factors, baseline 25(OH)D₃ concentrations, age, body weight and triglyceride concentrations were identified to affect the efficacy of 25(OH)D₃ response to vitamin D₃ intake. The finding, that changes in 25(OH)D₃ following vitamin D₃ treatment were inversely associated with the baseline levels of 25(OH)D₃ is in

Table 5
Subgroup analysis of vitamin D₃ supplementation response (Δ25(OH)D₃ and Δ24,25(OH)₂D₃) according to significant predictors.

	1st tertile	2nd tertile	3rd tertile	One-way ANOVA <i>p</i> -value
Δ25(OH)D₃				
Baseline 25(OH)D ₃ [nmol/L]	≤31	>31–44	>44	<0.01
Δ25(OH)D ₃ [nmol/L]	44 ± 13 ^a	35 ± 15 ^{a,b}	25 ± 20 ^b	
Age [years]	≤29	>29–45	>45	<0.05
Δ25(OH)D ₃ [nmol/L]	43 ± 16 ^a	34 ± 17 ^{a,b}	27 ± 18 ^b	
Body weight [kg]	≤65	>65–76	>76	0.07
Δ25(OH)D ₃ [nmol/L]	42 ± 18	30 ± 18	31 ± 14	
Triglycerides [mmol/L]	≤0.8	>0.8–1.0	>1.0	0.13
Δ25(OH)D ₃ [nmol/L]	31 ± 13	32 ± 23	40 ± 15	
Δ24,25(OH)₂D₃				
Baseline 24,25(OH) ₂ D ₃ [nmol/L]	≤1.3	>1.3–2.1	>2.1	0.05
Δ24,25(OH) ₂ D ₃ [nmol/L]	1.8 ± 0.8	1.8 ± 1.1	1.0 ± 1.6	
Age [years]	≤29	>29–45	>45	0.09
Δ24,25(OH) ₂ D ₃ [nmol/L]	2.0 ± 1.1	1.4 ± 1.3	1.2 ± 1.2	

Data are given as mean ± SD.

^{a,b}Significantly different between subgroups (Bonferroni post hoc test).

accordance with other studies [8–10,12] and is suggested to be caused by a negative feedback of 25-hydroxylase activity [22].

Age was identified as the second important predictor that independently influenced the response of 25(OH)D₃ to vitamin D₃ supplementation. We observed an inverse association between age and changes in 25(OH)D₃ concentration to vitamin D₃. Subjects of the lowest age tertile (≤ 29 years) appeared to be more responsive to supplemented vitamin D₃ than those of the highest age tertile (>45 years). Most of the previous vitamin D studies that found age as non-modifying factor of the 25(OH)D response addressed only a specific age group [10,23]. A recently published meta-regression found age positively associated with the 25(OH)D₃ response [24]. The reason for the contradictory finding in the meta-regression and our study could be the mean age of the volunteers which were on average 66 years (subgroups: <69 , 70–79 and >80 years) in the meta-regression study, and 39 years (subgroups: ≤ 29 , 29–45 and >45 years) in the current study.

Body weight which was negatively associated with the 25(OH)D₃ response represented a further predictor of the vitamin D₃ supplementation efficacy. Numerous studies found lower 25(OH)D₃ concentrations in overweight/obese subjects than in lean subjects (e.g. [25,26]), and postulated an inverse relation between BMI and changes of 25(OH)D₃ [8–10]. The authors attribute this observation to the differences in body fat mass, as vitamin D is stored in the adipose tissue and hence be less available for hydroxylation [8,27]. Body fat mass and BMI were also included to our regression model, but both were not associated with the $\Delta 25(\text{OH})\text{D}_3$. We speculate that total body mass rather than fat mass may modulate the 25(OH)D response. Although serum triglycerides contribute to improve the prediction model for $\Delta 25(\text{OH})\text{D}_3$ in response to vitamin D₃ treatment, there were no pronounced differences between subgroups. Nevertheless, triglyceride levels should be given more attention in future studies on vitamin D.

Based on the best subset regression analysis, we provided first evidence that the response of 24,25(OH)₂D₃ to vitamin D₃ administration was also modified by individual factors, although less than that of 25(OH)D₃. The current study found $\Delta 24,25(\text{OH})_2\text{D}_3$ to be affected by baseline 24,25(OH)₂D₃ concentrations and age, but not by body weight, triglycerides or baseline 25(OH)D₃ concentrations as demonstrated for 25(OH)D₃. Thus, serum 24,25(OH)₂D₃ could possibly provide a more robust marker of vitamin D status than 25(OH)D₃.

We further analyzed PTH as sensitive marker of serum calcium dysbalance, and found an increase of PTH concentration in placebo-treated subjects from baseline to week 12, but no changes in the group supplemented with vitamin D₃. The absence of the PTH response to vitamin D₃ supplementation was an unexpected result since 80% of the participants in this group showed baseline 25(OH)D₃ levels lower than 50 nmol/L and improved their vitamin D₃ status by vitamin D₃ treatment. Since PTH of young adults is known to respond less pronounced than that of older adults [28], we assume that the young age of individuals included in our study could explain the lack of PTH response.

Another unexpected finding was the decrease of serum calcium from baseline to week 12 in both study groups. We fail to explain this phenomenon, but we exclude calcium intake as a causal factor, as data from the Tromsø study showed no association between intake and serum levels of calcium [29]. Moreover, the current data provide no hint for an impact of vitamin D₃ supplementation on FGF-23 and klotho that are both linked to regulation of phosphate homeostasis [30].

However, data interpretation is somewhat restricted due to some limitations of our study. We neither have analyzed activities of enzymes involved in the conversion of vitamin D metabolites nor genetic polymorphisms of hydroxylating enzymes (CYP27B1,

CYP2R1, CYP24A1) and vitamin D binding protein, which are known to modulate the 25(OH)D₃ response to vitamin D₃. The intake of minerals and vitamin D which are determinants of vitamin D status and metabolites were not assessed.

In conclusion, the administration of 20 μg vitamin D₃ per day is suitable to improve deficient or insufficient concentrations of 25(OH)D₃ to at least 50 nmol/L during the winter months. The efficacy to increase 25(OH)D₃ serum levels depends on 25(OH)D₃ levels at baseline, age, body weight and circulating triglycerides. The 24,25(OH)₂D₃ concentration highly correlates with the 25(OH)D₃ concentration and appears to be less susceptible to modifying factors. However, further investigations are required to validate 24,25(OH)₂D₃ as biomarker of vitamin D status.

Statement of authorship

UL conducted the study, processed the samples, contributed to the analyses of 25(OH)D₃, FGF-23, PTH, 1,25(OH)₂D, calcium, glucose and triglycerides, reviewed the literature and drafted the manuscript. AR performed the statistical analyses, reviewed the literature and helped to draft the manuscript. FH analyzed 25(OH)D₃, FGF-23 and PTH. CB analyzed klotho, and contributed to measurement and evaluation of plasma parameters and anthropometric data. MG participated in the design of the study. CU conducted blood sampling, controlled the quality of blood analyses and helped to draft the manuscript. ES performed data analysis. CH analyzed 24,25(OH)₂D₃. MAG established the method and quality control of 24,25(OH)₂D₃. JD participated in the design of the study. GIS conceived the study, prepared the manuscript. All authors read and approved the final manuscript.

Funding sources

This work was funded by a grant from the German Federal Ministry of Education and Research, Grant No. 01EA1323A.

Conflict of interest

The authors declare that they have no conflict of interest.

Acknowledgments

We thank the staff of the Department of Transfusion Medicine (University Hospital Halle/Saale) for their assistance at recruitment and sampling.

Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.clnu.2015.04.021>.

References

- [1] Ross AC, Taylo CL, Yaktine AL, Valle HB. Dietary reference intakes for calcium and vitamin D: institute of medicine (US) committee to review dietary reference intakes for vitamin D and calcium. The National Academies Collection: Reports funded by National Institutes of Health. 2011.
- [2] Pludowski P, Holick MF, Pilz S, Wagner CL, Hollis BW, Grant WB, et al. Vitamin D effects on musculoskeletal health, immunity, autoimmunity, cardiovascular disease, cancer, fertility, pregnancy, dementia and mortality—a review of recent evidence. *Autoimmun Rev* 2013;12:976–89.
- [3] Spedding S, Vanlint S, Morris H, Scragg R. Does vitamin D sufficiency equate to a single serum 25-hydroxyvitamin D level or are different levels required for non-skeletal diseases? *Nutrients* 2013;5:5127–39.
- [4] Heaney RP. Toward a physiological referent for the vitamin D requirement. *J Endocrinol Investig* 2014;37:1127–30.
- [5] Kühn T, Kaaks R, Teucher B, Hirche B, Dierkes J, Weikert C, et al. Dietary, lifestyle, and genetic determinants of vitamin D status: a cross-sectional

- analysis from the European Prospective Investigation into Cancer and Nutrition (EPIC)-Germany study. *Eur J Nutr* 2014;53:731–41.
- [6] German Nutrition Society. New reference values for vitamin D. *Ann Nutr Metab* 2012;60:241–6.
- [7] Cashman KD, Hill TR, Lucey AJ, Taylor N, Seamans KM, Muldowney S, et al. Estimation of the dietary requirement for vitamin D in healthy adults. *Am J Clin Nutr* 2008;88:1535–42.
- [8] Blum M, Dallal GE, Dawson-Hughes B. Body size and serum 25 hydroxy vitamin D response to oral supplements in healthy older adults. *J Am Coll Nutr* 2008;27:274–9.
- [9] Didriksen A, Grimnes G, Hutchinson MS, Kjærgaard M, Svartberg J, Joakimsen RM, et al. The serum 25-hydroxyvitamin D response to vitamin D supplementation is related to genetic factors, BMI, and baseline levels. *Eur J Endocrinol* 2013;169:559–67.
- [10] Waterhouse M, Tran B, Armstrong BK, Baxter C, Ebeling PR, English DR, et al. Environmental, personal, and genetic determinants of response to vitamin D supplementation in older adults. *J Clin Endocrinol Metab* 2014;99:E1332–40.
- [11] Saliba W, Barnett-Griness O, Rennert G. The relationship between obesity and the increase in serum 25(OH)D levels in response to vitamin D supplementation. *Osteoporos Int* 2013;24:1447–54.
- [12] Nelson ML, Blum JM, Hollis BW, Rosen C, Sullivan SS. Supplements of 20 microg/d cholecalciferol optimized serum 25-hydroxyvitamin D concentrations in 80% of premenopausal women in winter. *J Nutr* 2009;139:540–6.
- [13] Faul F, Erdfelder E, Lang A-G, Buchner A. G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav Res Methods* 2007;39:175–91.
- [14] Schutkowski A, Krämer J, Kluge H, Hirche F, Krombholz A, Theumer T, et al. UVB exposure of farm animals: study on a food-based strategy to bridge the gap between current vitamin D intakes and dietary targets. *PLoS One* 2013;8:e69418.
- [15] Lehmann U, Hirche F, Stangl GI, Hinz K, Westphal S, Dierkes J. Bioavailability of vitamin D₂ and D₃ in healthy volunteers, a randomized placebo-controlled trial. *J Clin Endocrinol Metab* 2013;98:4339–45.
- [16] Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an endocrine society clinical practice guideline. *J Clin Endocrinol Metab* 2011;96:1911–30.
- [17] Heaney RP, Davies KM, Chen TC, Holick MF, Barger-Lux MJ. Human serum 25-hydroxycholecalciferol response to extended oral dosing with cholecalciferol. *Am J Clin Nutr* 2003;77:204–10.
- [18] Beckman MJ, Tadikonda P, Werner E, Pahl J, Yamada S, DeLuca HF. Human 25-hydroxyvitamin D₃-24-hydroxylase, a multicatalytic enzyme. *Biochemistry* 1996;35:8465–72.
- [19] Cashman KD, Hayes A, O'Donovan SM, Zhang JY, Kinsella M, Galvin K, et al. Dietary calcium does not interact with vitamin D₃ in terms of determining the response and catabolism of serum 25-hydroxyvitamin D during winter in older adults. *Am J Clin Nutr* 2014;99:1414–23.
- [20] Cipriani C, Romagnoli E, Pepe J, Russo S, Carlucci L, Piemonte S, et al. Long-term bioavailability after a single oral or intramuscular administration of 600,000 IU of ergocalciferol or cholecalciferol: implications for treatment and prophylaxis. *J Clin Endocrinol Metab* 2013;98:2709–15.
- [21] Wagner D, Hanwell HE, Schnabl K, Yazdanpanah M, Kimball S, Fu L, et al. The ratio of serum 24,25-dihydroxyvitamin D₃ to 25-hydroxyvitamin D₃ is predictive of 25-hydroxyvitamin D₃ response to vitamin D₃ supplementation. *J Steroid Biochem Mol Biol* 2011;126:72–7.
- [22] Bhattacharyya MH, DeLuca HF. The regulation of rat liver calciferol-25-hydroxylase. *J Biol Chem* 1973;248:2969–73.
- [23] Gallagher JC, Sai A, Templin T, Smith L. Dose response to vitamin D supplementation in postmenopausal women: a randomized trial. *Ann Intern Med* 2012;156:425–37.
- [24] Shab-Bidar S, Bours S, Geusens PP, Kessels AG, van den Bergh JP. Serum 25(OH)D response to vitamin D₃ supplementation: a meta-regression analysis. *Nutrition* 2014;30:975–85. <http://dx.doi.org/10.1016/j.nut.2013.12.020>.
- [25] Arunabh S, Pollack S, Yeh J, Aloia JF. Body fat content and 25-hydroxyvitamin D levels in healthy women. *J Clin Endocrinol Metab* 2003;88:157–61.
- [26] Vimalaswaran KS, Berry DJ, Lu C, Tikkanen E, Pilz S, Hiraki LT, et al. Causal relationship between obesity and vitamin D status: bi-directional Mendelian randomization analysis of multiple cohorts. *PLoS Med* 2013;10:e1001383.
- [27] Wortsman J, Matsuoka LY, Chen TC, Lu Z, Holick MF. Decreased bioavailability of vitamin D in obesity. *Am J Clin Nutr* 2000;72:690–3.
- [28] Valcour A, Blocki F, Hawkins DM, Rao SD. Effects of age and serum 25-OH-vitamin D on serum parathyroid hormone levels. *J Clin Endocrinol Metab* 2012;97:3989–95.
- [29] Jorde R, Sundsfjord J, Bønaa KH. Determinants of serum calcium in men and women. The Tromsø Study. *Eur J Epidemiol* 2001;17:1117–23.
- [30] Saito H, Maeda A, Ohtomo S, Hirata M, Kusano K, Kato S, et al. Circulating FGF-23 is regulated by 1alpha,25-dihydroxyvitamin D₃ and phosphorus in vivo. *J Biol Chem* 2005;280:2543–9.

3.3. Study 3

Efficacy of vitamin D-fortified rainbow trout on vitamin D status

3.3.1. Study Design

The study was conducted as a double blind, randomized, parallel study during November and December, when virtually no UVB irradiation is measurable in Halle and surroundings. Study visits were scheduled at baseline and after 4 weeks. Subjects were block randomized (stratified for body-mass index, sex and 25(OH)D₃-concentrations in serum as obtained during a screening visit 6-8 weeks prior to baseline) to receive either vitamin D₃-fortified or conventional fish. Fillets of trout's with skin were used for the study. The participants received their portion of rainbow trout (100g) six times per week at lunchtime to verify their compliance. The time of consumption was not specified. During each study visit, a venous blood sample was taken for analysis of 25(OH)D₃, PTH, serum calcium, total cholesterol, HDL and LDL cholesterol after a fasting period of at least 4 hours.

The study protocol was evaluated and approved by the ethic committee of the Medical Faculty at the Martin-Luther-University Halle-Wittenberg and each participant gave written, informed consent prior to the start of the study. The study was registered at clinicaltrials.gov, (NCT01696526).

3.3.2. Subjects

Participants were recruited among students and employees (only those who are unrelated to Nutritional science) on the campus through personal contacts and leaflets information in the library, cafeteria or hallways of the University. During a screening in autumn (about 1 months before the study start), the participants filled in a self-administered questionnaire on disease history, weight and height, lifestyle-behaviors (smoking, use of cosmetics containing sunblocker) and dietary habits concerning ingestion of food rich in vitamin D. Exclusion criteria were pregnancy and lactation, use of vitamin D supplements, vacation in regions with UVB light during an interval of 8 weeks before and during the study, subjects with known renal diseases or malignant diseases, participation in other clinical studies. Participants with elevated creatinine (in females ≥ 1.1 mg/dl, in males ≥ 1.3 mg/dl) were excluded. People who were already

well supplied with vitamin D (25-(OH)D₃-concentrations > 75 nmol/l) were excluded from study participation (n = 7). Five volunteers resigned from participation for personal reasons. In total, 56 subjects were recruited for the intervention study. Due to personal reasons, 3 participants dropped out during the study period. Finally, 53 subjects (age range 20 – 63 years) were included into the study. Characteristics of subjects are provided in Table 5.

3.3.3. Production of bio-fortified fish

The conventional and improved fish were provided and portioned by Forellenhof Thiessen (Coswig, Germany). The enrichment of the fillets with vitamin D₃ via UVB-radiation was held in Institute of Inland Fisheries (Potsdam-Sacrow, Germany). Fillets were irradiated with UVB light (G8 T5E UV-B, Sanyo Denki Germany GmbH, Eschborn, Germany) in vitro for 6 hours on ice. The irradiation intensity was measured in the medium at about 2500 mW/m² with distance of 27 cm from the light source. Bio-fortified fish and conventional fish were of identical appearance and indistinguishable from each other.

3.3.4. Methods

Blood samples for analysis were taken at the beginning and at the end of the study and collected in serum- and EDTA-tubes (Becton Dickinson, Heidelberg, Germany). Serum samples were centrifuged at 2000 g for 10 minutes at room temperature. EDTA samples were stored on ice till centrifugation at 2000 g for 10 minutes at 4° C. The samples were separated into aliquots and frozen at -80° C until the time of analysis. Serum concentrations of 25(OH)D₃ were determined by using a liquid chromatography tandem mass spectrometry (LC-MS/MS), MassChrom®25-OH Vitamin D₃ (Chromsystems GmbH, Munich, Germany), on a API 2000 (Applied Biosystems, Carlsbad, CA). The coefficient of variation for 25(OH)D₃ measurements was 6.5%, and the lower level of detection was 2.5 nmol/l. Serum creatinine was determined spectrophotometrically (DiaSys Diagnostic Systems GmbH, Holzheim, Germany). Total cholesterol, HDL cholesterol and LDL cholesterol were quantified spectrophotometrically (DiaSys Diagnostic Systems GmbH, Holzheim, Germany). Serum concentrations of parathyroid hormone (PTH) were measured using a

commercial ELISA Kit (IBL International GmbH, Hamburg, Germany). All measurements were made in duplicate.

3.3.5. Analysis of cholecalciferol concentration in fillet and skin of rainbow trout

Cholecalciferol in rainbow trout was determined by LC-MS/MS according to [Schutkowski et al., 2013] and [Higashi et al., 2008]. Samples were homogenized, mixed with deuterated internal standard (D_3 - d_3 , Sigma-Aldrich Chemie GmbH, Taufkirchen, Germany) and hydrolyzed under exclusion of oxygen. After extraction with n-hexane, hexane phase was washed with ultrapure water. Samples were fractionated by HPLC (Agilent 1100 HPLC, Agilent Technologies) and derivatized with 4-phenyl-1,2,4-triazolin-3,5-dione (solved in acetonitrile) according to [Mattila et al., 1995]. Ethanol and mobile phase were added to the dried residue and then analyzed by HPLC coupled to a MS system.

3.3.6. Statistical analysis

Statistical analysis was performed with SPSS, version 20.0 (Chicago, IL, USA). All data are expressed as mean \pm SD, and a p-level of <0.05 was regarded as significant. The primary outcome variables were $25(OH)D_3$ -concentrations which were compared between groups at baseline and after 4 weeks by Student's t-Test. Additionally, the change in these parameters within groups after 4 weeks to baseline were compared by paired t-test. The statistical power calculation revealed that 22 subjects per group would be required to show a difference of 15 nmol/l in the mean $25(OH)D_3$ -concentration after 4 weeks of fish consumption between the control and intervention group, (at an assumed standard variation of 15 nmol/l for each group, at a power of 80% and a significance level of 0.05). Only subjects who finished the study according to protocol were included into the analyses.

3.3.7. Results

The baseline characteristics were shown in Table 5. The study population consisted of 24 males and 29 females who were randomly assigned to either intervention ($n=26$) or control group ($n=27$). The study groups did not differ in body mass index (BMI) and age. On average, the participants were 29 ± 9.4 years and had a BMI of 23.2 ± 3.0

(kg/m²). Furthermore no significant differences between both groups were demonstrated in serum calcium, PTH, 25(OH)D₃, total cholesterol, LDL and HDL cholesterol.

Table 5: Baseline characteristics of the study population

	Intervention group	Control group	p-value
Sex (male/female)	13 / 13	11 / 16	0.508
Age [years]	30.3 ± 11.5	27.7 ± 6.8	0.325
BMI [kg/m ²]	23.1 ± 3.0	23.2 ± 3.0	0.912
25(OH)D₃ [nmol/l]	44.8 ± 14.0	43.0 ± 14.5	0.651
Calcium [nmol/l]	2.3 ± 0.1	2.3 ± 0.1	0.823
Total cholesterol [mmol/l]	4.5 ± 0.6	4.2 ± 0.6	0.090
LDL cholesterol [mmol/l]	2.6 ± 0.6	2.5 ± 0.6	0.477
HDL cholesterol [mmol/l]	1.5 ± 0.4	1.4 ± 0.4	0.394
PTH [pg/ml]	66.5 ± 27.2	66.1 ± 30.0	0.877

Student's t-test was used to compare intervention and control group at baseline

25-(OH)D₃-concentrations

At baseline, 25-(OH)D₃-concentrations did not differ between control and intervention group (44.8±14.0 vs. 43.0±14.5 nmol/l, $p=0.651$; respectively). In both groups, the 25(OH)D₃-concentrations decreased during the study. After four weeks 25(OH)D₃-concentrations were significantly higher in the intervention compared to control group (42.0±12.2 vs. 33.9±10.6 nmol/l, $p=0.013$; respectively). The 25(OH)D₃-concentrations after four weeks was significantly different from baseline in both groups (Figure 2). Decrease of 25(OH)D₃-concentration between baseline and 4 weeks showed a significant difference within the groups (control: -9.1±9.1; intervention: -2.7±6.9; $p=0.004$), with a smaller decline in the intervention group.

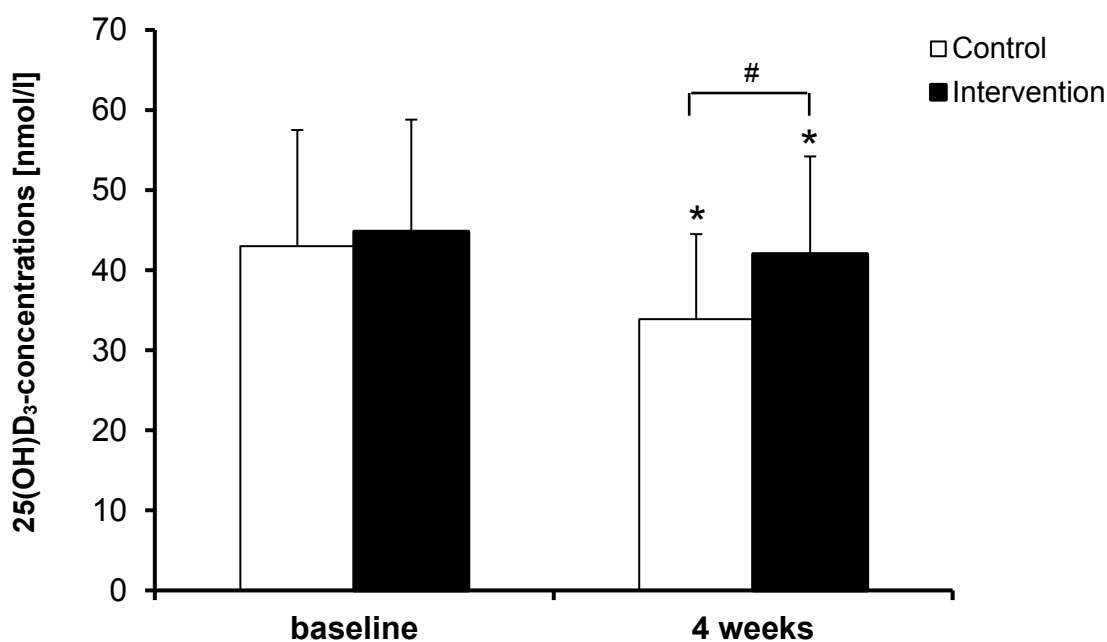


Figure 2: 25(OH)D₃ concentrations [nmol/l] in the control and intervention group at baseline and after four weeks.

A paired *t*-test was used to compare baseline and four weeks values.

Student's *t*-test was used to compare groups at baseline and four weeks

* significant different from baseline

significant different between groups

Concentrations of total cholesterol, HDL and LDL cholesterol

At baseline, neither total cholesterol nor HDL or LDL cholesterol was different between control and intervention group. After four weeks, total cholesterol was significantly increased in both groups ($p \leq 0.01$), but showed no differences between control and intervention group (4.65 ± 0.57 vs. 4.90 ± 0.82 mmol/l, $p=0.212$; respectively). Furthermore, HDL cholesterol did not differ between groups after 4 weeks (control: 1.71 ± 0.55 , intervention: 1.77 ± 0.54 mmol/l, $p=0.713$), but was increased significantly in both groups compared to baseline concentrations ($p \leq 0.01$). After 4 weeks LDL cholesterol did not differ between control and intervention group (2.56 ± 0.60 vs. 2.69 ± 0.76 mmol/l, $p=0.657$). In addition, LDL concentrations remained unchanged compared to baseline concentrations in both groups (control: $p=0.420$, intervention: $p=0.243$).

Concentrations of calcium and parathyroid hormone

Serum calcium concentrations did not show differences between control and intervention group after four weeks at second study visit (2.36 ± 0.12 , 2.32 ± 0.15 nmol/l, $p=0.823$; respectively) and remained unchanged compared to baseline concentrations

in both groups (control: $p=0.141$, intervention: $p=0.546$). Intact parathyroid hormone did not differ between groups after 4 weeks ($p=0.945$). Furthermore it did not change significantly within four weeks neither control nor intervention group ($p=0.780$, $p=0.793$; respectively).

3.4. Study 4

Lehmann, U., Gjessing, H.R., Hirche, F., Mueller-Belecke, A., Gudbrandsen, O.A., Ueland, P.M., Mellgran G., Laurizen L., Lindqvist H., Hansen A.L., Erkkilä, A.T., Pot G.K., Stangl G.I., & Dierkes J. (2015). Efficacy of fish intake on vitamin D status: a meta-analysis of randomized controlled trials. *The American journal of clinical nutrition*, 102(4), 837-847.

Efficacy of fish intake on vitamin D status: a meta-analysis of randomized controlled trials^{1,2}

Ulrike Lehmann,^{3,5} Hanne Rosendahl Gjessing,⁴ Frank Hirche,³ Andreas Mueller-Belecke,⁷ Oddrun Anita Gudbrandsen,⁵ Per Magne Ueland,⁸ Gunnar Mellgren,⁴ Lotte Lauritzen,⁹ Helen Lindqvist,¹⁰ Anita Lill Hansen,⁶ Arja T Erkkilä,¹¹ Gerda K Pot,¹² Gabriele I Stangl,³ and Jutta Dierkes^{5*}

³Institute of Agricultural and Nutritional Sciences, Martin Luther University of Halle-Wittenberg, Halle, Germany; Departments of ⁴Clinical Science, ⁵Clinical Medicine, and ⁶Psychosocial Science, University of Bergen, Bergen, Norway; ⁷Institute of Inland Fisheries, Potsdam-Sacrow, Germany; ⁸Bevital AS, Bergen, Norway; ⁹Department of Nutrition, Exercise, and Sports, Faculty of Sciences, University of Copenhagen, Copenhagen, Denmark; ¹⁰Department of Internal Medicine and Clinical Nutrition, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden; ¹¹Institute of Public Health and Clinical Nutrition, University of Eastern Finland, Kuopio, Finland; and ¹²Diabetes and Nutritional Sciences Division, King's College London, London, United Kingdom

ABSTRACT

Background: It is well known that fish is the major natural source of vitamin D in the diet; therefore, this meta-analysis investigated the influence of fish consumption in randomized controlled trials (RCTs) on serum 25-hydroxyvitamin D [25(OH)D] concentrations.

Objective: A literature search was carried out in Medline, Embase, Web of Science, and the Cochrane Library (up to February 2014) for RCTs that investigated the effect of fish consumption on 25(OH)D concentrations in comparison to other dietary interventions.

Results: Seven articles and 2 unpublished study data sets with 640 subjects and 14 study groups met the inclusion criteria and were included in this meta-analysis. Compared with controls, the consumption of fish increased 25(OH)D concentrations, on average, by 4.4 nmol/L (95% CI: 1.7, 7.1 nmol/L; $P < 0.0001$, $I^2 = 25\%$; 9 studies). The type of the fish also played a key role: the consumption of fatty fish resulted in a mean difference of 6.8 nmol/L (95% CI: 3.7, 9.9 nmol/L; $P < 0.0001$, $I^2 = 0\%$; 7 study groups), whereas for lean fish the mean difference was 1.9 nmol/L (95% CI: -2.3, 6.0 nmol/L; $P < 0.38$, $I^2 = 37\%$; 7 study groups). Short-term studies (4–8 wk) showed a mean difference of 3.8 nmol/L (95% CI: 0.6, 6.9 nmol/L; $P < 0.02$, $I^2 = 38\%$; 10 study groups), whereas in long-term studies (~6 mo) the mean difference was 8.3 nmol/L (95% CI: 2.1, 14.5 nmol/L; $P < 0.009$, $I^2 = 0\%$; 4 study groups).

Conclusion: As the major food source of vitamin D, fish consumption increases concentrations of 25(OH)D, although recommended fish intakes cannot optimize vitamin D status. *Am J Clin Nutr* 2015;102:837–47.

Keywords: fish intake, meta-analysis, randomized controlled trial, vitamin D, intervention studies, 25(OH)D, vitamin D status

INTRODUCTION

Vitamin D deficiency is a global problem and is associated with an increased risk of cardiovascular diseases (1–4), autoimmune diseases (5), type 1 diabetes (6, 7), osteoporosis (6), and probably various types of cancer (8–10). Although vitamin D is synthesized in the skin on exposure to UV-B radiation, it is not possible to maintain an adequate vitamin D status during winter

at high latitudes when UV-B radiation is absent (11). Fish, egg yolk, cheese, and mushrooms are the only dietary sources that contain natural vitamin D (12). Among these, fish has, in general, the highest content of vitamin D (12, 13) and is the major natural food source in many populations within (14–17) and outside of (18, 19) Europe. Other significant food sources are fortified items such as margarine, skimmed milk, and orange juice (20, 21). Although, in general, fish is a good source of vitamin D, there are considerable differences in vitamin D content between different fish species (13, 22). Other important factors are environmental conditions, such as season, and the fat content of the fish (13), but more research is needed in this area.

In observational studies (23, 24) fish consumption was shown to have a beneficial effect on cardiovascular morbidity and mortality, although it must be considered that these health effects could also be due to other constituents present in fish, such as long-chain n-3 PUFAs, amino acids, iodine, or selenium, in addition to vitamin D. The effects of short- to medium-term fish interventions on PUFAs (25–28, 30), blood lipids (25–27, 29–34), vitamin B-12 and selenium status (28, 35), insulin and leptin concentrations (29), eicosanoids and adhesion molecule concentrations (36), heart rate variability (25, 34), and vitamin D status have been investigated in several randomized controlled trials (RCTs)¹³ (25–28, 34, 35, 37), but systematic studies of the

¹ Supported by grant 01EA1323A from the German Federal Ministry of Education and Research and by personal grants from The Research Council of Norway (to UL; grant 227506/F11) and from Bergen Medical Research Foundation (to OAG).

² Supplemental Text and Supplemental Tables 1–3 are available from the “Supplemental data” link in the online posting of the article and from the same link in the online table of contents at <http://ajcn.nutrition.org>.

*To whom correspondence should be addressed. E-mail: jutta.dierkes@med.uib.no.

¹³ Abbreviations used: LC-MS/MS, liquid chromatography–tandem mass spectrometry; RCT, randomized controlled trial; 25(OH)D, 25-hydroxyvitamin D; 25(OH)D₃, 25-hydroxycholecalciferol.

Received December 12, 2014. Accepted for publication July 8, 2015.

First published online September 9, 2015; doi: 10.3945/ajcn.114.105395.

extent to which fish consumption may contribute to dietary status or to biomarkers for nutrient uptake are lacking. To the best of our knowledge, the effect of fish consumption on vitamin D status has not been investigated systematically. Because increased vitamin D intake due to regular fish consumption may be one explanation for the beneficial health effects of fish, the aim of this study was to conduct a meta-analysis of RCTs on the effect of fish consumption on serum 25-hydroxyvitamin D [25(OH)D] concentrations as the outcome.

METHODS

Search strategies and data collection

To identify relevant studies, Medline, Embase, Web of Science, and the Cochrane Library databases were searched between January 1950 and 12 February 2014. The following search terms were used: vitamin D, cholecalciferol, ergocalciferol, hydroxycholecalciferols, dihydroxycholecalciferol, calcitriol, 24,25-dihydroxyvitamin D, 25-hydroxyvitamin D, 1,25-dihydroxyvitamin D, calcidiol or calciol, fishes, seafood, shellfish, clinical trial, and random trial or parallel trial (as shown in **Supplemental Table 1**). Additional studies were identified by manual searches through references or the clinicaltrials.gov database. The search was restricted to studies published in English.

The studies were assessed by 2 independent investigators (UL and JD), taking the inclusion criteria into account. Data on the primary patients were collected by personal communication with the relevant investigators by e-mail. Standard data files were provided for this purpose. Investigators who agreed to collaborate were asked to provide data for each participant, including the measured serum 25(OH)D concentration, the definition of the fish consumption group, age and sex, BMI, and the season in which blood samples were taken.

Study eligibility criteria

Any randomized intervention trial that involved human adults and investigated the effects of fish meals on serum 25(OH)D concentrations was included in the analysis. We excluded studies that used only a food-frequency questionnaire to calculate fish intake and studies with only 1 fish meal/wk as an intervention (38). In addition to studies in healthy participants, those that included patients who had survived a myocardial infarction or overweight subjects were also included in the meta-analysis. Studies that involved children, adolescents, or pregnant or breastfeeding women were excluded. Differences between the extracted studies in daily fish intake, the frequency of meals containing fish, or study duration were not a cause for exclusion.

Data collection

The quality of the included studies was checked manually by careful examination of the original publications. Several studies did not originally intend to evaluate the effect of fish consumption on vitamin D status, and therefore most studies did not adequately report the methods of 25(OH)D measurement or the season of blood collection. Because this meta-analysis was concerned with the effect of real food, the issue of blinding was not applicable to the participants. Indeed, only 1 study (U Lehmann, unpublished data, 2012) was sufficiently blinded to participants, as expected in

studies investigating natural food. In most studies, meat was used as the comparator or no food was provided to the participants in the control group. In 2 studies, fish with a low vitamin D content was used as the control intervention (27; U Lehmann, unpublished data, 2012). The accepted quality-control measures, such as the Jadad scale (43) or the CONSORT(Consolidated Standards of Reporting Trials) statement (44), were therefore not appropriate for estimating study quality. The quality of the studies was instead assessed on the basis of compliance, number of dropouts, measurements of the vitamin D content in the fish, season of the intervention, the type of vitamin D analysis, and the type of randomization. One score point was given for each item of information included. Scores of 5–6 denote good quality, 3–4 indicate moderate quality, and 0–2 points denote low quality.

Analysis of the data

Studies were analyzed by using RevMan 5.2, which was provided by the Cochrane Collaboration. After consultation with the relevant authors, we received individual patient data from 6 trials (26–28, 35; U Lehmann, unpublished data, 2012; OA Gudbrandsen, unpublished data, 2014). For each study we recorded the number of subjects and mean (SD) baseline and postintervention 25(OH)D concentrations separately for controls and for the intervention group. The mean change in 25(OH)D was calculated by subtracting the mean baseline 25(OH)D concentration from the mean postintervention 25(OH)D concentration. For calculation of the SD of the change in 25(OH)D we applied a correlation coefficient of 0.82 in the control group and a correlation coefficient of 0.77 in the intervention group. These correlation coefficients were calculated from studies with access to individual data ($n = 6$), according to the Cochrane Handbook (39).

Studies that included >1 intervention group (26–28, 37; OA Gudbrandsen, unpublished data, 2014) were treated by dividing the number of subjects in the control group by the number of comparisons while retaining the mean and SD of the change according to the Cochrane Handbook (40).

The changes in 25(OH)D concentrations were calculated as weighted mean differences with 95% CIs. Statistical heterogeneity between the studies was tested by using the Cochrane Q-test (41). A random-effects model was applied. Publication bias was assessed by a funnel plot (**Figure 1**) (42). In addition to the main analysis, we conducted several sensitivity analyses taking into account study duration, type of fish, mean baseline 25(OH)D concentrations, season of blood collection, access to individual data or calculated data, participants' health status, the amount of fish consumed during the trial, measurements of total 25(OH)D or 25-hydroxycholecalciferol [25(OH)D₃], the determination of the vitamin D content in the fish, and the method of determination of 25(OH)D concentrations [ELISA/radioimmunoassay or liquid chromatography–tandem mass spectrometry (LC-MS/MS)]. Two studies (27; U Lehmann, unpublished data, 2012) compared fish with different vitamin D contents. These 2 fish interventions were compared in an additional separate analysis.

Included studies

In addition to published studies, we included 2 unpublished RCTs involving fish consumption in healthy adults. One of these was conducted at the University of Bergen in Norway and the

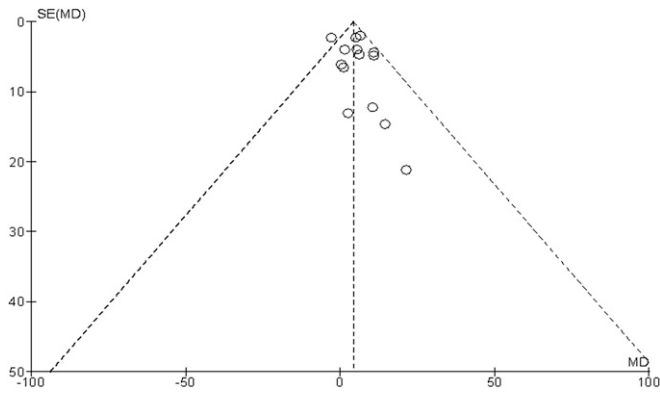


FIGURE 1 Funnel plot with pseudo 95% CIs for the effect of fish intake on serum 25-hydroxyvitamin D concentrations. MD, mean difference.

other at the Martin Luther University of Halle-Wittenberg in Germany. These studies are described briefly below and in greater detail in the **Supplemental Text**.

Lehmann study

The study in Halle (Saale) at the Martin Luther University of Halle-Wittenberg (latitude 51° north) was conducted during the late autumn of 2012. The major aim was to compare the effect of vitamin D-enriched rainbow trout on 25(OH)D₃ concentrations in comparison with conventional rainbow trout in healthy volunteers (*n* = 53) over a 4-wk period. The participants consumed 6 times/wk 100-g portions of rainbow trout enriched with vitamin D by postmortem irradiation with UV-B or 100-g portions of conventional, untreated rainbow trout fillets. Consumption was usually at lunchtime and was supervised on weekdays. Both participants and investigators were blinded to the type of trout. Blood samples were collected at baseline and after 4 wk for determination of 25(OH)D₃

concentrations by LC-MS/MS (MassChrom 25-OH Vitamin D₃ reagent kit for LC-MS/MS analysis; Chromsystems GmbH) on an API2000 LC-MS/MS system (Applied Biosystems), as described elsewhere (51). Characteristics of participants are provided in **Supplemental Table 2**.

Gudbrandsen study

This was a randomized controlled intervention study with a parallel design and 3 intervention arms: cod, salmon, or chicken in weekly doses of 750 g (5 meals of 150 g)/wk for 4 wk, with study visits at baseline and after 4 wk. The study included 57 participants recruited in Bergen, Norway, and randomly assigned to the intervention groups. Because of the reduced number of blood samples (*n* = 5 with missing data) and dropouts (*n* = 3), samples for the 25(OH)D analyses were only available for 19, 18, and 12 participants, respectively. Fasting blood samples were collected at baseline and after 4 wk, and 25(OH)D was determined in serum by LC-MS/MS according to methods of Middtun and Ueland (45). Characteristics of participants are provided in **Supplemental Table 3**.

RESULTS

A systematic search of the literature led to the identification of 3277 possibly relevant articles (**Figure 2**). A first examination identified 61 studies as appropriate to be included in the analysis by reviewing titles and abstracts. After detailed consideration, 54 studies were rejected from the analysis, because they did not measure 25(OH)D as an outcome, were not RCTs, gave no detailed information on amount of fish, or were duplicates of included studies. In total, 7 published and 2 unpublished studies that fulfilled the inclusion criteria were included in the present meta-analysis investigating the effect of fish intake on serum 25(OH)D concentrations.

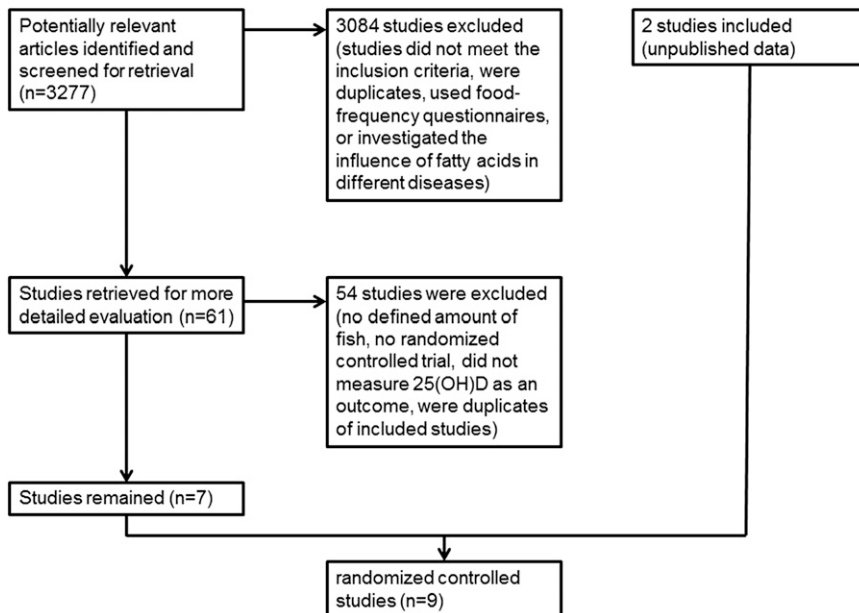


FIGURE 2 Flow diagram for the selection of studies of the effects of fish intake on serum 25(OH)D concentrations in the present meta-analysis, indicating numbers of articles reviewed and later excluded or included for the meta-analysis. 25(OH)D, 25-hydroxyvitamin D.

TABLE 1
 Characteristics of the randomized controlled trials included in the present meta-analysis¹

Authors (ref)	Year	Country	Intervention, dose, and frequency ²	Included in analysis, <i>n</i>	Sex and age	Follow-up	Baseline 25(OH)D (fish group), nmol/L
Erkkilä et al. (26) ³	2008	Finland	1) Control (lean meat or chicken) [10] 2) Lean fish (400–600 g/wk) [11] 3) Fatty fish (400–600 g/wk) [12]	33	27 men, 6 women; 61.0 ± 5.8 y	8 wk	— 96 ± 30 ⁴ 124 ± 43
Lucey et al. (37) ³	2008	Iceland, Ireland, Spain	1) Control (sunflower oil, 3 g/d) [66] 2) Cod (450 g/wk) [70] 3) Salmon (450 g/wk) [74]	210	92 men, 118 women; 20–40 y	8 wk	— 59.0 ± 22.1 61.9 ± 33.8
Pot et al. (28) ³	2009	United Kingdom, Netherlands	1) Control (dietary advice) [23] 2) Cod (300 g/wk) [22] 3) Salmon (300 g/wk) [29]	74	37 men, 37 women; 18–79 y	6 mo	— 71.6 ± 30 71.7 ± 26.5
Hallund et al. (27) ³	2010	Denmark	1) Control (chicken, 1050 g/wk) [22] 2) Trout raised on marine feed (1050 g/wk) [23] 3) Trout raised on vegetable-based feed (1050 g/wk) [23]	68	All men; 40–70 y	8 wk	— 45.9 ± 20.9 48.1 ± 22.1
Hansen et al. (34)	2010	Norway	1) Control (alternative dinner) [14] 2) Seafood, mainly salmon (600 g/wk) [15] 3) Chicken (750 g/wk) [12] 4) Cod (750 g/wk) [18] 5) Salmon (750 g/wk) [19]	29	All men; 20–60 y	23 wk	— 48 ± 15 66.5 ± 17.8 77.0 ± 23.1
OA Gudbrandsen (unpublished data, 2014) ³ U Lehmann (unpublished data, 2012)	2013	Germany	1) Control (common rainbow trout) (600 g/wk) [27] 2) Vitamin D-enriched rainbow trout (600 g/wk) [26]	53	24 men, 29 women; 20–63 y	4 wk	— 44.8 ± 14
Scheers et al. (35)	2013	Sweden	1) Control (650 g pork or 750 g chicken/wk) [21] 2) Herring (750 g/wk) [21] 3) Control (3 times meat/wk) [42] 4) Salmon (900 g/wk over 5 mo, 450 g/wk during the past 4 wk) [40]	21	All men; 35–60 y	2 × 6 wk (crossover)	— 66.9 ± 22.1 85 ± 36
Hansen et al. (25)	2014	Norway	1) Control (3 times meat/wk) [42] 2) Salmon (900 g/wk over 5 mo, 450 g/wk during the past 4 wk) [40]	82	All men; 18–61 y	6 mo	— 66.9 ± 22.1 85 ± 36

¹ref, reference; 25(OH)D, 25-hydroxyvitamin D.

²*n* in brackets.

³These studies included 2 intervention groups (fatty compared with lean fish).

⁴Mean ± SD (all such values).

TABLE 2
Quality evaluation of the 9 randomized controlled studies included in the meta-analysis¹

Authors, year (ref)	Compliance	Dropouts, <i>n</i> (%)	Measured vitamin D content in fish	Intervention period	Analytic method for 25(OH)D	Randomization	Participants	Quality score ²
Erkkilä et al., 2008 (26)	Checked by self-report and serum fatty acid composition	2 (5.7)	No data	Spring, August–September, October	ELISA	Stratified by sex	Survivors of myocardial infarction	4
Lucey et al., 2008 (37)	86%	34 (13.9)	Data cited but not measured (9.6 µg/100 g)	October–May	ELISA	No information	Overweight; consuming a low-calorie diet	4
Pot et al., 2009 (28)	Serum n-3 very-long-chain PUFAs	22 missing blood samples	No information	November 2004–July 2007	ELISA	Randomization in blocks (<i>n</i> = 6)	Only participants with healthy colon included	5
Hallund et al., 2010 (27)	99% evaluated in study diaries	7 (9.3)	Measured trout raised on marine feed: 0.62 µg/100 g; trout raised on vegetarian feed: <0.1 µg/100 g	Spring or autumn	Chemiluminescence immunoassay	Randomization without taking baseline characteristics into account	Healthy volunteers	6
Hansen et al., 2010 (34)	No information	24 (45.3)	No data	April–November	Radioimmunoassay	No information	Prisoners	3
OA Gudbrandsen (unpublished data, 2014)	Checked by self-report	8 (14)	No data	Winter	LC-MS/MS	Consideration of sex, BMI, and age	Healthy volunteers	4
U Lehmann (unpublished data, 2012)	93%	4 (7)	2.8 µg/100 g in intervention trout; 0.6 µg/100 g in control trout	November–December	LC-MS/MS	Randomization in blocks (<i>n</i> = 2); consideration of sex, BMI, and 25(OH)D ₃ concentrations at screening	Healthy volunteers	6
Scheers et al., 2013 (35)	Checked by self-report (24-h recall) and fatty acid concentration in blood	<i>n</i> = 5 dropouts; <i>n</i> = 19 for this analysis who were missing blood samples for 25(OH)D (19.2)	8.5 µg/100 g vitamin D in herring	April–June, September–November	HPLC	No randomization because crossover study	Healthy, overweight	6
Hansen et al., 2014 (25)	No information	11 (11.8)	15 µg/300 g in salmon but no source added	September – February	Chemiluminescence immunoassay	No information	Sex offenders	3

¹The quality score was based on assessment of compliance, number of dropouts, measurements of vitamin D content in fish, season of intervention, type of intervention, type of vitamin D analysis, and type of randomization. One score point was given for each item of information included. LC-MS/MS, liquid chromatography–tandem mass spectrometry; ref, reference; 25(OH)D, 25-hydroxyvitamin D; 25(OH)D₃, 25-hydroxycholecalciferol.

²Scores of study quality: 5–6 denotes good quality, 3–4 indicates moderate quality, and 0–2 points denotes low quality.

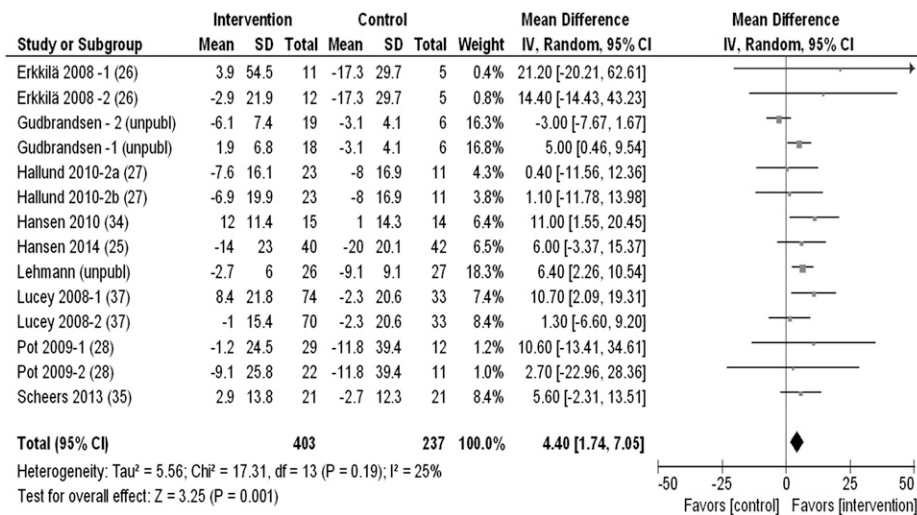


FIGURE 3 Random-effects meta-analysis comparing the effects of fish intervention with the control food on the 25(OH)D concentrations (in nmol/L). The forest plot indicates a significant effect of the absolute change in 25(OH)D concentrations compared with the control food by fish intervention. “Total” denotes the cumulative n from all of the included studies. IV, inverse variance; unpubl, unpublished; 25(OH)D, 25-hydroxyvitamin D; -1, study groups who received fatty fish; -2, study groups who received lean fish.

Study characteristics

The 9 studies considered comprised 619 participants [640 participants were included in the meta-analysis on account of 1 crossover study (34)] aged between 18 and 79 y. Descriptive study information is shown in **Table 1**. Sixty-four percent of the study population were men ($n = 396$) and 36% were women ($n = 223$). The studies were conducted in Finland, Iceland, Ireland, Spain, the United Kingdom, Netherlands, Denmark, Norway, the United States, Germany, and Sweden. Two studies (28, 37) were multicenter studies. Most of the participants were white, although a number of studies did not specify this explicitly. The change in 25(OH)D concentration served as the primary outcome in only 1 case (U Lehmann, unpublished data, 2012), whereas in the other studies the 25(OH)D concentration was measured as a secondary outcome (25–28, 34, 35) or was measured post hoc (OA Gudbrandsen, unpublished data, 2014).

The interventions differed between the studies in dosage, time, and fish species. We included 3 long-term studies with an intervention period of 6 mo or 23 wk (25, 28, 34). Six studies (26, 27, 35, 37; OA Gudbrandsen, unpublished data, 2014; U Lehmann, unpublished data, 2012) investigated the influence of short-term fish intake (4–8 wk) on 25(OH)D concentrations. The planned amount of fish to be consumed varied from 300 to 1050 g/wk. In 6 studies the weekly fish intake was planned to be between 300 and 600 g (25, 26, 28, 34, 37; U Lehmann, unpublished data, 2012), whereas in 3 studies (27, 35; OA Gudbrandsen, unpublished data, 2014) the intake was between 750 and 1050 g. The selected fish species differed between studies. The intake of fatty fish (salmon, herring) was investigated in 3 studies (25, 34, 35), whereas 4 studies compared both fatty and lean fish (cod, rainbow trout; 26, 28, 37; OA Gudbrandsen, unpublished data, 2014). One study included rainbow trout in the fatty fish group (26), and 2 studies investigated rainbow trout that differed in either the feeding regimen (27) or in postmortem treatment (U Lehmann, unpublished data, 2012). Total serum 25(OH)D concentrations measured by ELISA/immunoassay

were reported in 6 studies (25–28, 34, 37), whereas in 3 studies serum 25(OH)D₃ concentrations were measured by chromatographic methods (35; U Lehmann, unpublished data, 2012; OA Gudbrandsen, unpublished data, 2014).

All of the studies were designed as RCTs and included a control group. Because of the visible differences between the meals, 8 studies were not blinded. Only 1 study (U Lehmann, unpublished data, 2012) was double-blinded. Details of the randomization scheme and criteria were reported in 4 cases (27, 28; OA Gudbrandsen, unpublished data, 2014; U Lehmann, unpublished data, 2012), and 1 study had a crossover design (35). All of the studies provided general information on season of the intervention period, but an exact timing (month or season) of the blood collection procedures was usually not possible. Exact compliance rates were reported in only 3 studies (27, 37; U Lehmann, unpublished data, 2012), but the drop-out rates were given in all studies (25–28, 34, 35, 37; OA Gudbrandsen, unpublished data, 2014; U Lehmann, unpublished data, 2012).

Individual data were available for 6 of the 9 studies [298 participants, although 319 individual data are included because of 1 crossover study (35)] to calculate the change in 25(OH)D concentrations (26–28, 35; OA Gudbrandsen, unpublished data, 2014; U Lehmann, unpublished data, 2012), whereas the mean (SD) 25(OH)D concentrations at baseline and during follow-up in the study groups were available in 3 studies (25, 34, 37). The vitamin D₃ concentration in the fish was reported in 3 studies (27, 35; U Lehmann, unpublished results) but was explicitly measured only by Hallund et al. (27) and Lehmann (unpublished data, 2012). Average fish vitamin D₃ concentrations were cited by 2 studies (25, 37).

Results of the study quality assessment are presented in **Table 2**. The quality score was high in 5 studies (27, 28, 35; OA Gudbrandsen, unpublished data, 2014; U Lehmann, unpublished data, 2012) and moderate in 4 studies (25, 26, 34, 37). None of the studies had a low score.

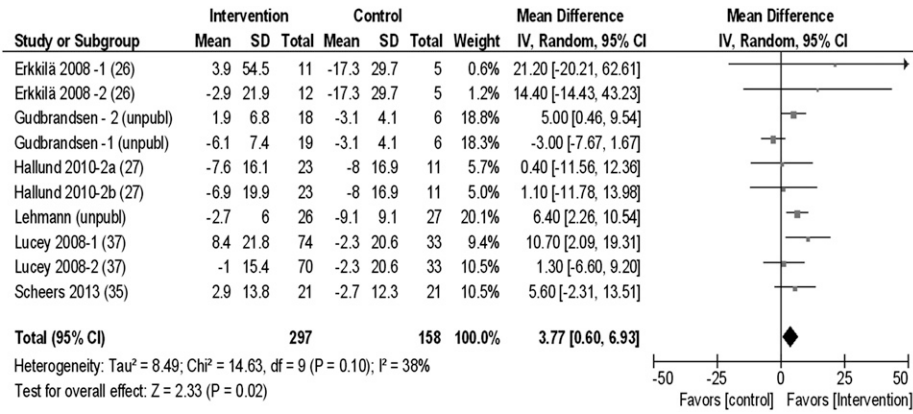


FIGURE 4 Random-effects meta-analysis comparing the effects of short-term (4–8 wk; 10 study groups) fish intervention with the control food on 25(OH)D concentrations (in nmol/L). The forest plot indicates a significant effect of the absolute change in 25(OH)D concentrations compared with the control food by fish intervention. “Total” denotes the cumulative *n* from all of the included studies. IV, inverse variance; unpubl, unpublished; 25(OH)D, 25-hydroxyvitamin D; -1, study groups who received fatty fish; -2, study groups who received lean fish.

Meta-analysis

Comparison of the changes in 25(OH)D concentrations between the fish intervention and the control groups including all studies (*n* = 9; 14 study groups) resulted in a weighted mean difference of 4.4 nmol/L (95% CI: 1.7, 7.1 nmol/L; *P* < 0.001, *I*² = 25%) (Figure 3).

The effect of the fish intervention varied depending on the study duration. Short-term studies (4–8 wk) showed a mean difference of 3.8 nmol/L (95% CI: 0.6, 6.9 nmol/L; *P* < 0.02, *I*² = 38%), whereas long-term studies (~6 mo) showed a mean difference of 8.3 nmol/L (95% CI: 2.1, 14.5 nmol/L; *P* < 0.009, *I*² = 0%) (Figures 4 and 5).

The type of fish also had an effect. Fatty fish (salmon, herring; *n* = 7 study groups) resulted in a mean difference of 6.8 nmol/L (95% CI: 3.7, 9.9 nmol/L; *P* < 0.0001, *I*² = 0%), whereas studies that used lean fish (trout, cod; *n* = 6 study groups) showed a mean difference of -1.1 nmol/L (95% CI: -4.7, 2.5 nmol/L; *P* < 0.55, *I*² = 0%). When the unpublished study by Lehmann was included (*n* = 7 study groups), which used lean fish that were biofortified with vitamin D, the weighted mean difference changed to 1.9 nmol/L (95% CI: -2.3, 6.0 nmol/L; *P* < 0.38, *I*² = 37%) (Figures 6 and 7).

In 2 studies, different types of rainbow trout were compared: the intervention group received trout that had been biofortified either by the feeding regimen (26) or by postmortem irradiation (U Lehmann, unpublished data, 2012). A separate meta-analysis

of these studies showed a weighted mean difference of 5.4 nmol/L (95% CI: 1.6, 9.1 nmol/L; *P* < 0.005, *I*² = 0%) between the intervention groups and controls.

An additional sensitivity analysis was carried out to investigate the influence of the mean baseline 25(OH)D concentration. In 3 studies that included 4 study groups, mean baseline 25(OH)D in the intervention groups was <50 nmol/L (27, 34; U Lehmann, unpublished data, 2012). The weighted mean difference was 6.1 nmol/L (95% CI: 2.7, 9.6 nmol/L; *P* < 0.0006, *I*² = 0%), compared with 3.9 nmol/L (95% CI: 0.4, 7.3 nmol/L; *P* < 0.03, *I*² = 30%) in 6 studies with 10 study groups in which mean baseline 25(OH)D concentrations were >50 nmol/L. A meta-analysis that used individual patients’ data that were available from 6 trials (26–28, 35; OA Gudbrandsen, unpublished data, 2014; U Lehmann, unpublished data, 2012) did not show results that were different from those in the analysis of aggregated data (data not shown).

DISCUSSION

In this meta-analysis we investigated whether fish intake increases serum 25(OH)D concentrations in healthy adults under controlled conditions and included 9 RCTs with good or moderate quality. The main result was that the consumption of at least 2 fish meals, corresponding to ~300 g fish/wk over a period of at least 4 wk, was associated with a significant increase in 25(OH)D

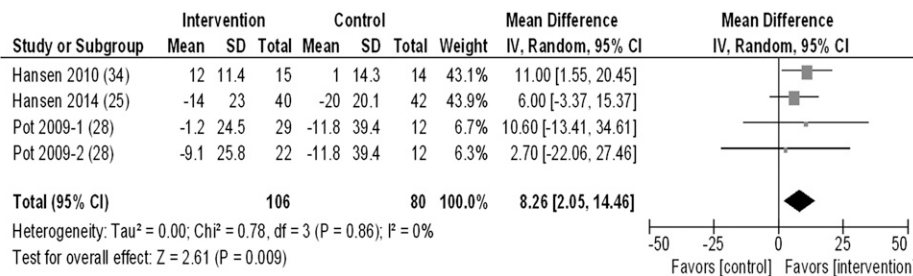


FIGURE 5 Random-effects meta-analysis comparing the effects of long-term (6 mo; 4 study groups) fish intervention with the control food on 25(OH)D concentrations (in nmol/L). The forest plot indicates a significant effect of the absolute change in 25(OH)D concentrations compared with the control food by fish intervention. “Total” denotes the cumulative *n* from all of the included studies. IV, inverse variance; 25(OH)D, 25-hydroxyvitamin D; -1, study groups who received fatty fish; -2, study groups who received lean fish.

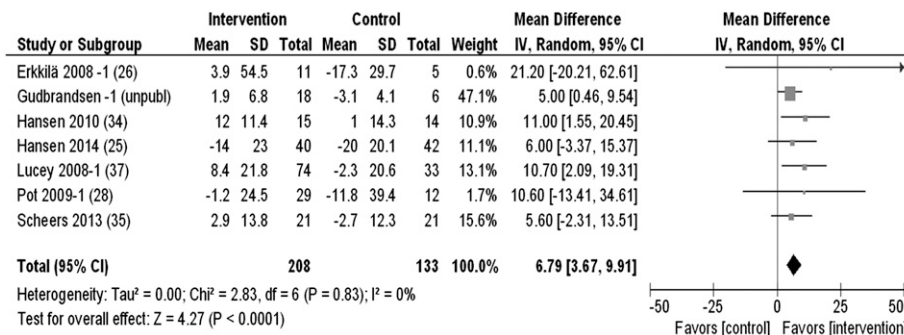


FIGURE 6 Random-effects meta-analysis comparing the effects of fatty fish intervention (7 study groups) with the control food on 25(OH)D concentrations (in nmol/L). The forest plot indicates a significant effect of the absolute change in 25(OH)D concentrations compared with the control food by fish intervention. "Total" denotes the cumulative *n* from all of the included studies. IV, inverse variance; unpubl, unpublished; 25(OH)D, 25-hydroxyvitamin D; -1, study groups who received fatty fish.

concentrations. In addition, fatty fish, longer study durations, and lower baseline 25(OH)D concentrations were associated with larger increases in 25(OH)D concentrations.

Although it has long been known that the consumption of fish is healthy, and this finding is included in most dietary recommendations, so far there have been few quantitative analyses supporting this effect on intermediate endpoints. The present analysis adds to our knowledge of the health-related effects of fish consumption and allows a quantitative estimate of what may be expected from increased fish consumption. This type of analysis is not very common in nutrition research and has not so far been included in recommendations and guidelines.

The present meta-analysis showed that the type of the fish is an important factor. Lean fish, mostly cod, did not increase vitamin D status to a significant extent, although it should be noted that differentiation between lean and fatty may be arbitrary in some species that could also be classified as medium-fatty fish. We observed a significant increase in 25(OH)D concentrations only when biofortified rainbow trout was included in the lean fish group. Thus, the consumption of fatty or biofortified fish should be recommended from the standpoint of improving vitamin D status.

In 2 studies, different types of rainbow trout were investigated: fish that were biofortified with vitamin D either by feeding or by postmortem exposure to UV-B radiation. These studies showed that there is a huge potential for improving the vitamin D content,

which is more pronounced by using postmortem irradiation than by feeding. However, both technologies should be developed further, because the absolute amounts of vitamin D in the treated fish were still relatively low. In this respect, it is interesting to note that preliminary data on freshwater fish species also indicate an effect of different living conditions on vitamin D content (46).

One side effect of high fish consumption may be an increased exposure to environmental toxins that accumulate in fish and in seafood. Health authorities such as the Norwegian Scientific Committee for Food Safety therefore recommended in 2007 an upper intake limit of 400 g fatty fish/wk (47). It has been shown that the accumulation of toxins was high in fatty fish species such as herring, salmon, and sprat (48). Within the same fish species contamination may vary depending on age, fat content, and geographic region (49). In our meta-analysis none of the studies investigated the association between fish intake and toxins, but this clearly should be taken into account when recommending high fatty fish intakes to improve vitamin D status and should be explored in future studies.

Our knowledge of the variation in vitamin D content in fish is limited. For example, the vitamin D content of the fish used throughout the study was only measured in 2 studies (27; U Lehmann, unpublished data, 2012). It may be assumed that, even within a given fish species, there is a wide variation in vitamin D content depending on growth, feed, and other factors such as season (13). For example, it has been hypothesized that farmed

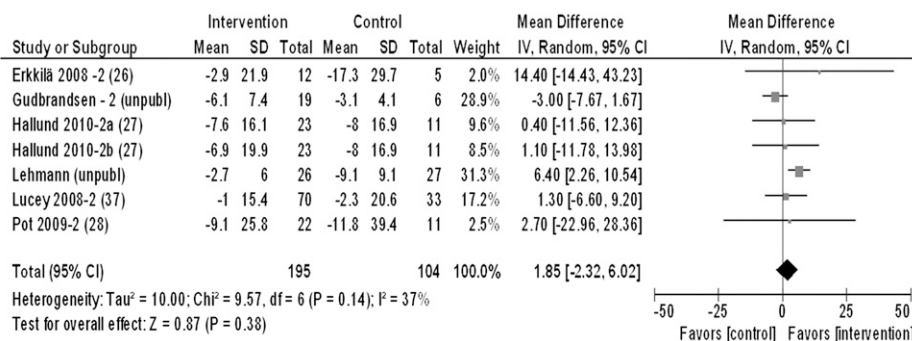


FIGURE 7 Random-effects meta-analysis comparing the effects of lean fish intervention (7 groups) with the control food on 25(OH)D concentrations (in nmol/L). The forest plot indicates a nonsignificant effect of the absolute change in 25(OH)D concentrations compared with the control food by fish intervention. "Total" denotes the cumulative *n* from all of the included studies. IV, inverse variance; unpubl, unpublished; 25(OH)D, 25-hydroxyvitamin D; -2, study groups who received lean fish.

salmon contains less vitamin D than does wild salmon (13, 50). Thus, there is a need for more detailed and accurate data on the determinants of vitamin D content in wild and farmed fish.

Although fish is one of the few foods that contain vitamin D (12), there is still an ongoing discussion whether fish intake contributes to a sufficient supply of vitamin D. Several observational studies (51–53) investigated the relation of fish intake and vitamin D status with the aid of food-frequency questionnaires. With the use of data from EPIC (European Prospective Investigation into Cancer and Nutrition)—Germany, Kühn et al. (51) reported a low, but positive, significant association between fish intake and 25(OH)D concentrations. In the United Kingdom, people who consumed meat and fish had higher 25(OH)D concentrations than did vegetarians and vegans (52). In Swedish women, fatty fish was one important predictor of serum 25(OH)D concentrations (53).

On the basis of all of the available data on fish intake, we observed a mean increase of 4.4 nmol 25(OH)D/L and an increase of 6.8 nmol/L when only fatty fish was considered. The application of the proposed increase of 25(OH)D of 1.97 nmol/L per additional microgram of vitamin D intake (54) suggests an intake of 2.2 μg vitamin D/d for all types of fish and 3.5 μg vitamin D/d for fatty fish such as salmon. According to the available data from food-composition tables, 300 g raw salmon/wk (corresponding to 2–3 portions) would provide 4.3, 6.9, or 2.5 μg vitamin D/d when using Norwegian, German, or UK databases, respectively (55–57). Whether these differences reflect natural variation or differences in analytic methods is unclear at present. Efforts to harmonize food-composition databases have been undertaken, e.g., by European Food Safety Authority or the EuroFIR project (www.eurofir.org).

The above calculations also show clearly that this fish intake is insufficient and does not fulfill the revised recommendations for a daily dietary amount of vitamin D that would improve vitamin D status (58–61). Indeed, it may be misleading to recommend fish consumption alone to improve vitamin D status. A daily intake of 2.2 or 3.5 μg vitamin D—which is achieved by consuming ~300–600 g fish/wk—will not increase serum 25(OH)D concentrations to an optimal level (>50 nmol/L) in most people and will only result in increases of 4.4 or 6.8 nmol/L, respectively. Our results are in line with dose-response studies conducted in older adults (54), which showed that subjects supplemented with 5 μg vitamin D/d were able to maintain 25(OH)D concentrations during wintertime, whereas supplementation with 10 $\mu\text{g}/\text{d}$ increased 25(OH)D concentrations by ~12 nmol/L, on average. In healthy postmenopausal women, a daily supplement of 800 IU vitamin D (corresponding to 20 μg) was needed to increase 25(OH)D concentrations to >50 nmol/L in almost all of the women (62).

Strengths and limitations of the study

A major strength of the study is the inclusion of only RCTs and the collection of individual patients' data for at least some of the studies. All of the studies reached either a high- or moderate-quality score, although the use of established quality scores was prevented by the use of real food and therefore lack of participant blinding in 8 of the 9 studies. Limitations included that, due to the low number of studies, no further sensitivity analyses with respect to the amount or type of fish, length of intervention, or

analytic method for determination of 25(OH)D were possible. In particular, the different analytic methods used for 25(OH)D measurements may have affected the results, because only 3 studies used chromatographic (LC-MS/MS or HPLC) methods.

Conclusions

We conclude that fish, as an important food source of vitamin D, increases 25(OH)D concentrations but cannot optimize vitamin D status. The side effects of the accumulation of environmental pollutants must be taken into account and investigated further. It should be clarified which foods are effective in improving vitamin D status; however, it seems to be difficult to increase the vitamin D concentrations sufficiently without using either supplements or fortified food in the absence of UV-B radiation.

The authors' responsibilities were as follows—UL and JD: designed the study and applied for funding; UL, HRG, and JD: performed the literature search and the meta-analysis; FH, AM-B, OAG, PMU, GM, GIS, and JD: were involved in the planning, design, and conduct of the 2 unpublished studies; LL, HL, ALH, ATE, and GKP: provided original data from their studies; and all authors: were substantially involved in the writing process. None of the authors stated a conflict of interest.

REFERENCES

- Pilz S, Marz W, Wellnitz B, Seelhorst U, Fahrleitner-Pammer A, Dimai HP, Boehm BO, Dobnig H. Association of vitamin D deficiency with heart failure and sudden cardiac death in a large cross-sectional study of patients referred for coronary angiography. *J Clin Endocrinol Metab* 2008;93:3927–35.
- Giovannucci E, Liu Y, Hollis BW, Rimm EB. 25-Hydroxyvitamin D and risk of myocardial infarction in men: a prospective study. *Arch Intern Med* 2008;168:1174–80.
- Dobnig H, Pilz S, Scharnagl H, Renner W, Seelhorst U, Wellnitz B, Kinkeldei J, Boehm BO, Weihrauch G, Maerz W. Independent association of low serum 25-hydroxyvitamin D levels with all-cause and cardiovascular mortality. *Arch Intern Med* 2008;168:1340–9.
- Forman JP, Giovannucci E, Holmes MD, Bischoff-Ferrari HA, Tworoger SS, Willett WC, Curhan GC. Plasma 25-hydroxyvitamin D levels and risk of incident hypertension. *Hypertension* 2007;49:1063–9.
- Adorini L, Penna G. Control of autoimmune diseases by the vitamin D endocrine system. *Nat Clin Pract Rheumatol* 2008;4:404–12.
- Holick MF. Vitamin D: importance in the prevention of cancers, type 1 diabetes, heart disease and osteoporosis. *Am J Clin Nutr* 2004;79:362–71.
- Chiu KC, Chu A, Go VLW, Saad MF. Hypovitaminosis D is associated with insulin resistance and B cell dysfunction. *Am J Clin Nutr* 2004;79:820–5.
- Holick MF. Vitamin D deficiency. *N Engl J Med* 2007;357:266–81.
- Holick MF. Vitamin D: its role in cancer prevention and treatment. *Prog Biophys Mol Biol* 2006;92:49–59.
- Gorham ED, Garland CF, Garland FC, Grant WB, Mohr SB, Lipkin M, Newmark HL, Giovannucci E, Wei M, Holick MF. Vitamin D and prevention of colorectal cancer. *J Steroid Biochem Mol Biol* 2005;97:179–94.
- Holick MF. Sunlight and vitamin D for bone health and prevention on autoimmune diseases, cancers, and cardiovascular disease. *Am J Clin Nutr* 2004;80(Suppl):1678S–88S.
- Lamberg-Allardt C. Vitamin D in foods and as supplements. *Prog Biophys Mol Biol* 2006;92:33–8.
- Lu Z, Chen TC, Zhang A, Persons KS, Kohn N, Berkowitz R, Martinello S, Holick MF. An evaluation of the vitamin D3 content in fish: is the vitamin D content adequate to satisfy the dietary requirement for vitamin D? *J Steroid Biochem Mol Biol* 2007;103:642–4.
- Max-Rubner-Institut. National Consumption Survey II. 2008 [cited 2014 Jul 10]. Available from: http://www.mri.bund.de/fileadmin/Institute/EV/NVSII_Abschlussbericht_Teil_2.pdf.

15. González-Rodríguez LG, Estaire P, Penaz-Ruiz C, Ortega RM; UCM Research Group VALORNUT. Vitamin D intake and dietary sources in a representative sample of Spanish adults. *J Hum Nutr Diet* 2013;26:64–72.
16. Zgaga L, Theodoratou E, Farrington SM, Agakov F, Tenesa A, Walker M, Knox S, Wallace AM, Cetnarskyj R, McNeill G, et al. Diet, environmental factors, and lifestyle underlie the high prevalence of vitamin D deficiency in healthy adults in Scotland and supplementation reduces the proportion that are severely deficient. *J Nutr* 2011;141:1535–42.
17. van der Meer IM, Boeke AJ, Lips P, Grootjans-Geerts I, Wuister JD, Deville WL, Wielders JP, Bouter LM, Middelkoop BJ. Fatty fish and supplements are the greatest modifiable contributors to the serum 25-hydroxyvitamin D concentration in a multiethnic population. *Clin Endocrinol (Oxf)* 2008;68:466–72.
18. Nakamura K, Nashimoto M, Okuda Y, Ota T, Yamamoto M. Fish as a major source of vitamin D in the Japanese diet. *Nutrition* 2002;18:415–6.
19. Slater J, Larcombe L, Green C, Slivinski C, Singer M, Denecheze L, Whaley C, Nickerson P, Orr P. Dietary intake of vitamin D in another Canadian Dene First Nation community. *Int J Circumpolar Health* 2013;72:20723.
20. Holden JM, Lemar LE. Assessing vitamin D contents in foods and supplements: challenges and needs. *Am J Clin Nutr* 2008;88(Suppl):551S–3S.
21. US Food and Drug Administration. Direct food substances affirmed as generally recognized as safe. Silver Spring (MD): FDA; 2008.
22. Aro TL, Larmo PS, Bäckman CH, Kallio HP, Tahvonen RL. Fatty acids and fat-soluble vitamins in salted herring (*Clupea harengus*) products. *J Agric Food Chem* 2005;53:1482–8.
23. Mozaffarian D, Rimm EB. Fish intake, contaminants, and human health: evaluating the risks and the benefits. *JAMA* 2006;296:1885–99.
24. He K, Song Y, Davi GL, Liu K, Van Horn L, Dyer AR, Greenland P. Accumulated evidence on fish consumption and coronary heart disease mortality: a meta-analysis of cohort studies. *Circulation* 2004;109:2705–11.
25. Hansen AL, Dahl L, Olson G, Thornton D, Graff IE, Frøyland L, Thayer JF, Pallesen S. Fish consumption, sleep, daily functioning, and heart rate variability. *J Clin Sleep Med* 2014;10:567–75.
26. Erkkilä AT, Schwab US, de Mello VDF, Lappalainen T, Mussalo H, Lehto S, Kemi V, Lamberg-Allardt C, Uusitupa MIJ. Effects of fatty and lean fish intake on blood pressure in subjects with coronary heart disease using multiple medications. *Eur J Nutr* 2008;47:319–28.
27. Hallund J, Overgaard Madsen B, Bügel SH, Jacobsen C, Jakobsen J, Krarup H, Holm J, Nielsen HH, Lauritzen L. The effect of farmed trout on cardiovascular risk markers in healthy men. *Br J Nutr* 2010;104:1528–36.
28. Pot GK, Majsak-Newman G, Geelen A, Harvey LJ, Nagengast FM, Witteman BJM, van de Meeberg BC, Timmer R, Tan A, Wahab PJ, et al. Fish consumption and markers of colorectal cancer risk: a multicenter randomized controlled trial. *Am J Clin Nutr* 2009;90:354–61.
29. Abete I, Parra D, Crujeiras AB, Goyenechea E, Martínez JA. Specific insulin sensitivity and leptin responses to a nutritional treatment of obesity via a combination of energy restriction and fatty fish intake. *J Hum Nutr Diet* 2008;21:591–600.
30. Lindqvist H, Langkilde AM, Undeland I, Radendal T, Sandberg AS. Herring (*Clupea harengus*) supplemented diet influences risk factors for CVD in overweight subjects. *Eur J Clin Nutr* 2007;61:1106–13.
31. Seierstad SL, Seljeflot I, Johansen O, Hansen R, Haugen M, Rosenlund G, Frøyland L, Arnesen H. Dietary intake of differently fed salmon; the influence on markers of human atherosclerosis. *Eur J Clin Invest* 2005;35:52–9.
32. Vázquez C, Botella-Carretero JJ, Corella D, Fiol M, Lage M, Lurbe E, Richart C, Fernández-Real JM, Fuentes F, Ordóñez A, et al. White fish reduces cardiovascular risk factors in patients with metabolic syndrome: the WISH-CARE study, a multicenter randomized clinical trial. *Nutr Metab Cardiovasc Dis* 2014;24:328–35.
33. Zhang J, Wang C, Li L, Man Q, Meng L, Song P, Frøyland L, Du ZY. Dietary inclusion of salmon, herring and pompano as oily fish reduces CVD risk markers in dyslipidaemic middle-aged and elderly Chinese women. *Br J Nutr* 2012;108:1455–65.
34. Hansen AL, Dahl L, Bakke L, Frøyland, Thayer JF. Fish consumption and heart rate variability. *J Psychophysiol* 2010;24:41–7.
35. Scheers N, Lindqvist H, Langkilde AM, Undeland I, Sandberg AS. Vitamin B12 as a potential compliance marker for fish intake. *Eur J Nutr* 2014;53:1327–33.
36. Chiang YL, Haddad E, Rajaram S, Shavlik D, Sabate J. The effect of dietary walnuts compared to fatty fish on eicosanoids, cytokines, soluble endothelial adhesion molecules and lymphocyte subsets: a randomized, controlled crossover trial. *Prostaglandins Leukot Essent Fatty Acids* 2012;87:111–7.
37. Lucey AJ, Paschos GK, Cashman KD, Martínéz JA, Thorsdottir I, Kiely M. Influence of moderate energy restriction and seafood consumption on bone turnover in overweight young adults. *Am J Clin Nutr* 2008;87:1045–52.
38. Brustad M, Sandager T, Wilsgaard T, Aksnes L, Lund E. Change in plasma levels of vitamin D after consumption of cod-liver and fresh cod-liver oil as part of the traditional north Norwegian fish dish “Molje”. *Int J Circumpolar Health* 2003;62:40–53.
39. Cochrane Handbook, chapter 16.1.3.2. Imputing standard deviations for changes from baseline [cited 2014 Sep 13]. Available from: http://handbook.cochrane.org/chapter_16/16_1_3_2_imputing_standard_deviations_for_changes_from_baseline.htm.
40. Cochrane Handbook, chapter 16.5.4. Including multiple groups from one study [cited 2014 Sep 13]. Available from: http://handbook.cochrane.org/chapter_16/16_5_4_how_to_include_multiple_groups_from_one_study.htm.
41. Whitehead A. Meta-analysis of controlled clinical trials. Chichester (United Kingdom): John Wiley & Sons; 2002.
42. Egger M, Davey Smith G, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* 1997;315:629–34.
43. Jadad AR, Moore RA, Carroll D, Jenkinson C, Reynolds DJ, Gavaghan DJ, McQuay HJ. Assessing the quality of reports of randomized clinical trials: is blinding necessary? *Control Clin Trials* 1996;17:1–12.
44. Schulz KF, Altman DG, Moher D. CONSORT 2010 statement: updated guidelines for reporting parallel group randomised trials. *J Pharmacol Pharmacother* 2010;1:100–7.
45. Midttun Ø, Ueland PM. Determination of vitamins A, D and E in a small volume of human plasma by a high-throughput method based on liquid chromatography/tandem mass spectrometry. *Rapid Commun Mass Spectrom* 2011;25:1942–8.
46. Müller-Belecke A, Harbach H, Kaufhold S, Seeburg N, Glomb M. Vitamin D-Gehalte wichtiger Wirtschaftsfischarten in Deutschland—Untersuchungen zum Einfluss der Haltungsumwelt und zu Möglichkeiten der Erhöhung. *Fischer & Teichwirt* 2014;9–11 (in German).
47. Norwegian Scientific Committee for Food Safety. A comprehensive assessment of fish and other seafood in the Norwegian diet. Oslo (Norway): Norwegian Scientific Committee for Food Safety; 2007.
48. Ankarberg A, Bjerselius R, Aune M, Damerud PO, Larsson L, Anderson A, Tysklind M, Berge S, Lundstedt-Enkel K, Karlsson L, et al. Study of dioxin and dioxin-like PCB levels in fatty fish from Sweden 2000–2002. *Organohalogen Compd* 2004;66:2035–9.
49. Pandelova M, Henkelmann B, Roots O, Simm M, Järn L, Benfenati E, Schramm KW. Levels of PCDD/F and dioxin-like PCB in Baltic fish of different age and gender. *Chemosphere* 2008;71:369–78.
50. Chen TC, Chimeh F, Lu Z, Mathieu J, Person KS, Zhang A, Kohn N, Martinello S, Berkowitz R, Holick MF. Factors that influence the cutaneous synthesis and dietary sources of vitamin D. *Arch Biochem Biophys* 2007;460:213–7.
51. Kühn T, Kaaks R, Teucher B, Hirche F, Dierkes J, Weikert C, Katzke V, Boeing H, Stangl GI, Buijsse B. Dietary, lifestyle and genetic determinants of vitamin D status: a cross-sectional analysis from the European Prospective Investigation into Cancer and Nutrition (EPIC)-Germany study. *Eur J Nutr* 2014;53:731–41.
52. Crowe FL, Appleby PN, Allen NE, Key TJ. Diet and risk of diverticular disease in Oxford cohort of European Prospective Investigation into Cancer and Nutrition (EPIC): prospective study of British vegetarians and non-vegetarians. *BMJ* 2011;343:d4131.
53. Burgaz A, Akesson A, Oster A, Michaelsson K, Wolk A. Associations of diet, supplement use, and ultraviolet B radiation exposure with vitamin D status in Swedish women during winter. *Am J Clin Nutr* 2007;86:1399–404.
54. Cashman KD, Hill TR, Lucey AJ, Taylor N, Seamans KM, Muldowney S, Fitzgerald AP, Flynn A, Barnes MS, Horigan G, Bonham MP, Duffy EM, Strain JJ, Wallace JM, Kiely M. Estimations of the dietary requirement for vitamin D in free-living adults >64 years of age. *Am J Clin Nutr* 2009;89:1366–74.

55. National Institute of Nutrition and Seafood Research. Seafood data [cited 2014 Jul 10]. Available from: http://www2.nifes.no/index.php?page_id=164&lang_id=2.
56. Souci-Fachmann-Kraut food database (Germany) [cited 2014 Sep 13].
57. McCance and Widdowson's food database (United Kingdom) [cited 2014 Sep 13]. Available from: <http://tna.europarchive.org/20110116113217/http://www.food.gov.uk/science/dietarysurveys/dietsurveys>.
58. Institute of Medicine. Dietary Reference Intakes for calcium and vitamin D. Washington (DC): The National Academies Press; 2011.
59. German Nutrition Society. New reference values for vitamin D. *Ann Nutr Metab* 2012;60:241–6.
60. Health Council of the Netherlands. Evaluation of dietary reference values for vitamin D. The Hague (Netherlands): Health Council of the Netherlands; 2012. Publication No.: 2012/15E.
61. NORDEN. Nordic nutrition recommendations: NNR5—vitamin D. 5th ed. 2012 [cited 2014 Sep 13]. Available from: <http://www.slv.se/upload/NNR5/Vitamin%20D%20NNR%202012.pdf>.
62. Gallagher JC, Sai A, Templin T II, Smith L. Dose response to vitamin D supplementation in postmenopausal women: a randomized trial. *Ann Intern Med* 2012;156:425.

4. Discussion

It has been described earlier that vitamin D deficiency is a major global and national problem. In Germany, daily intake of vitamin D from food is less than 3 µg/d [Max Rubner-Institut, 2008] which deviates enormously to German recommendations [German Nutrition Society, 2012]. This deficit can be compensated through either endogenous vitamin D synthesis, vitamin D supplementation or vitamin D food fortification.

This thesis aimed to investigate dietary measures for the increase of the vitamin D status by either vitamin D supplementation or food intake.

4.1. Efficacy of vitamin D supplementation to optimize vitamin D status in humans

It has been a debate for many years whether vitamin D₂ and D₃ are bioequivalent with respect to the treatment of deficiency, the increase in 25(OH)D-concentrations, and their effect on bone health. Therefore, it was the aim of this thesis to investigate the efficacy of vitamin D₂ and D₃ on vitamin D status (Study 1).

While vitamin D₂ is effective in the treatment of rickets [Jones et al., 1998], the efficacy of increasing the 25(OH)D-concentrations is still unclear. Thus, a number of studies have investigated the ability to correct vitamin D deficiency and to increase the total 25(OH)D-concentration in human. These studies applied different study designs, e.g. high iv. application routes [Romagnoli et al., 2008], daily [Tjellesen et al., 1986; Trang et al., 1998; Holick et al., 2008b, 2008b, 2008a; Glendenning et al., 2009; Biancuzzo et al., 2010; Binkley et al., 2011; Hammami and Yusuf, 2017], weekly [Heaney et al., 2011; Hammami and Yusuf, 2017] or monthly oral application [Binkley et al., 2011; Hammami and Yusuf, 2017] or high single doses of either vitamin D₂ or D₃ [Armas et al., 2004; Romagnoli et al., 2008; Leventis and Kiely, 2009]. However, there was no clear conclusion on the bioequivalence of both forms, which is also due to the fact that many studies were underpowered. Even a recent meta-analysis [Tripkovic et al., 2012] that included one study with intramuscular vs. oral application [Romagnoli et al., 2008]

and seven studies with oral applications only [Trang et al., 1998; Holick et al., 2008b; Glendenning et al., 2009; Biancuzzo et al., 2010; Binkley et al., 2011; Heaney et al., 2011; Hammami and Yusuf, 2017], did not reveal a definitive conclusion whether vitamin D₃ was superior to vitamin D₂ in increasing the 25(OH)D-concentration when applied daily. The results shown in Study 1 clearly demonstrate the superior effect of daily oral vitamin D₃ applications in increasing the total 25(OH)D concentrations, and even more in increasing the 25(OH)D₃-concentrations. Indeed, vitamin D₂ was associated with a significant decrease in 25(OH)D₃-concentrations. This effect was neglected in those studies that measured total 25(OH)D-concentrations by immunoassays [Trang et al., 1998; Romagnoli et al., 2008; Heaney et al., 2011]. Including Study 1 to the meta-analysis of Tripkovic et al. [Tripkovic et al., 2012], the conclusion would change to a clear superiority of vitamin D₃ compared to D₂ by daily supplementation. (Figure 3)

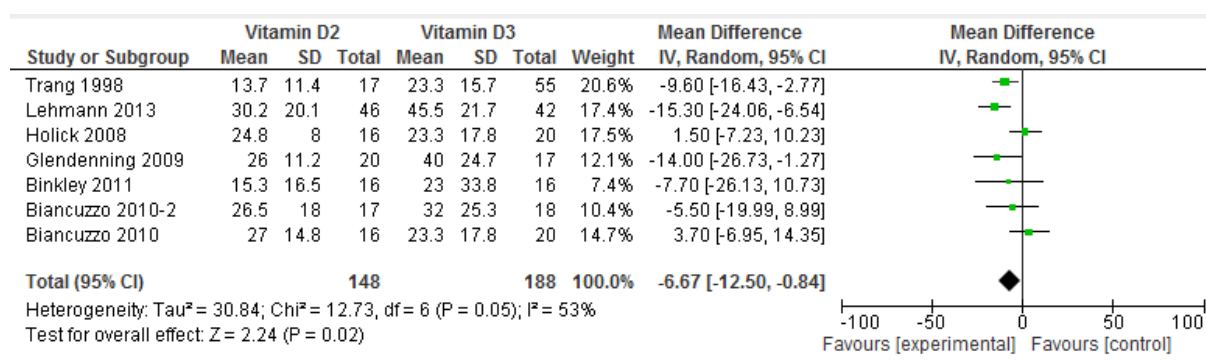


Figure 3: Forest plot on the effect of daily supplementation with either vitamin D₂ or vitamin D₃ on plasma total 25(OH)D concentrations.

experimental = vitamin D₃; control = vitamin D₂. The figure is equivalent to Figure 4 from Tripkovic et al. [2012], only data from the present study are included (data are derived from Table 2B of the article [Lehmann et al., 2013]. Review Manager release 5.2 was used to generate the analysis and the figure.

The different conclusion from the meta-analysis after inclusion of the data of Study 1 might be explained by the increase in the number of subjects included in the meta-analysis. Many prior studies lacked in statistical power and included insufficient sample numbers to draw firm conclusions. This is in contrast to Study 1, that was designed to find a difference in 25(OH)D between vitamin D₃ and D₂ and was thus sufficiently powered.

Currently, it is unclear whether there is also a difference in clinical outcomes between vitamin D₂ and D₃. The group of Bischoff-Ferrari et al. [2009] evaluated in her meta-analysis on vitamin D supplementation and falls supplements containing vitamin D₂

and D₃ separately. While vitamin D₃ supplementation was associated with a 26% decrease of the risk of falls, the effect was only 12% decrease for vitamin D₂ supplementation. There is a lack of studies that compare vitamin D₂ and D₃ for clinical outcomes.

One aspect is of particular interest, and that is the decrease of 25(OH)D₃ upon supplementation with vitamin D₂. It should be noted that only analytical methods using chromatography or mass spectrometry can differentiate between the two compounds, as immunoassays can measure only the total 25(OH)D-concentration. The decrease of 25(OH)D₃ has been also observed by others, [Armas et al., 2004; Glendenning et al., 2009; Binkley et al., 2011; Hammami and Yusuf, 2017]. However, most of these authors did not discuss this specifically. The group of [Holick et al., 2008b] did not observe this effect after 1000 IU vitamin D daily for 11 weeks. Hammami and Yusuf [2017] reported an inverse association of the increase in 25(OH)D₂ with the decrease of 25(OH)D₃ after supplementation with vitamin D₂ with in total 250,000 IU for 140 days ($r = -0.48$). Even if total amounts and also the observed increase in 25(OH)D₂ are comparable to the results of Study 1 (2000 IU/d for 56 days), the association suggested by the authors is in contrast to the association calculated from the present results ($r = -0.07$). In agreement with others [Binkley et al., 2011], a large interindividual variation in the response of both 25(OH)D₃ and 25(OH)D₂ upon vitamin D₂ or D₃ supplementation was observed. This may mask the ability to observe a true association between the changes of either metabolite.

Several mechanisms have been suggested to explain the differences between 25(OH)D-concentrations after similar amounts of vitamin D₃ and D₂, including differences in the affinity for transport proteins or enzymatic activity [Trang et al., 1998; Armas et al., 2004]. However, the present results do not reveal differences in 25-hydroxylation as the increase of 25(OH)D₃ or 25(OH)D₂. Consequently, vitamin D₃ or vitamin D₂ do not differ significantly, suggesting, in agreement with others, [Cheng et al., 2003; Shinkyō et al., 2004; Strushkevich et al., 2008] that the 25-hydroxylation for both vitamin D forms is equal and do not cause differences in bioefficiency.

However, several enzymes are capable to hydroxylize vitamin D₃, among them in the liver microsomal fraction *CYP2R1* and *CYP27A1*. However, the *CYP27A1* cannot 25-hydroxylize vitamin D₂, but hydroxylates vitamin D₂ at C₂₄ [Tuckey et al., 2019]. Thus, the reduction in circulating 25(OH)D₃ following vitamin D₂ supplementation could be due to substrate competition [Glendenning et al., 2009].

Although vitamin D₂ and D₃ only differ in the side chain, they show different metabolic fates. Obviously, both ergocaliferol or 25(OH)D₂, have lower affinity to DBP than cholecalciferol or 25(OH)D₃ [Nilsson et al., 1972; Hollis, 1984; Hollis et al., 1986; Glendenning et al., 2009]. Differences in the side chain of vitamin D₂ and D₃ might cause the different binding capacity [Hollis, 1984] and lead to higher stability of vitamin D₃ binding with consequently higher half-life [Jones et al., 2014] and increased clearance of vitamin D₂ metabolites [Horst et al., 1986]. Further, Horst et al. [1986] demonstrated a 40% higher degradation rate for 1,24,25-(OH)₃D₂ compared to 1,24,25-(OH)₃D₃ in rats.

Study 1 demonstrate that vitamin D₃ is more effective than vitamin D₂ in increasing the total serum 25(OH)D- and 25(OH)D₃-concentration, while vitamin D₂ supplementation is associated with a significant decrease in 25(OH)D₃. Therefore, vitamin D₃ should be recommended as preferred supplement to prevent or treat vitamin D deficiency in humans.

After identifying vitamin D₃ as the most promising supplement (Study 1), it was further aimed to investigate whether 20 µg vitamin D₃ representing the recalculated German recommendations are suitable to optimize vitamin D status in healthy volunteers (Study 2).

Dietary recommendations for vitamin D intake have been revised in many countries during the past 10 years [Ross et al., 2011; German Nutrition Society, 2012; Nordic Council of Ministers, 2012; Scientific Advisory Committee on Nutrition, 2016] following the observations of widespread vitamin D deficiency and low vitamin D intake in many countries [Moore et al., 2005; Hyppönen and Power, 2007; Hintzpeter et al., 2008; Totland et al., 2012; Rabenberg et al., 2015; Pilz et al., 2018]. Numerous vitamin D supplementation studies have been published which have been summarized in several meta-analyses [Cranney et al., 2007; Shab-Bidar et al., 2014; Whiting et al., 2015]. Although these meta-analyses differ in inclusion and exclusion criteria, in the publication periods covered, and in the number of studies, the main findings can be summarized as following: First, all meta-analysis reported significant heterogeneity in the increase of 25(OH)D-concentrations among studies, and this is also true for studies using similar doses of vitamin D. Second, the main determinants of the 25(OH)D-response to vitamin D₃ supplementation are dose, age, baseline 25(OH)D-concentrations and trial duration [Shab-Bidar et al., 2014]. Of these factors, dose,

baseline 25(OH)D-concentrations and trial duration were also identified by Whiting et al. [2015]. Third, both Cranney et al [2007] and Whiting et al. [2015] reported that the response to vitamin D can be described by a regression line (calculated as 2.19 ± 0.97 nmol/l increase in 25(OH)D per μg vitamin D by Whiting et al. [2015], and $0.016 + 19.65$ nmol/l increase in 25(OH)D per IU vitamin D by Cranney et al. [2007]). Interestingly, the regression line by Whiting et al. [2015] is quite similar to the slope of the regression line obtained by Cashman et al. [2009] from several dose response studies in elderly (1.97 nmol/l per μg vitamin D intake).

Applying these calculations to the results of Study 2 would display the average increase in 25(OH)D₃-concentrations in the studied population almost perfectly (observed mean increase after 12 weeks: 35 nmol/l, calculated with Whiting et al. [2015]: 44 nmol/l, calculated with Cranney et al. [2007]: 32.6 nmol/l. It has to be taken into account that the response of 25(OH)D to vitamin D₃ supplementation is most likely not linear at higher doses, but can best be described by a curvilinear function [Shab-Bidar et al., 2014; Whiting et al., 2015].

Both meta-analyses identified study duration as a factor for the achieved 25(OH)D level. Studies with study duration of less than 6 months had a lower increase in 25(OH)D-concentrations than studies of longer duration [Shab-Bidar et al., 2014; Whiting et al., 2015]. At present, it is not clear which trial duration is required to achieve a plateau of 25(OH)D concentrations. Study 2 revealed only a small and non-significant increase in 25(OH)D₃-concentrations between 8 and 12 weeks, however, as the trial finished after 12 weeks, it is difficult to estimate whether the mean 25(OH)D₃-concentrations would have further increased with continued vitamin D supplementation.

Even more interesting than the mean concentration would be the development of 25(OH)D₃-concentrations both at the lower end and the upper end. It was one aim of the Study 2 to increase the 25(OH)D₃-concentration in almost all participants to levels exceeding 50 nmol/l. This aim was already achieved after 8 weeks (when 50 out of 54 participants had 25(OH)D₃-concentrations >50 nmol/l). None of the participants in the vitamin D₃ group had 25(OH)D₃ concentrations <40 nmol/l at this time.

Thus, Study 2 confirmed that 20 µg/d of vitamin D₃ are sufficient to increase the 25(OH)D₃-concentrations in almost all healthy subjects during wintertime. However, when comparing the recommendations, it is obvious that in other countries, lower vitamin D intakes are recommended, despite similar or more northern latitude [Nordic Council of Ministers, 2012]. Taking into account the average 25(OH)D-concentration in Germany (mean 45.6 nmol/l [Rabenberg et al., 2015] or median 45 nmol/l [Hintzpeter et al., 2008] and applying the above mentioned regression lines from the meta-analyses by Whiting et al. [2015] or Cranney et al. [2007], the average 25(OH)D concentration would also exceed 50 nmol/l at additional daily intakes of 4 µg vitamin D. When almost all people should exceed 50 nmol/l, these amounts are not sufficient. Hintzpeter et al. [2008] provided the median and interquartile range of the distribution of 25(OH)D. Choosing the 25th percentile from the NVS 1998 which was 30.5 nmol/l in men and 30.7 nmol/l in women [Hintzpeter et al., 2008] and applying the regression line by Whiting et al. [2015] again, these persons would need about 10 µg additional vitamin D per day to achieve 50 nmol/l. Thus, it can be concluded that a substantial part of the population would need more than 10 µg additional vitamin D to achieve 50 nmol/l 25(OH)D-concentrations. However, 20 µg/d, as recommended by DACH in periods with no endogenous synthesis of vitamin D [German Nutrition Society, 2012], might be too high. Additionally, it has to be considered that the 25(OH)D-concentrations show huge variations among certain groups of the population, e.g. low 25(OH)D-levels are associated with high BMI, during winter and spring, higher age, and low physical activity and low socio-economic status [Rabenberg et al., 2015]. Thus, certain groups clearly need more additional vitamin D than other, and this would call for personalized nutritional advice (and supplementation strategies) instead of general recommendations.

4.2. Efficacy of fish and food consumption to optimize vitamin D status in humans

In addition to vitamin D supplementation, the consumption of vitamin D-rich foods is important to counteract vitamin D deficiency. Since only a few foods contain vitamin D in significant amounts, the enrichment of vitamin D in natural foods is of growing importance. National dietary surveys indicate fish and fish dishes as the most important source of dietary vitamin D in many countries [Max Rubner-Institut, 2008; Amcoff et al., 2012; Totland et al., 2012] and besides fortified food in some countries

(Netherlands, Finland). However, in population-based investigations on determinants of vitamin D status, fish has often a minor role [Kuhn et al., 2014]. Overall low fish consumption and large proportions of non-consumers in certain populations might cause this. Official recommendations of fish intake, usually to consume fish two or three times a week, are met by less than 40% of the population in many countries [Dutch National Institute for Public Health and the Environment; Max Rubner-Institut, 2008; Totland et al., 2012]. These considerations led to the question whether fish intake contributes to the vitamin D status.

Study 3 demonstrated that enrichment of vitamin D content in fish (rainbow trout) is feasible, but is further insufficient to compensate the seasonal drop of 25(OH)D₃-concentrations. In detail, the regular consumption of vitamin D-fortified rainbow trout reduced the decrease of 25(OH)D₃-concentrations that was observed in participants in the conventional fish group during the study period, leading to significantly different 25(OH)D₃-concentrations after 4 weeks. Interestingly, although fish is an important food providing vitamin D, there are no other intervention studies with fish and 25(OH)D₃-concentrations as primary outcome. There are several fish intervention studies with 25(OH)D-concentrations as secondary outcome which are included in the meta-analysis (Study 4) and discussed below.

Thus, the effect of vitamin D-enriched rainbow trout (Study 3) will be compared to low-dose supplement studies in adults, although there only limited number of studies available with supplements that provided 5 µg [Viljakainen et al., 2006; Cashman et al., 2008; Cashman et al., 2009] and none with lower doses. These studies have shown that daily 5 µg vitamin D supplements could either increase 25(OH)D-concentrations in elderly [Viljakainen et al., 2006], maintain 25(OH)D-concentrations in elderly [Cashman et al., 2009], or was associated with a decrease of 25(OH)D-concentrations in adults [Cashman et al., 2008]. Reasons for the different findings may be more related to the higher baseline 25(OH)D-concentrations in the adults than in the elderly [Cashman et al., 2008; Cashman et al., 2009]. Baseline levels in Study 3 resembled closer the baseline levels in the Finnish study, but the amount of vitamin D provided by the fish was less than 5 µg [Viljakainen et al., 2006].

As RCTs are the gold standard to investigate such a hypothesis, we aimed to summarize available studies from the literature. The systematic literature search revealed nine relevant studies (Study 4). It is noteworthy to mention that only Study 3 had the effect of fish consumption on vitamin D status as primary outcome. The other included studies measured vitamin D status as secondary outcome, and reported details of the 25(OH)D-concentration on request (personal communication). The meta-analysis (Study 4) clearly showed that regular fish consumption is associated with an increase in 25(OH)D-concentrations, and fatty fish and study duration were of importance. Indeed, fatty fish like salmon, herring and mackerel have much higher vitamin D contents than lean fish like cod, pollock and freshwater fish like pike and pikeperch. However, comparing the different food databases, large differences in the vitamin D content of fish species among databases become evident (Table 4, Introduction). It is unclear at present whether these differences reflect true differences due to season, habitat or feeding, or whether these differences are due to analytical methods used or other reasons. However, these differences are impeding transnational comparisons on vitamin D intake from fish, and also add insecurity to estimate the amount of fish needed to achieve serum 25(OH)D-concentrations exceeding 50 nmol/l.

Further, it was found that study duration had a strong effect on the results (Study 4). The longest observation period, was 6 months, and the increase of 25(OH)D-concentration was higher in these studies, compared to studies lasting 4-8 weeks. This is particularly interesting as the food based dietary guidelines are intended for life-long use, and are aimed at regular fish consumption as part of regular meals. Thus, it cannot be excluded that the present meta-analysis underestimated the effect of regular fish consumption on 25(OH)D-concentrations in the long term. On the other hand, it is also not known whether vitamin D intake from fish would be high enough to result in a plateau of 25(OH)D-concentrations, as seen in supplement studies, even at low vitamin D dose [Viljakainen et al., 2006]. The uncertainty of vitamin D content of fish, the different vitamin D content of fish species and the dependence of the total 25(OH)D increase on baseline levels makes it difficult to estimate the required amount of fish to increase 25(OH)D-concentrations to specific levels for the general population.

Nevertheless, Study 3 showed that post-mortem radiation of rainbow trout fillet can increase the vitamin D content of this type of fish creating a kind of “functional food”. The efficacy of this principle (UVB-radiation of food to increase vitamin D content) has also been shown in mushrooms [Urbain et al., 2011] thus increasing vitamin D₂ content, and also in living animals like hens which led to increased vitamin D₃ and 25(OH)D₃ content in their eggs [Kühn et al., 2014]. Additional UVB exposure of free-range hens increased vitamin D₃ content by 2.58 µg per egg (3.8 µg in total). During Study 3 content of vitamin D-enriched trout was 2.8 µg/100g fish, less than in eggs containing 3.8 µg per egg following UVB-radiation [Kühn et al., 2014]. It is not known whether upscaling of vitamin D-enriched eggs would lead to same challenges than in fish or could be a more promising strategy with comparable amounts of vitamin D even in higher quantities. Further, the efficacy of vitamin D-enriched eggs on vitamin D-status in humans remains unclear so far, but would lead to doubling of current vitamin D intake in the German population [Max Rubner-Institut, 2008].

For other foods such as UVB-exposed mushrooms a meta-analysis [Cashman and Kiely et al., 2016] showed small increase of 25(OH)D-concentrations in terms of low baseline 25(OH)D-concentrations in humans. The number of RCTs included in this meta-analysis were small (n=6), showed large heterogeneity and inconsistent results. It can be assumed, that vitamin D food fortification via UVB-radiation is a promising strategy. So far, it can be stated that fish is suitable to reduce seasonal 25(OH)D decrease. Further intervention studies on bio fortified foods (e.g. UVB-exposed eggs and mushrooms) targeting vitamin D status are needed to derive recommendations for food fortification.

4.3. Effects of vitamin D supplementation on vitamin D metabolites and cardiovascular risk factors

Further the present results indicated a significant increase of vitamin D-metabolites (24,25(OH)₂D₃) following or a significant difference between placebo and vitamin D₃-group after 12 weeks of vitamin D₃ supplementation (1,25(OH)₂D₃-concentrations). Even other vitamin D supplementation studies showed significant increases of vitamin D-metabolites [Binkley et al., 2017; Saleh et al., 2017; Ketha et al., 2018; Vaes et al., 2018; Martucci et al., 2019]. This was presented for daily [Vaes et al., 2018] and high

single dose [Saleh et al., 2017]. One study comparing single and daily doses showed significant higher 24,25(OH)₂D₃-concentrations for a high single dose, but did not showed differences after 28 days [Ketha et al., 2018].

As a secondary outcome the effect of vitamin D₃ supplementation on cardiovascular risk factors like PTH and FGF-23 (Study 2) was evaluated. There was no significant decrease of PTH concentrations following 12 weeks of vitamin D₃ supplementation.

This is in line with a meta-analysis [Moslehi et al., 2015] which demonstrated high heterogeneity among PTH response according to vitamin D supplementation. This study suggested that PTH response depend on calcium dose, trial duration, baseline PTH-levels, BMI, age and sex. Further they conclude that >75 µg vitamin D per day for at least 12 months a necessary to reach a PTH plateau [Moslehi et al., 2015]. Even another meta-analysis indicated higher dosages and 25(OH)D-concentrations (>86 nmol/l) to decrease PTH concentrations [Mirhosseini et al., 2018].

Studies on the effect on FGF-23 showed different results: While the EVITA-Trial demonstrated an increase of FGF-23 concentrations after 3 years of daily 100µg Vitamin D supplementation [Zittermann et al., 2018], another study did not find a significant change of FGF-23 with a high single oral dose (300.000 IU) [Chitalia et al., 2014]. Even another study using different doses (2.000 IU or 40.000 IU) or time points of vitamin D supplementation (daily or monthly) did not find a significant reduction of FGF-23 concentrations [Mager et al., 2017]. It has to be taken into account that FGF-23 concentrations were mainly investigated in association with patients with cardiovascular diseases [Zittermann et al., 2018] or in patients with chronic kidney disease [Seibert et al., 2013; Mager et al., 2017] and not in healthy adults as we elucidated in Study 2.

4.4. Health and adverse effects of vitamin D supplementation and fish consumption

The meaning of high 25(OH)D-concentrations for health is discussed controversially. While levels around 100 nmol/l have been suggested by meta-analysis to prevent falls, cancer and respiratory infections [Spedding et al., 2013] such levels have recently been associated with increased risk for mortality in patients with coronary heart disease [Degerud et al., 2018] and increased need for mechanical circulatory support

implantation in patients with heart failure [Zittermann, 2017]. However, in Study 2 only one participant had 25(OH)D₃-concentrations exceeding 100 nmol/l and two participants had concentrations exceeding 95 µmol/l both after 8 weeks and after 12 weeks of supplementation. It would have been interesting to study the effect of ongoing supplementation, and more attention to the effect of vitamin D supplementation on high 25(OH)D₃-concentrations should be paid.

Results of Study 4 clearly showed that fish consumption increases concentrations of 25(OH)D, although recommended fish intakes cannot optimize vitamin D status. High intake of fish exceeding recommendation of two to three portions per week might be associated with higher 25(OH)D-concentrations but will also be associated with other health effects. In particular, increased consumption of fatty fish also leads to a high intake of dioxins. Dioxins show negative health effects and promotes cellular growth and differentiation [Döhr et al., 1994] and inhibits estrogen receptor [Tian et al., 1998]. In addition, high intake of dioxins increases insulin resistance and consequently increases diabetic risk [Ruzzin et al., 2010]. The consumption of lean fish might be associated with lower intake of dioxin but is further accompanied with lower intake of vitamin D.

For the environment, increased fish consumption poses the risk of overfishing. Therefore, both, health effects and adverse effects of fish, have to be taken into account while discussing increased fish intake exceeding the recommendations.

4.5. Limitations of vitamin D supplementation and food enrichment

Results showed (Study 2) that the response of 25(OH)D₃-concentrations on supplementation depends on various factors such as baseline vitamin D levels and revealed large heterogeneity among the study participants. This has been described in other studies which additionally identified age [Chen et al., 2008], doses [Heaney et al., 2003] and BMI [Gallagher et al., 2012] as predictors of vitamin D response. People with lower baseline 25(OH)D-concentrations benefit more from vitamin D supplementation than people with higher baseline concentrations [Trang et al., 1998; Aloia et al., 2008; Mazahery et al., 2015].

Results of Study 1 further showed that the decrease of 25(OH)D₃ due to vitamin D₂ supplementation may be of particular importance for the growing part of the population that follows a vegan diet, as this diet does not contain vitamin D₃. Thus, vegans and most vegetarians are dependent on either sun exposure or vitamin D supplements. As vegetarians and especially vegans want to avoid food and nutrients of animal origin, their vitamin D would be mainly vitamin D₂ from either supplements or the few foods (mushrooms, and yeast) that contain vitamin D₂. Although vitamin D₂ improves vitamin D status, it is obviously less effective than vitamin D₃ in comparable amounts (Study 1). Therefore, it is recommended to tailor recommendations according to individual needs and living conditions for optimization of vitamin D status.

The enrichment of foods with vitamin D using UVB-radiation has been identified as a promising strategy for the supply of the population in the future. Indeed, the vitamin D content of fortified fish in Study 3 was unexpectedly low (2.8 µg/100g fillet). Pre-trial measurements had indicated higher vitamin D values in the fillets (18.1 µg/100g). This demonstrates the challenges associated with upscaling food technological processes from the experimental scale to intermediate scale. Irradiation of food using UVB is also a very cost- and time-consuming process that has to be considered in relation to benefits of food enrichment.

Furthermore, vitamin D content in fish showed large variations depending on species [Lu et al., 2007], feeding [Mattila et al., 1997] and season [Mattila et al., 1995] which has to be taken into account while enriching fish with vitamin D.

For the German population fish consumption per capita is low [BLE, 2018], while eggs, which also have been identified as a promising target for UVB-radiation [Kühn et al., 2014], are widely used in German diets. However, if fish will be successfully fortified with vitamin D (*based on preliminary results of Study 3*) and in higher quantities, low amounts of fish would be needed to fulfill vitamin D recommendations.

4.6. Suggestions for future research

During the studies of this thesis, it became apparent that the response to supplementation is mainly dependent on both, the dose of vitamin D and the baseline vitamin D status. Differences in baseline 25(OH)D-concentrations may explain to a large proportion the heterogeneity of the response to similar doses of vitamin D. However, baseline vitamin D status was not included into the regression formula by neither Whiting et al. [2015] or Cashman et al. [2008]. Thus, there is a need for a recalculation of the response to supplemental or dietary vitamin D with consideration of the baseline 25(OH)D-concentrations.

Even though it is well known that there is a large seasonal variation in total 25(OH)D-concentration, it is not clear whether the decrease in 25(OH)D-concentrations during winter season can be predicted by summer or autumn 25(OH)D-concentrations. More longitudinal data are needed, which also take into account potential health effects of high summer and low winter values.

Although, it is now established that vitamin D₂ is associated with a decrease in 25(OH)D₃, this form is further used as a supplement. This applies especially for the growing part of the population who follows a vegan or plant based diet. Long-term effects of vitamin D₂ supplementation in this special group should be investigated.

Work for this thesis also revealed that there are substantial gaps of knowledge in the vitamin D content of foods. Here, international comparisons and application of validated methods both for analysis and sampling should be applied. This would be also a prerequisite for the implementation and monitoring of food fortification strategies, which are more promising than supplementation for the improvement of the vitamin D status of the general population and especially of vulnerable groups.

4.7. Conclusion

The main conclusion derived from the bioavailability study on vitamin D₂ and D₃ is the higher effectiveness of vitamin D₃ in comparison to vitamin D₂ in increasing the 25(OH)D-concentrations in healthy individuals. Thus, vitamin D₃ should be preferred as supplement and for food fortification instead of vitamin D₂. Biochemical, physiological and health effects of long-term vitamin D₂ use, for example in vegans, warrant further investigations.

The second study showed that 12 weeks of supplementation with 20 µg/d vitamin D₃ were efficient to increase the 25(OH)D₃-concentration in wintertime in nearly all participants without increasing the concentrations beyond levels of 100 nmol/l. It became apparent that this dose is safe, but may not be required by all, as seen by the large variation in the achieved concentrations. Thus, a more personalized approach considering individual baseline vitamin D status, dietary habits, health status and lifestyle habits would be warranted to achieve an optimized result in the individual person.

It was shown that the biofortification of fish with vitamin D was successful and efficient, although the achieved vitamin D content was lower than expected. Fish is a major source of dietary vitamin D, and regular fish intake, especially fatty fish can increase the 25(OH)D₃-concentration.

Biofortification seems to be a promising strategy to increase the dietary vitamin D intake and should be promoted more intensely.

This thesis demonstrates the possibility to optimize the vitamin D status by vitamin D supplementation, and also partly by fish intake. Even though recent randomized clinical studies (EVITA, VITAL) did not show improvements in clinical outcomes [Zittermann et al., 2017; Manson et al., 2020], this does not preclude the prevention and treatment of vitamin D deficiency through either dietary fortification or supplementation.

5. Zusammenfassung

Eine unzureichende Vitamin D-Versorgung ist ein weltweites Problem und ist assoziiert mit einem erhöhten Risiko für Bluthochdruck, Diabetes, kardiovaskuläre Erkrankungen und begünstigt weiterhin die Entstehung verschiedener Krebserkrankungen. Es war das Ziel der vorliegenden Arbeit in 3 Studien zu untersuchen inwieweit Vitamin D-Supplemente und der Verzehr von Fisch als wertvolle Vitamin D-Quelle zur Optimierung des Vitamin D-Status bei gesunden Probanden beitragen. Weiterhin sollte der Einfluss von Fischverzehr auf den Vitamin D-Status systematisch untersucht werden.

In Studie 1 wurde die Effekte einer täglichen Dosis von 50 µg Vitamin D₂ und D₃ auf den Vitamin D-Status über einen Zeitraum von 8 Wochen untersucht. Dabei konnte gezeigt werden, dass Vitamin D₃ die 25(OH)D-Konzentration signifikant erhöht und eine Vitamin D₂-Supplementierung zu einer signifikanten Minderung der 25(OH)D₃-Konzentration führt.

In Studie 2 wurde die Effektivität der neuen Vitamin D-Empfehlungen der DGE von täglich 20 µg Vitamin D₃ auf eine Optimierung des Vitamin D-Status nach 12 Wochen untersucht. Die Ergebnisse konnten zeigen, dass in den Vitamin D₃-supplementierten Probanden die 25(OH)D₃-Konzentration signifikant anstieg und 94% der Probanden Konzentrationen > 50 nmol/l erreichten.

Im Rahmen der Studie 3 wurde der Einfluss eines 4-wöchigen Verzehrs von Vitamin D-angereichertem Fisch auf die 25(OH)D-Konzentration untersucht. Die Ergebnisse zeigten, dass nach Verzehr des Vitamin D-angereicherten Fisches die 25(OH)D-Konzentration signifikant weniger sank als durch den Verzehr von konventionellem Fisch. Eine systematische Literatursuche und Meta-Analyse von Fischinterventionsstudien (Studie 4) konnte zeigen, dass der Verzehr von Fisch, insbesondere Fettfisch, zu einer Erhöhung der Vitamin D-Konzentration beiträgt, aber nicht für eine Optimierung des Vitamin D-Status ausreichend ist.

Die Ergebnisse der vorliegenden Arbeit konnten zeigen, dass eine Optimierung des Vitamin D-Status durch Vitamin D-Supplementierung und teilweise durch Fischverzehr möglich ist. Die Anreicherung von Lebensmitteln mit Vitamin D ist dabei eine vielversprechende Strategie zur Verbesserung der Vitamin D-Aufnahme über die Nahrung. Eine personalisierte Betrachtung von Vitamin D-Ausgangswerten, der

Nahrungszufuhr und Lebensstilfaktoren könnte zusätzlich den Effekt von Vitamin D-Supplementierungen beeinflussen.

6. Summary

Inadequate vitamin D supply is a worldwide problem, is associated with increased risk of high blood pressure, diabetes, cardiovascular diseases, and further promotes development of various types of cancer. It was the aim of the present work to investigate the extent to which vitamin D supplements and the consumption of fish as a valuable source of vitamin D contributing to the optimization of vitamin D status in healthy volunteers in three different studies. Furthermore, the influence of fish consumption on vitamin D status was systematically examined.

Study 1 investigated the effects of a daily dose of 50 µg vitamin D₂ and D₃ on vitamin D status over a period of 8 weeks. It was shown that vitamin D₃ significantly increases the 25(OH)D-concentrations and that vitamin D₂ supplementation significantly decreases 25(OH)D₃-concentrations.

Study 2 examined the effectiveness of the new German recommendations for vitamin D intake (20 µg vitamin D₃ daily) to optimize vitamin D status after 12 weeks. Subjects receiving vitamin D₃ showed significantly increases 25(OH)D₃-concentration of which 94% reached concentrations >50 nmol/l.

Study 3 investigated the influence of a 4-week intake of vitamin D-enriched fish on the 25(OH)D-concentration. The results showed that 25(OH)D-concentrations decreased significantly less in the group receiving vitamin D-enriched fish, than in the group consuming conventional fish. A systematic literature search and meta-analysis of fish intervention studies (Study 4) showed that the consumption of fish, particularly fatty fish, contributes to an increase in vitamin D concentration, but is not sufficient to optimize the vitamin D status.

The results of the present work were able to show that an optimization of the vitamin D status is possible through vitamin D supplementation and partly through fish consumption. Enriching foods with vitamin D is a promising strategy for improving vitamin D intake through food. Personalized consideration of baseline vitamin D concentrations, food intake and lifestyle factors could additionally influence vitamin D supplementation

References

- Adams JS, Gacad MA (1985): Characterization of 1 alpha-hydroxylation of vitamin D3 sterols by cultured alveolar macrophages from patients with sarcoidosis. *The Journal of experimental medicine* 161 (4):755–765. DOI: 10.1084/jem.161.4.755.
- Ahonen MH, Tenkanen L, Teppo L, Hakama M, Tuohimaa P (2000): Prostate cancer risk and prediagnostic serum 25-hydroxyvitamin D levels (Finland). *Cancer causes & control* 11 (9):847–852.
- Alemzadeh R, Kichler J, Babar G, Calhoun M (2008): Hypovitaminosis D in obese children and adolescents: relationship with adiposity, insulin sensitivity, ethnicity, and season. *Metabolism: clinical and experimental* 57 (2):183–191. DOI: 10.1016/j.metabol.2007.08.023.
- Aloia JF, Patel M, Dimaano R, Li-Ng M, Talwar SA, Mikhail M, Pollack S, Yeh JK (2008): Vitamin D intake to attain a desired serum 25-hydroxyvitamin D concentration. *Am J Clin Nutr* 87 (6):1952–1958. DOI: 10.1093/ajcn/87.6.1952.
- Amcoff E, Edberg A, Enghardt Barbieri H, Lindroos AK, Nälsen C, Pearson M (2012): Riksmaten 2010–11, Food and Nutrient Intake Among Adults in Sweden (in Swedish) Report. 2012. Swedish National Food Agency, Uppsala.
- ANSES – French Agency for Food, Environmental and Occupational Health & Safety (2013): Vitamin D presentation, food sources and nutritional needs. <http://www.anses.fr/en/content/vitamin-d>. Accessed 02 January 2020.
- Armas LAG, Hollis BW, Heaney RP (2004): Vitamin D2 is much less effective than vitamin D3 in humans. *The Journal of Clinical Endocrinology & Metabolism* 89 (11):5387–5391.
- Avioli LV, Lee SW, McDonald JE, Lund J, DeLuca HF (1967): Metabolism of vitamin D 3-3 H in human subjects: distribution in blood, bile, feces, and urine. *The Journal of clinical investigation* 46 (6):983–992.

References

- Barger-Lux MJ, Heaney RP, Dowell S, Chen TC, Holick MF (1998): Vitamin D and its major metabolites: serum levels after graded oral dosing in healthy men. *Osteoporosis International* 8 (3):222–230.
- Bates B, Lennox A, Prentice A, Bates CJ, Page P, Nicholson S, Swan G (2014): National Diet and Nutrition Survey: Results from Years 1, 2, 3 and 4 (combined) of the Rolling Programme (2008/2009-2011/2012): A survey carried out on behalf of Public Health England and the Food Standards Agency, Public Health England.
- Biancuzzo RM, Young A, Bibuld D, Cai MH, Winter MR, Klein EK, Ameri A, Reitz R, Salameh W, Chen TC (2010): Fortification of orange juice with vitamin D2 or vitamin D3 is as effective as an oral supplement in maintaining vitamin D status in adults. *The American journal of clinical nutrition* 91 (6):1621–1626.
- Bikle D (2009): Nonclassic actions of vitamin D. *The Journal of Clinical Endocrinology & Metabolism* 94 (1):26–34. DOI: 10.1210/jc.2008-1454.
- Bikle DD, Halloran BP, Riviere JE (1994): Production of 1,25 dihydroxyvitamin D3 by perfused pig skin. *The Journal of investigative dermatology* 102 (5):796–798. DOI: 10.1111/1523-1747.ep12378190.
- Bikle DD, Siiteri PK, Ryzen E, Haddad JG, Gee E (1985): Serum protein binding of 1, 25-dihydroxyvitamin D: a reevaluation by direct measurement of free metabolite levels. *The Journal of Clinical Endocrinology & Metabolism* 61 (5):969–975.
- Binkley N, Borchardt G, Siglinsky E, Krueger D (2017): DOES VITAMIN D METABOLITE MEASUREMENT HELP PREDICT 25(OH)D CHANGE FOLLOWING VITAMIN D SUPPLEMENTATION? *Endocrine practice official journal of the American College of Endocrinology and the American Association of Clinical Endocrinologists* 23 (4):432–441. DOI: 10.4158/EP161517.OR.
- Binkley N, Gemar D, Engelke J, Gangnon R, Ramamurthy R, Krueger D, Drezner MK (2011): Evaluation of ergocalciferol or cholecalciferol dosing, 1,600 IU daily or 50,000 IU monthly in older adults. *The Journal of Clinical Endocrinology & Metabolism* 96 (4):981–988. DOI: 10.1210/jc.2010-0015.

- Bischof MG, Heinze G, Vierhapper H (2006): Vitamin D status and its relation to age and body mass index. *Hormone research* 66 (5):211–215. DOI: 10.1159/000094932.
- Bischoff-Ferrari HA, Dawson-Hughes B, Staehelin HB, Orav JE, Stuck AE, Theiler R, Wong JB, Egli A, Kiel DP, Henschkowski J (2009): Fall prevention with supplemental and active forms of vitamin D: a meta-analysis of randomised controlled trials. *BMJ (Clinical research ed.)* 339:b3692. DOI: 10.1136/bmj.b3692.
- Black LJ, Seamans KM, Cashman KD, Kiely M (2012): An updated systematic review and meta-analysis of the efficacy of vitamin D food fortification. United States, vol 6.
- BLE (2018): Der Markt für Fischereierzeugnisse Der Markt für Fischereierzeugnisse in der Bundesrepublik Deutschland im Jahre 2018.
- Brouwer-Brolsma EM, Bischoff-Ferrari HA, Bouillon R, Feskens EJM, Gallagher CJ, Hypponen E, Llewellyn DJ, Stoecklin E, Dierkes J, Kies AK (2013): Vitamin D: do we get enough? *Osteoporosis International* 24 (5):1567–1577.
- Brumbaugh PF, Haussler MR (1975): Nuclear and cytoplasmic binding components for vitamin D metabolites. *Life sciences* 16 (3):353–362.
- Bundesministerium der Justiz und für Verbraucherschutz - Verordnung über vitaminisierte Lebensmittel; in: *Bundesgesetzblatt Teil III*.
- Burgaz A, Åkesson A, Öster A, Michaëlsson K, Wolk A (2007): Associations of diet, supplement use, and ultraviolet B radiation exposure with vitamin D status in Swedish women during winter. *The American journal of clinical nutrition* 86 (5):1399–1404.
- Burgaz A, Orsini N, Larsson SC, Wolk A (2011): Blood 25-hydroxyvitamin D concentration and hypertension: a meta-analysis. *Journal of hypertension* 29 (4):636–645. DOI: 10.1097/HJH.0b013e32834320f9.
- Burnand B, Sloutskis D, Gianoli F, Cornuz J, Rickenbach M, Paccaud F, Burckhardt P (1992): Serum 25-hydroxyvitamin D: distribution and determinants in the Swiss population. *The American journal of clinical nutrition* 56 (3):537–542.

References

- Calvo MS, Whiting SJ, Barton CN (2005): Vitamin D intake: a global perspective of current status. *The Journal of nutrition* 135 (2):310–316.
- Cardus A, Panizo S, Encinas M, Dolcet X, Gallego C, Aldea M, Fernandez E, Valdivielso JM (2009): 1, 25-dihydroxyvitamin D3 regulates VEGF production through a vitamin D response element in the VEGF promoter. *Atherosclerosis* 204 (1):85–89.
- Carnevale V, Modoni S, Pileri M, Di Giorgio A, Chiodini I, Minisola S, Vieth R, Scillitani A (2001): Longitudinal evaluation of vitamin D status in healthy subjects from southern Italy: seasonal and gender differences. *Osteoporosis International* 12 (12):1026–1030.
- Cashman KD (2012): The role of vitamers and dietary-based metabolites of vitamin D in prevention of vitamin D deficiency. *Food & nutrition research* 56. DOI: 10.3402/fnr.v56i0.5383.
- Cashman KD, Dowling KG, Škrabáková Z, Gonzalez-Gross M, Valtueña J, Henauw S de, Moreno L, Damsgaard CT, Michaelsen KF, Mølgaard C (2016): Vitamin D deficiency in Europe: pandemic? *The American journal of clinical nutrition* 103 (4):1033–1044.
- Cashman KD, Hill TR, Lucey AJ, Taylor N, Seamans KM, Muldowney S, Fitzgerald AP, Flynn A, Barnes MS, Horigan G (2008): Estimation of the dietary requirement for vitamin D in healthy adults. *Am J Clin Nutr* 88 (6):1535–1542.
- Cashman KD, Kiely M, Kinsella M, Durazo-Arvizu RA, Tian L, Zhang Y, Lucey A, Flynn A, Gibney MJ, Vesper HW (2013): Evaluation of Vitamin D Standardization Program protocols for standardizing serum 25-hydroxyvitamin D data: a case study of the program's potential for national nutrition and health surveys. *The American journal of clinical nutrition* 97 (6):1235–1242.
- Cashman KD, Kiely M, Seamans KM, Urbain P (2016): Effect of Ultraviolet Light–Exposed Mushrooms on Vitamin D Status: Liquid Chromatography–Tandem Mass Spectrometry Reanalysis of Biobanked Sera from a Randomized Controlled Trial and a Systematic Review plus Meta-Analysis. *J Nutr* 146 (3):565–575. DOI: 10.3945/jn.115.223784.
- Cashman KD, Seamans KM, Lucey AJ, Stöcklin E, Weber P, Kiely M, Hill TR (2012): Relative effectiveness of oral 25-hydroxyvitamin D3 and vitamin D3 in raising wintertime

References

- serum 25-hydroxyvitamin D in older adults. *The American journal of clinical nutrition* 95 (6):1350–1356.
- Cashman KD, Wallace JM, Horigan G, Hill TR, Barnes MS, Lucey AJ, Bonham MP, Taylor N, Duffy EM, Seamans K, Muldowney S, Fitzgerald AP, Flynn A, Strain JJ, Kiely M (2009): Estimation of the dietary requirement for vitamin D in free-living adults =64 y of age. *The American journal of clinical nutrition* 89 (5):1366–1374. DOI: 10.3945/ajcn.2008.27334.
- Castro M, King TS, Kunselman SJ, Cabana MD, Denlinger L, Holguin F, Kazani SD, Moore WC, Moy J, Sorkness CA, Avila P, Bacharier LB, Bleecker E, Boushey HA, Chmiel J, Fitzpatrick AM, Gentile D, Hundal M, Israel E, Kraft M, Krishnan JA, LaForce C, Lazarus SC, Lemanske R, Lugogo N, Martin RJ, Mauger DT, Naureckas E, Peters SP, Phipatanakul W, Que LG, Sheshadri A, Smith L, Solway J, Sullivan-Vedder L, Sumino K, Wechsler ME, Wenzel S, White SR, Sutherland ER (2014): Effect of vitamin D3 on asthma treatment failures in adults with symptomatic asthma and lower vitamin D levels: the VIDA randomized clinical trial. *Jama* 311 (20):2083–2091. DOI: 10.1001/jama.2014.5052.
- Chapuy M-C, Preziosi P, Maamer M, Arnaud S, Galan P, Hercberg S, Meunier PJ (1997): Prevalence of vitamin D insufficiency in an adult normal population. *Osteoporosis International* 7 (5):439–443.
- Chen JS, Sambrook PN, March L, Cameron ID, Cumming RG, Simpson JM, Seibel MJ (2008): Hypovitaminosis D and parathyroid hormone response in the elderly: effects on bone turnover and mortality. *Clinical endocrinology* 68 (2):290–298.
- Chen TC, Chimeh F, Lu Z, Mathieu J, Person KS, Zhang A, Kohn N, Martinello S, Berkowitz R, Holick MF (2007): Factors that influence the cutaneous synthesis and dietary sources of vitamin D. *Archives of biochemistry and biophysics* 460 (2):213–217.
- Cheng JB, Motola DL, Mangelsdorf DJ, Russell DW (2003): De-orphanization of cytochrome P450 2R1: a microsomal vitamin D 25-hydroxylase. *Journal of Biological Chemistry* 278 (39):38084–38093. DOI: 10.1074/jbc.M307028200.
- Chick H (1976): Study of rickets in Vienna 1919–1922. *Medical history* 20 (1):41–51.

References

- Chick H, Roscoe MH (1926): Influence of diet and sunlight upon the amount of vitamin A and vitamin D in the milk afforded by a cow. *Biochemical Journal* 20 (3):632.
- Chitalia N, Ismail T, Tooth L, Boa F, Hampson G, Goldsmith D, Kaski JC, Banerjee D (2014): Impact of vitamin D supplementation on arterial vasomotion, stiffness and endothelial biomarkers in chronic kidney disease patients. *PloS one* 9 (3):e91363. DOI: 10.1371/journal.pone.0091363.
- Chun RF, Peercy BE, Orwoll ES, Nielson CM, Adams JS, Hewison M (2014): Vitamin D and DBP: the free hormone hypothesis revisited. *The Journal of steroid biochemistry and molecular biology* 144:132–137.
- Compston JE, Horton LW, Thompson RP (1979): Treatment of osteomalacia associated with primary biliary cirrhosis with parenteral vitamin D2 or oral 25-hydroxyvitamin D3. *Gut* 20 (2):133–136.
- Cooke NE, Haddad JG (1989): Vitamin D binding protein (Gc-globulin). *Endocrine reviews* 10 (3):294–307.
- Costa EM, Blau HM, Feldman D (1986): 1,25-dihydroxyvitamin D3 receptors and hormonal responses in cloned human skeletal muscle cells. *Endocrinology* 119 (5):2214–2220. DOI: 10.1210/endo-119-5-2214.
- Cranney A, Horsley T, O'Donnell S, Weiler H, Puil L, Ooi D, Atkinson S, Ward L, Moher D, Hanley D (2007): Effectiveness and safety of vitamin D in relation to bone health. *Evidence report/technology assessment* (158):1.
- Crowle AJ, Ross EJ, May MH (1987): Inhibition by 1,25(OH)₂-vitamin D3 of the multiplication of virulent tubercle bacilli in cultured human macrophages. *Infection and immunity* 55 (12):2945–2950.
- Danish National Food Institute: Danish food composition table. <https://frida.fooddata.dk/>. Accessed 11 November 2020.
- Dawson-Hughes B, Heaney RP, Holick MF, Lips P, Meunier PJ, Vieth R (2005): Estimates of optimal vitamin D status, Springer.

References

- Deeb KK, Trump DL, Johnson CS (2007): Vitamin D signalling pathways in cancer: potential for anticancer therapeutics. *Nature reviews cancer* 7 (9):684–700.
- Degerud E, Hoff R, Nygård O, Strand E, Nilsen DW, Nordrehaug JE, Midttun Ø, Ueland PM, Vogel S de, Dierkes J (2016): Cosinor modelling of seasonal variation in 25-hydroxyvitamin D concentrations in cardiovascular patients in Norway. *European journal of clinical nutrition* 70 (4):517.
- Degerud E, Nygård O, Vogel S de, Hoff R, Svingen GFT, Pedersen ER, Nilsen DWT, Nordrehaug JE, Midttun Ø, Ueland PM (2018): Plasma 25-hydroxyvitamin D and mortality in patients with suspected stable angina pectoris. *The Journal of Clinical Endocrinology & Metabolism* 103 (3):1161–1170.
- Delmez JA, Tindira C, Grooms P, Dusso A, Windus DW, Slatopolsky E (1989): Parathyroid hormone suppression by intravenous 1,25-dihydroxyvitamin D. A role for increased sensitivity to calcium. *J Clin Invest* 83 (4):1349–1355. DOI: 10.1172/JCI114022.
- DeLuca HF (2004): Overview of general physiologic features and functions of vitamin D. *Am J Clin Nutr* 80 (6):1689S-1696S. DOI: 10.1093/ajcn/80.6.1689S.
- DeLuca HF (2014): History of the discovery of vitamin D and its active metabolites. *BoneKey reports* 3.
- Dobnig H, Pilz S, Scharnagl H, Renner W, Seelhorst U, Wellnitz B, Kinkeldei J, Boehm BO, Weihrauch G, Maerz W (2008): Independent association of low serum 25-hydroxyvitamin D and 1, 25-dihydroxyvitamin D levels with all-cause and cardiovascular mortality. *Archives of internal medicine* 168 (12):1340–1349.
- Döhr O, Vogel C, Abel J (1994): Modulation of Growth Factor Expression by 2,3,7,8-Tetrachlorodibenzo-p-dioxin; in: pp 142–148.
- Driscoll RH, Meredith SC, Sitrin M, Rosenberg IH (1982): Vitamin D deficiency and bone disease in patients with Crohn's disease. *Gastroenterology* 83 (6):1252–1258.

References

- Dueland S, Helgerud P, Pedersen JI, Berg T, Drevon CA (1983): Plasma clearance, transfer, and distribution of vitamin D₃ from intestinal lymph. *American Journal of Physiology-Endocrinology And Metabolism* 245 (4):E326-E331.
- Dutch National Institute for Public Health and the Environment: NEVO food composition table. nevo-online.rivm.nl. Accessed 11 November 2020.
- Duval D, Durant S, Homo-Delarche F (1983): Non-genomic effects of steroids Interactions of steroid molecules with membrane structures and functions. *Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes* 737 (3-4):409–442.
- EFSA NDA (2016): Panel (EFSA Panel on Dietetic Products, Nutrition and Allergies Scientific Opinion on Dietary Reference Values for Vitamin D, *EFSA Journal*.
- Elmadfa I (2009): *European nutrition and health report 2009*, Karger Medical and Scientific Publishers, vol 62.
- ETH Zurich and BAG: Swiss Food Composition Database. <https://naehwertdaten.ch/en/>. Accessed 11 November 2020.
- Evans RM (1988): The steroid and thyroid hormone receptor superfamily. *Science (New York, N.Y.)* 240 (4854):889–895.
- Evatt ML, DeLong MR, Khazai N, Rosen A, Triche S, Tangpricha V (2008): Prevalence of vitamin D insufficiency in patients with Parkinson disease and Alzheimer disease. *Archives of neurology* 65 (10):1348–1352.
- FIZ (2019): *Fischfavoriten in Deutschland*.
https://www.fischinfo.de/images/presse/2020/Die_Top_5_2019.jpg.
- Fleet JC (2004): Rapid, membrane-initiated actions of 1,25 dihydroxyvitamin D: what are they and what do they mean? *The Journal of nutrition* 134 (12):3215–3218. DOI: 10.1093/jn/134.12.3215.

References

- Flynn A, Hirvonen T, Mensink GBM, Ocké MC, Serra-Majem L, Stos K, Szponar L, Tetens I, Turrini A, Fletcher R (2009): Intake of selected nutrients from foods, from fortification and from supplements in various European countries. *Food & nutrition research* 53 (1):2038.
- Gallagher JC, Rapuri P, Smith L (2007): Falls are associated with decreased renal function and insufficient calcitriol production by the kidney. *The Journal of steroid biochemistry and molecular biology* 103 (3-5):610–613. DOI: 10.1016/j.jsbmb.2006.12.082.
- Gallagher JC, Sai A, Templin T, Smith L (2012): Dose response to vitamin D supplementation in postmenopausal women: a randomized trial. *Annals of internal medicine* 156 (6):425–437.
- Garland C, Garland F, Shaw E, Comstock G, Helsing K, Gorham E (1989): Serum 25-hydroxyvitamin D and colon cancer: eight-year prospective study. *The Lancet* 334 (8673):1176–1178.
- Gattineni J, Bates C, Twombly K, Dwarakanath V, Robinson ML, Goetz R, Mohammadi M, Baum M (2009): FGF23 decreases renal NaPi-2a and NaPi-2c expression and induces hypophosphatemia in vivo predominantly via FGF receptor 1. *American Journal of Physiology-Renal Physiology* 297 (2):F282-F291.
- Gellert S, Ströhle A, Hahn A (2017): Breastfeeding women are at higher risk of vitamin D deficiency than non-breastfeeding women - insights from the German VitaMinFemin study. *Int Breastfeed J* 12:19. DOI: 10.1186/s13006-017-0105-1.
- German Nutrition Society (2012): New reference values for vitamin D. *Annals of nutrition & metabolism* 60 (4):241.
- Glendenning P, Chew GT, Seymour HM, Gillett MJ, Goldswain PR, Inderjeeth CA, Vasikaran SD, Taranto M, Musk AA, Fraser WD (2009): Serum 25-hydroxyvitamin D levels in vitamin D-insufficient hip fracture patients after supplementation with ergocalciferol and cholecalciferol. *Bone* 45 (5):870–875. DOI: 10.1016/j.bone.2009.07.015.
- Gomez JM, Maravall FJ, Gomez N, Navarro MA, Casamitjana R, Soler J (2004): Relationship between 25-(OH) D3, the IGF-I system, leptin, anthropometric and body composition

References

- variables in a healthy, randomly selected population. *Hormone and Metabolic Research* 36 (01):48–53.
- González-Gross M, Valtuena J, Breidenassel C, Moreno LA, Ferrari M, Kersting M, Henauw S de, Gottrand F, Azzini E, Widhalm K (2012): Vitamin D status among adolescents in Europe: the Healthy Lifestyle in Europe by Nutrition in Adolescence study. *British journal of nutrition* 107 (5):755–764.
- Haddad JG (1995): Plasma vitamin D-binding protein (Gc-globulin): multiple tasks. *The Journal of steroid biochemistry and molecular biology* 53 (1-6):579–582.
- Haddad JG, Matsuoka LY, Hollis BW, Hu YZ, Wortsman J (1993): Human plasma transport of vitamin D after its endogenous synthesis. *The Journal of clinical investigation* 91 (6):2552–2555.
- Hammami MM, Yusuf A (2017): Differential effects of vitamin D2 and D3 supplements on 25-hydroxyvitamin D level are dose, sex, and time dependent: a randomized controlled trial. *BMC endocrine disorders* 17 (1):12. DOI: 10.1186/s12902-017-0163-9.
- Hathcock JN, Shao A, Vieth R, Heaney R (2007): Risk assessment for vitamin D. *The American journal of clinical nutrition* 85 (1):6–18.
- Havinga E (1973): Vitamin D, example and challenge. *Experientia* 29 (10):1181–1193.
- Heaney RP, Davies KM, Chen TC, Holick MF, Barger-Lux MJ (2003): Human serum 25-hydroxycholecalciferol response to extended oral dosing with cholecalciferol. *The American journal of clinical nutrition* 77 (1):204–210.
- Heaney RP, Horst RL, Cullen DM, Armas LAG (2009): Vitamin D3 distribution and status in the body. *Journal of the American College of Nutrition* 28 (3):252–256.
- Heaney RP, Recker RR, Grote J, Horst RL, Armas LAG (2011): Vitamin D(3) is more potent than vitamin D(2) in humans. *The Journal of Clinical Endocrinology & Metabolism* 96 (3):E447-52. DOI: 10.1210/jc.2010-2230.

References

- Henderson L, Gregory J, Swan G (2003): The National Diet and Nutrition Survey: adults aged 19 to 64 years. Vitamin and mineral intake and urinary analytes 3.
- Henry HL (2011): Regulation of vitamin D metabolism. Best practice & research. Clinical endocrinology & metabolism 25 (4):531–541. DOI: 10.1016/j.beem.2011.05.003.
- Higashi T, Shibayama Y, Fuji M, Shimada K (2008): Liquid chromatography–tandem mass spectrometric method for the determination of salivary 25-hydroxyvitamin D 3: a noninvasive tool for the assessment of vitamin D status. Analytical and bioanalytical chemistry 391 (1):229–238.
- Hintzpeter B, Mensink GBM, Thierfelder W, Müller MJ, Scheidt-Nave C (2008): Vitamin D status and health correlates among German adults. European journal of clinical nutrition 62 (9):1079.
- Hohman EE, Martin BR, Lachcik PJ, Gordon DT, Fleet JC, Weaver CM (2011): Bioavailability and Efficacy of Vitamin D₂ from UV-Irradiated Yeast in Growing, Vitamin D-Deficient Rats. Journal of agricultural and food chemistry 59 (6):2341–2346. DOI: 10.1021/jf104679c.
- Holden JM, Lemar LE (2008): Assessing vitamin D contents in foods and supplements: challenges and needs. The American journal of clinical nutrition 88 (2):551S-553S. DOI: 10.1093/ajcn/88.2.551S.
- Holick MF (1996): Vitamin D and bone health. The Journal of nutrition 126 (suppl_4):1159S-1164S.
- Holick MF (2004): Vitamin D: importance in the prevention of cancers, type 1 diabetes, heart disease, and osteoporosis. The American journal of clinical nutrition 79 (3):362–371.
- Holick MF (2006): Resurrection of vitamin D deficiency and rickets. The Journal of clinical investigation 116 (8):2062–2072.
- Holick MF (2007a): Vitamin D deficiency. New England Journal of Medicine 357 (3):266–281.

References

Holick MF (2007b): Vitamin D deficiency. *New England Journal of Medicine* 357 (3):266–281. DOI: 10.1056/NEJMra070553.

Holick MF (2009): Vitamin D status: measurement, interpretation, and clinical application. *Annals of epidemiology* 19 (2):73–78.

Holick MF (2010): The vitamin D deficiency pandemic: a forgotten hormone important for health. *Public health reviews* 32 (1):267.

Holick MF (2011): Vitamin D: a d-lightful solution for health. *Journal of Investigative Medicine* 59 (6):872–880.

Holick MF (2012): Vitamin D: extraskkeletal health. *Rheumatic Disease Clinics* 38 (1):141–160.

Holick MF, Biancuzzo RM, Chen TC, Klein EK, Young A, Bibuld D, Reitz R, Salameh W, Ameri A, Tannenbaum AD (2008a): Vitamin D₂ is as effective as vitamin D₃ in maintaining circulating concentrations of 25-hydroxyvitamin D. *The Journal of Clinical Endocrinology & Metabolism* 93 (3):677–681.

Holick MF, Biancuzzo RM, Chen TC, Klein EK, Young A, Bibuld D, Reitz R, Salameh W, Ameri A, Tannenbaum AD (2008b): Vitamin D₂ is as effective as vitamin D₃ in maintaining circulating concentrations of 25-hydroxyvitamin D. *The Journal of Clinical Endocrinology & Metabolism* 93 (3):677–681. DOI: 10.1210/jc.2007-2308.

Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH, Weaver CM (2011): Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *The Journal of Clinical Endocrinology & Metabolism* 96 (7):1911–1930. DOI: 10.1210/jc.2011-0385.

Holick MF, Chen TC, Lu Z, Sauter E (2007): Vitamin D and skin physiology: AD-lightful story. *Journal of Bone and Mineral Research* 22 (S2):V28-V33.

Holick MF, MacLaughlin JA, Doppelt SH (1981): Regulation of cutaneous previtamin D₃ photosynthesis in man: skin pigment is not an essential regulator. *Science (New York, N.Y.)* 211 (4482):590–593. DOI: 10.1126/science.6256855.

References

- Hollis BW (1984): Comparison of equilibrium and disequilibrium assay conditions for ergocalciferol, cholecalciferol and their major metabolites. *Journal of steroid biochemistry* 21 (1):81–86. DOI: 10.1016/0022-4731(84)90063-3.
- Hollis BW, Pittard III WB, Reinhardt TA (1986): Relationships among vitamin D, 25-hydroxyvitamin D, and vitamin D-binding protein concentrations in the plasma and milk of human subjects. *The Journal of Clinical Endocrinology & Metabolism* 62 (1):41–44.
- Horst RL, Reinhardt TA, Ramberg CF, Koszewski NJ, Napoli JL (1986): 24-Hydroxylation of 1,25-dihydroxyergocalciferol. An unambiguous deactivation process. *Journal of Biological Chemistry* 261 (20):9250–9256.
- Hu MC, Shiizaki K, Kuro-o M, Moe OW (2013): Fibroblast growth factor 23 and Klotho: physiology and pathophysiology of an endocrine network of mineral metabolism. *Annual review of physiology* 75:503–533.
- Huhtakangas JA, Olivera CJ, Bishop JE, Zanella LP, Norman AW (2004): The vitamin D receptor is present in caveolae-enriched plasma membranes and binds 1 α , 25 (OH) 2-vitamin D3 in vivo and in vitro. *Molecular Endocrinology* 18 (11):2660–2671.
- Hyppönen E, Power C (2007): Hypovitaminosis D in British adults at age 45 y: nationwide cohort study of dietary and lifestyle predictors. *The American journal of clinical nutrition* 85 (3):860–868.
- Irish Universities Nutrition Alliance (2011): National Adult Nutrition Survey. <https://irp-cdn.multiscreensite.com/46a7ad27/files/uploaded/The%20National%20Adult%20Nutrition%20Survey%20Summary%20Report%20March%202011.pdf>. Accessed 11 November 2020.
- Jääskeläinen T, Itkonen ST, Lundqvist A, Erkkola M, Koskela T, Lakkala K, Dowling KG, Hull GL, Kröger H, Karppinen J, Kyllönen E, Härkänen T, Cashman KD, Männistö S, Lamberg-Allardt C (2017): The positive impact of general vitamin D food fortification policy on vitamin D status in a representative adult Finnish population: evidence from an 11-y follow-up based on standardized 25-hydroxyvitamin D data. *The American journal of clinical nutrition* 105 (6):1512–1520. DOI: 10.3945/ajcn.116.151415.

- Jäpelt RB, Silvestro D, Smedsgaard J, Jensen PE, Jakobsen J (2013): Quantification of vitamin D₃ and its hydroxylated metabolites in waxy leaf nightshade (*Solanum glaucophyllum* Desf.), tomato (*Solanum lycopersicum* L.) and bell pepper (*Capsicum annuum* L.). *Food chemistry* 138 (2-3):1206–1211.
- Jetter A, Egli A, Dawson-Hughes B, Staehelin HB, Stoecklin E, Goessl R, Henschkowski J, Bischoff-Ferrari HA (2014): Pharmacokinetics of oral vitamin D₃ and calcifediol. *Bone* 59:14–19. DOI: 10.1016/j.bone.2013.10.014.
- John EM, Schwartz GG, Dreon DM, Koo J (1999): Vitamin D and breast cancer risk: the NHANES I epidemiologic follow-up study, 1971–1975 to 1992. *Cancer Epidemiology and Prevention Biomarkers* 8 (5):399–406.
- Jones G (1999): Metabolism and catabolism of vitamin D, its metabolites, and clinically relevant analogs; in: *Vitamin D*, Springer, pp 57–84.
- Jones G, Strugnell SA, DeLuca HF (1998): Current understanding of the molecular actions of vitamin D. *Physiological reviews* 78 (4):1193–1231. DOI: 10.1152/physrev.1998.78.4.1193.
- Jones KS, Assar S, Vanderschueren D, Bouillon R, Prentice A, Schoenmakers I (2014): Predictors of 25(OH)D half-life and plasma 25(OH)D concentration in The Gambia and the UK. *Osteoporosis International* 26 (3):1137–1146. DOI: 10.1007/s00198-014-2905-0.
- Jorde R, Sneve M, Hutchinson M, Emaus N, Figenschau Y, Grimnes G (2010): Tracking of serum 25-hydroxyvitamin D levels during 14 years in a population-based study and during 12 months in an intervention study. *American journal of epidemiology* 171 (8):903–908.
- Jurutka PW, Whitfield GK, Hsieh JC, Thompson PD, Haussler CA, Haussler MR (2001): Molecular nature of the vitamin D receptor and its role in regulation of gene expression. *Reviews in endocrine & metabolic disorders* 2 (2):203–216. DOI: 10.1023/a:1010062929140.
- Keegan R-JH, Lu Z, Bogusz JM, Williams JE, Holick MF (2013): Photobiology of vitamin D in mushrooms and its bioavailability in humans. *Dermato-endocrinology* 5 (1):165–176.

- Ketha H, Thacher TD, Oberhelman SS, Fischer PR, Singh RJ, Kumar R (2018): Comparison of the effect of daily versus bolus dose maternal vitamin D3 supplementation on the 24,25-dihydroxyvitamin D3 to 25-hydroxyvitamin D3 ratio. *Bone* 110:321–325. DOI: 10.1016/j.bone.2018.02.024.
- Khaw K-T, Stewart AW, Waayer D, Lawes CMM, Toop L, Camargo Jr CA, Scragg R (2017): Effect of monthly high-dose vitamin D supplementation on falls and non-vertebral fractures: secondary and post-hoc outcomes from the randomised, double-blind, placebo-controlled ViDA trial. *The Lancet Diabetes & Endocrinology* 5 (6):438–447.
- Kleiner-Bossaller A, DeLuca HF (1974): Formation of 1, 24-24-trihydroxyvitamin D3 from 1, 25-dihydroxyvitamin D3. *Biochimica et Biophysica Acta (BBA)-General Subjects* 338 (2):489–495.
- Kuhn T, Kaaks R, Teucher B, Hirche F, Dierkes J, Weikert C, Katzke V, Boeing H, Stangl GI, Buijsse B (2014): Dietary, lifestyle, and genetic determinants of vitamin D status: a cross-sectional analysis from the European Prospective Investigation into Cancer and Nutrition (EPIC)-Germany study. *European journal of nutrition* 53 (3):731–741. DOI: 10.1007/s00394-013-0577-8.
- Kühn J, Schutkowski A, Kluge H, Hirche F, Stangl GI (2014): Free-range farming: A natural alternative to produce vitamin D-enriched eggs. *Nutrition* 30 (4):481–484. DOI: 10.1016/j.nut.2013.10.002.
- Kurosu H, Ogawa Y, Miyoshi M, Yamamoto M, Nandi A, Rosenblatt KP, Baum MG, Schiavi S, Hu M-C, Moe OW, Kuro-o M (2006): Regulation of fibroblast growth factor-23 signaling by klotho. *J Biol Chem* 281 (10):6120–6123. DOI: 10.1074/jbc.C500457200.
- Lagunova Z, Porojnicu AC, Lindberg F, Hexeberg S, Moan J (2009): The dependency of vitamin D status on body mass index, gender, age and season. *Anticancer research* 29 (9):3713–3720.
- Lamberg-Allardt C, Brustad M, Meyer HE, Steingrimsdottir L (2013): Vitamin D—a systematic literature review for the 5th edition of the Nordic Nutrition Recommendations. *Food & nutrition research* 57 (1):22671.

References

- Lamberg-Allardt C, Kärkkäinen M, Seppänen R, Biström H (1993): Low serum 25-hydroxyvitamin D concentrations and secondary hyperparathyroidism in middle-aged white strict vegetarians. *Am J Clin Nutr* 58 (5):684–689. DOI: 10.1093/ajcn/58.5.684.
- Lamberg-Allardt CJE, Outila TA, Kärkkäinen MUM, Rita HJ, Valsta LM (2001): Vitamin D deficiency and bone health in healthy adults in Finland: could this be a concern in other parts of Europe? *Journal of Bone and Mineral Research* 16 (11):2066–2073.
- Lehmann B, Meurer M (2010): Vitamin D metabolism. *Dermatologic therapy* 23 (1):2–12.
- Lehmann B, Tiebel O, Meurer M (1999): Expression of vitamin D3 25-hydroxylase (CYP27) mRNA after induction by vitamin D3 or UVB radiation in keratinocytes of human skin equivalents--a preliminary study. *Archives of dermatological research* 291 (9):507–510. DOI: 10.1007/s004030050445.
- Lehmann U, Hirche F, Stangl GI, Hinz K, Westphal S, Dierkes J (2013): Bioavailability of vitamin D(2) and D(3) in healthy volunteers, a randomized placebo-controlled trial. *The Journal of Clinical Endocrinology & Metabolism* 98 (11):4339–4345. DOI: 10.1210/jc.2012-4287.
- Leventis P, Kiely PDW (2009): The tolerability and biochemical effects of high-dose bolus vitamin D2 and D3 supplementation in patients with vitamin D insufficiency. *Scandinavian journal of rheumatology* 38 (2):149–153. DOI: 10.1080/03009740802419081.
- Li C, Li Y, Gao L-B, Wang Y-Y, Zhou B, Lv M-L, Lu H-M, Zhang L (2009): Vitamin D receptor gene polymorphisms and the risk of colorectal cancer in a Chinese population. *Digestive diseases and sciences* 54 (3):634–639.
- Linseisen J, Bechthold A, Bischoff-Ferrari HA, Hintzpeter B, Leschik-Bonnet E, Reichrath J, Stehle P, Volkert D, Wolfram G, Zittermann A (2011): *Vitamin D und Prävention ausgewählter chronischer Krankheiten*. Bonn: Deutsche Gesellschaft für Ernährung eV.
- Lips P (2004): Which circulating level of 25-hydroxyvitamin D is appropriate? *The Journal of steroid biochemistry and molecular biology* 89:611–614.

- Lips P, Cashman KD, Lamberg-Allardt C, Bischoff-Ferrari HA, Obermayer-Pietsch B, Bianchi ML, Stepan J, Fuleihan GE-H, Bouillon R (2019): Current vitamin D status in European and Middle East countries and strategies to prevent vitamin D deficiency: a position statement of the European Calcified Tissue Society. *European Journal of Endocrinology* 180 (4):P23-P54.
- Littlejohns TJ, Henley WE, Lang IA, Annweiler C, Beauchet O, Chaves PHM, Fried L, Kestenbaum BR, Kuller LH, Langa KM (2014): Vitamin D and the risk of dementia and Alzheimer disease. *Neurology* 83 (10):920–928.
- Lombardi G, Ziemann E, Banfi G, Corbetta S (2020): Physical Activity-Dependent Regulation of Parathyroid Hormone and Calcium-Phosphorous Metabolism. *Int J Mol Sci* 21 (15):5388. DOI: 10.3390/ijms21155388.
- Lu Z, Chen TC, Zhang A, Persons KS, Kohn N, Berkowitz R, Martinello S, Holick MF (2007): An evaluation of the vitamin D3 content in fish: Is the vitamin D content adequate to satisfy the dietary requirement for vitamin D? *The Journal of steroid biochemistry and molecular biology* 103 (3):642–644. DOI: 10.1016/j.jsbmb.2006.12.010.
- MacDonald PN (1999): Molecular biology of the vitamin D receptor; in: *Vitamin D*, Springer, pp 109–128.
- Mager DR, Jackson ST, Hoffmann MR, Jindal K, Senior PA (2017): Vitamin D3 supplementation, bone health and quality of life in adults with diabetes and chronic kidney disease: Results of an open label randomized clinical trial. *Clinical nutrition (Edinburgh, Scotland)* 36 (3):686–696. DOI: 10.1016/j.clnu.2016.05.012.
- Manson JE, Bassuk SS, Cook NR, Lee I-M, Mora S, Albert CM, Buring JE (2020): Vitamin D, Marine n-3 Fatty Acids, and Primary Prevention of Cardiovascular Disease Current Evidence. *Circulation research* 126 (1):112–128. DOI: 10.1161/CIRCRESAHA.119.314541.
- Manson JE, Cook NR, Lee I-M, Christen W, Bassuk SS, Mora S, Gibson H, Gordon D, Copeland T, D'Agostino D, Friedenberg G, Ridge C, Bubes V, Giovannucci EL, Willett WC, Buring JE (2019): Vitamin D Supplements and Prevention of Cancer and

- Cardiovascular Disease. *The New England journal of medicine* 380 (1):33–44. DOI: 10.1056/NEJMoa1809944.
- Martucci G, McNally D, Parekh D, Zajic P, Tuzzolino F, Arcadipane A, Christopher KB, Dobnig H, Amrein K (2019): Trying to identify who may benefit most from future vitamin D intervention trials: a post hoc analysis from the VITDAL-ICU study excluding the early deaths. *Critical care (London, England)* 23 (1):200. DOI: 10.1186/s13054-019-2472-z.
- Mattila P, Piironen V, Haapala R, Hirvi T, Uusi-Rauva E (1997): Possible Factors Responsible for the High Variation in the Cholecalciferol Contents of Fish. *Journal of agricultural and food chemistry* 45 (10):3891–3896. DOI: 10.1021/jf970243j.
- Mattila PH, Piironen VI, Uusi-Rauva EJ, Koivistoinen PE (1994): Vitamin D contents in edible mushrooms. *Journal of agricultural and food chemistry* 42 (11):2449–2453.
- Mattila PH, Piironen VI, Uusi-Rauva EJ, Koivistoinen PE (1995): Contents of cholecalciferol, ergocalciferol, and their 25-hydroxylated metabolites in milk products and raw meat and liver as determined by HPLC. *Journal of agricultural and food chemistry* 43 (9):2394–2399.
- Mattila PH, Valkonen E, Valaja J (2011): Effect of different vitamin D supplementations in poultry feed on vitamin D content of eggs and chicken meat. *Journal of agricultural and food chemistry* 59 (15):8298–8303.
- Max Rubner-Institut (2008): *Nationale Verzehrsstudie II: Ergebnisbericht Teil 2*.
- Mazahery H, Stonehouse W, Hurst PR von (2015): The effect of monthly 50,000 IU or 100,000 IU vitamin D supplements on vitamin D status in premenopausal Middle Eastern women living in Auckland. *European journal of clinical nutrition* 69 (3):367–372. DOI: 10.1038/ejcn.2014.264.
- McCance RA, Widdowson EM (2014): *McCance and Widdowson's the Composition of Foods*, Royal Society of Chemistry.

References

- McCollum EV, Simmonds N, Becker JE, Shipley PG (1922): Studies on experimental rickets XXI. An experimental demonstration of the existence of a vitamin which promotes calcium deposition. *Journal of Biological Chemistry* 53 (2):293–312.
- McDonnell DP, Mangelsdorf DJ, Pike JW, Haussler MR, O'Malley BW (1987): Molecular cloning of complementary DNA encoding the avian receptor for vitamin D. *Science (New York, N.Y.)* 235 (4793):1214–1217. DOI: 10.1126/science.3029866.
- Minisola S, Cianferotti L, Biondi P, Cipriani C, Fossi C, Franceschelli F, Giusti F, Leoncini G, Pepe J, Bischoff-Ferrari HA, Brandi ML (2017): Correction of vitamin D status by calcidiol: pharmacokinetic profile, safety, and biochemical effects on bone and mineral metabolism of daily and weekly dosage regimens. *Osteoporosis international a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA* 28 (11):3239–3249. DOI: 10.1007/s00198-017-4180-3.
- Mirhosseini N, Rainsbury J, Kimball SM (2018): Vitamin D Supplementation, Serum 25(OH)D Concentrations and Cardiovascular Disease Risk Factors: A Systematic Review and Meta-Analysis. *Frontiers in cardiovascular medicine* 5:87. DOI: 10.3389/fcvm.2018.00087.
- Moore CE, Murphy MM, Holick MF (2005): Vitamin D intakes by children and adults in the United States differ among ethnic groups. *The Journal of nutrition* 135 (10):2478–2485.
- Mosekilde L (2005): Vitamin D and the elderly. *Clinical endocrinology* 62 (3):265–281. DOI: 10.1111/j.1365-2265.2005.02226.x.
- Moslehi N, Shab-Bidar S, Mirmiran P, Hosseinpanah F, Azizi F (2015): Determinants of parathyroid hormone response to vitamin D supplementation: a systematic review and meta-analysis of randomised controlled trials. *England*, vol 9.
- Mousa A, Naderpoor N, Courten MP de, Teede H, Kellow N, Walker K, Scragg R, Courten B de (2017): Vitamin D supplementation has no effect on insulin sensitivity or secretion in vitamin D-deficient, overweight or obese adults: a randomized placebo-controlled trial. *The American journal of clinical nutrition* 105 (6):1372–1381. DOI: 10.3945/ajcn.117.152736.

- Müller-Belecke A., Harbach H., Kaufhold S., Seeburg N., Glomb M (2014): Vitamin D-Gehalte wichtiger Wirtschaftsfischarten in Deutschland— Untersuchungen zum Einfluss der Haltungsumwelt und zu Möglichkeiten der Erhöhung. *Rapid Commun Mass Spectrom.*
- Mulligan ML, Felton SK, Riek AE, Bernal-Mizrachi C (2010): Implications of vitamin D deficiency in pregnancy and lactation. *Am J Obstet Gynecol* 202 (5):429.e1-429.e4299. DOI: 10.1016/j.ajog.2009.09.002.
- Napoli JL, Horst RL (1983): C (24)-and C (23)-oxidation, converging pathways of intestinal 1, 25-dihydroxyvitamin D3 metabolism: identification of 24-keto-1, 23, 25-trihydroxyvitamin D3. *Biochemistry* 22 (25):5848–5853.
- Nilsson SF, Östberg L, Peterson PA (1972): Binding of vitamin D to its human carrier plasma protein. *Biochemical and biophysical research communications* 46 (3):1380–1387.
- Nordic Council of Ministers (2012): *Nordic Nutrition Recommendations 2012.*
- Norman AW (2008): From vitamin D to hormone D: fundamentals of the vitamin D endocrine system essential for good health. *The American journal of clinical nutrition* 88 (2):491S-499S.
- Norman AW, Bishop JE, Bula CM, Olivera CJ, Mizwicki MT, Zanello LP, Ishida H, Okamura WH (2002): Molecular tools for study of genomic and rapid signal transduction responses initiated by 1 alpha,25(OH)(2)-vitamin D(3). *Steroids* 67 (6):457–466. DOI: 10.1016/s0039-128x(01)00167-2.
- Norwegian Food Safety Authority: Norwegian food composition table. www.matvaretabellen.no. Accessed 11 November 2020.
- Nykjaer A, Fyfe JC, Kozyraki R, Leheste J-R, Jacobsen C, Nielsen MS, Verroust PJ, Aminoff M, La Chapelle A de, Moestrup SK (2001): Cubilin dysfunction causes abnormal metabolism of the steroid hormone 25 (OH) vitamin D3. *Proceedings of the National Academy of Sciences* 98 (24):13895–13900.

- O'Mahony L, Stepien M, Gibney MJ, Nugent AP, Brennan L (2011): The potential role of vitamin D enhanced foods in improving vitamin D status. *Nutrients* 3 (12):1023–1041.
- O'Donnell S, Cranney A, Horsley T, Weiler HA, Atkinson SA, Hanley DA, Ooi DS, Ward L, Barrowman N, Fang M, Sampson M, Tsertsvadze A, Yazdi F (2008): Efficacy of food fortification on serum 25-hydroxyvitamin D concentrations: systematic review. United States, vol 6.
- Ohnuma N, Norman AW (1982): Identification of a new C-23 oxidation pathway of metabolism for 1, 25-dihydroxyvitamin D₃ present in intestine and kidney. *Journal of Biological Chemistry* 257 (14):8261–8271.
- Outila TA, KÄRKKÄINEN MUM, SEPPÄNEN RH, Lamberg-Allardt CJE (2000): Dietary intake of vitamin D in premenopausal, healthy vegans was insufficient to maintain concentrations of serum 25-hydroxyvitamin D and intact parathyroid hormone within normal ranges during the winter in Finland. *Journal of the American Dietetic Association* 100 (4):434–441.
- Ovesen L, Brot C, Jakobsen J (2003): Food contents and biological activity of 25-hydroxyvitamin D: a vitamin D metabolite to be reckoned with? *Annals of nutrition & metabolism* 47 (3-4):107–113. DOI: 10.1159/000070031.
- Owen GI, Zelent A (2000): Origins and evolutionary diversification of the nuclear receptor superfamily. *Cellular and Molecular Life Sciences CMLS* 57 (5):809–827.
- Park EA (1940): The therapy of rickets. *Journal of the American Medical Association* 115 (5):370–379.
- Park YK, Barton CN, Calvo MS (2001): Dietary contributions to serum 25 (OH) vitamin D levels [25 (OH) D] differ in black and white adults in the United States: Results from NHANES III. *J. Bone Miner. Res*:S212-S212.
- Perwad F, Zhang MYH, Tenenhouse HS, Portale AA (2007): Fibroblast growth factor 23 impairs phosphorus and vitamin D metabolism in vivo and suppresses 25-hydroxyvitamin D-1 α -hydroxylase expression in vitro. *American journal of physiology. Renal physiology* 293 (5):F1577-83. DOI: 10.1152/ajprenal.00463.2006.

References

- Pike JW, Gooz  LL, Haussler MR (1980): Biochemical evidence for 1, 25-dihydroxyvitamin D receptor macromolecules in parathyroid, pancreatic, pituitary, and placental tissues. *Life sciences* 26 (5):407–414.
- Pilz S, Marz W, Cashman KD, Kiely ME, Whiting SJ, Holick MF, Grant WB, Pludowski P, Hilgsmann M, Trummer C, Schwetz V, Lerchbaum E, Pandis M, Tomaschitz A, Grubler MR, Gaksch M, Verheyen N, Hollis BW, Rejnmark L, Karras SN, Hahn A, Bischoff-Ferrari HA, Reichrath J, Jorde R, Elmadfa I, Vieth R, Scragg R, Calvo MS, van Schoor NM, Bouillon R, Lips P, Itkonen ST, Martineau AR, Lamberg-Allardt C, Zittermann A (2018): Rationale and Plan for Vitamin D Food Fortification: A Review and Guidance Paper. *Frontiers in endocrinology* 9:373. DOI: 10.3389/fendo.2018.00373.
- Pittas AG, Lau J, Hu FB, Dawson-Hughes B (2007): The role of vitamin D and calcium in type 2 diabetes. A systematic review and meta-analysis. *The Journal of Clinical Endocrinology & Metabolism* 92 (6):2017–2029.
- Prentice A, Goldberg GR, Schoenmakers I (2008): Vitamin D across the lifecycle: physiology and biomarkers. *The American journal of clinical nutrition* 88 (2):500S-506S. DOI: 10.1093/ajcn/88.2.500S.
- Provedini DM, Tsoukas CD, Deftos LJ, Manolagas SC (1983): 1, 25-dihydroxyvitamin D₃ receptors in human leukocytes. *Science (New York, N.Y.)* 221 (4616):1181–1183.
- Rabenberg M, Scheidt-Nave C, Busch MA, Rieckmann N, Hintzpeter B, Mensink GBM (2015): Vitamin D status among adults in Germany—results from the German Health Interview and Examination Survey for Adults (DEGS1). *BMC public health* 15 (1):641.
- Reichrath J, Schilli M, Kerber A, Bahmer FA, Czarnetzki BM, Paus R (1994): Hair follicle expression of 1, 25-dihydroxyvitamin D₃ receptors during the murine hair cycle. *British Journal of Dermatology* 131 (4):477–482.
- Rizzoli R, Stoermann C, Ammann P, Bonjour J-P (1994): Hypercalcemia and hyperosteolysis in vitamin D intoxication: effects of clodronate therapy. *Bone* 15 (2):193–198.
- Romagnoli E, Mascia ML, Cipriani C, Fassino V, Mazzei F, D'Erasmo E, Carnevale V, Scillitani A, Minisola S (2008): Short and long-term variations in serum calciotropic

References

- hormones after a single very large dose of ergocalciferol (vitamin D₂) or cholecalciferol (vitamin D₃) in the elderly. *The Journal of Clinical Endocrinology & Metabolism* 93 (8):3015–3020. DOI: 10.1210/jc.2008-0350.
- Ross AC, Taylor CL, Yaktine AL, Del Valle, Heather B., Institute of Medicine (2011): Dietary Reference Intakes for Calcium and Vitamin D. The National Academies Collection: Reports funded by National Institutes of Health. Washington (DC).
- Rossini M, Bongi SM, La Montagna G, Minisola G, Malavolta N, Bernini L, Cacace E, Sinigaglia L, Di Munno O, Adami S (2010): Vitamin D deficiency in rheumatoid arthritis: prevalence, determinants and associations with disease activity and disability. *Arthritis research & therapy* 12 (6):R216.
- Rowling MJ, Kemmis CM, Taffany DA, Welsh J (2006): Megalin-mediated endocytosis of vitamin D binding protein correlates with 25-hydroxycholecalciferol actions in human mammary cells. *The Journal of nutrition* 136 (11):2754–2759.
- Ruzzin J, Petersen R, Meugnier E, Madsen L, Lock E-J, Lillefosse H, Ma T, Pesenti S, Sonne SB, Marstrand TT, Malde MK, Du Z-Y, Chavey C, Fajas L, Lundebye A-K, Brand CL, Vidal H, Kristiansen K, Frøylund L (2010): Persistent organic pollutant exposure leads to insulin resistance syndrome. *Environmental health perspectives* 118 (4):465–471. DOI: 10.1289/ehp.0901321.
- Saleh L, Tang J, Gawinecka J, Boesch L, Fraser WD, Eckardstein A von, Nowak A (2017): Impact of a single oral dose of 100,000 IU vitamin D₃ on profiles of serum 25(OH)D₃ and its metabolites 24,25(OH)₂D₃, 3-epi-25(OH)D₃, and 1,25(OH)₂D₃ in adults with vitamin D insufficiency. *Clinical chemistry and laboratory medicine* 55 (12):1912–1921. DOI: 10.1515/cclm-2016-1129.
- Sanders KM, Stuart AL, Williamson EJ, Simpson JA, Kotowicz MA, Young D, Nicholson GC (2010): Annual high-dose oral vitamin D and falls and fractures in older women: a randomized controlled trial. *Jama* 303 (18):1815–1822.
- Saxholt E, Christensen AT, Møller A, Hartkopp HB, Hess Ygil K, Hels OH (2008): Danish food composition databank, revision 7. Department of Nutrition, National Food Institute, Technical University of Denmark.

- Schmid A, Walther B (2013): Natural vitamin D content in animal products. *Advances in nutrition* (Bethesda, Md.) 4 (4):453–462. DOI: 10.3945/an.113.003780.
- Schutkowski A, Krämer J, Kluge H, Hirche F, Krombholz A, Theumer T, Stangl GI (2013): UVB exposure of farm animals: study on a food-based strategy to bridge the gap between current vitamin D intakes and dietary targets. *PloS one* 8 (7):e69418.
- Scientific Advisory Committee on Nutrition (2016): SACN Vitamin D and Health report. https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/537616/SACN_Vitamin_D_and_Health_report.pdf. Accessed 02 January 2020.
- Seibert E, Heine GH, Ulrich C, Seiler S, Kohler H, Girndt M (2013): Influence of cholecalciferol supplementation in hemodialysis patients on monocyte subsets: a randomized, double-blind, placebo-controlled clinical trial. *Nephron. Clinical practice* 123 (3-4):209–219. DOI: 10.1159/000354717.
- Selby PL, Davies M, Marks JS, Mawer EB (1995): Vitamin D intoxication causes hypercalcaemia by increased bone resorption which responds to pamidronate. *Clinical endocrinology* 43 (5):531–536. DOI: 10.1111/j.1365-2265.1995.tb02916.x.
- Serra-Majem L, Ribas-Barba L, Salvador G, Jover L, Raidó B, Ngo J, Plasencia A (2007): Trends in energy and nutrient intake and risk of inadequate intakes in Catalonia, Spain (1992–2003). *Public Health Nutrition* 10 (11A):1354–1367.
- Shab-Bidar S, Bours S, Geusens PPMM, Kessels AGH, van den Bergh JPW (2014): Serum 25(OH)D response to vitamin D3 supplementation: a meta-regression analysis. *Nutrition* (Burbank, Los Angeles County, Calif.) 30 (9):975–985. DOI: 10.1016/j.nut.2013.12.020.
- Shimada T, Urakawa I, Yamazaki Y, Hasegawa H, Hino R, Yoneya T, Takeuchi Y, Fujita T, Fukumoto S, Yamashita T (2004): FGF-23 transgenic mice demonstrate hypophosphatemic rickets with reduced expression of sodium phosphate cotransporter type IIa. *Biochemical and biophysical research communications* 314 (2):409–414. DOI: 10.1016/j.bbrc.2003.12.102.

References

- Shinkyo R, Sakaki T, Kamakura M, Ohta M, Inouye K (2004): Metabolism of vitamin D by human microsomal CYP2R1. *Biochemical and biophysical research communications* 324 (1):451–457. DOI: 10.1016/j.bbrc.2004.09.073.
- Simpson RU, Thomas GA, Arnold AJ (1985): Identification of 1, 25-dihydroxyvitamin D₃ receptors and activities in muscle. *Journal of Biological Chemistry* 260 (15):8882–8891.
- Smolders J, Schuurman KG, van Strien ME, Melief J, Hendrickx D, Hol EM, van Eden C, Luchetti S, Huitinga I (2013): Expression of vitamin D receptor and metabolizing enzymes in multiple sclerosis-affected brain tissue. *Journal of neuropathology and experimental neurology* 72 (2):91–105. DOI: 10.1097/NEN.0b013e31827f4fcc.
- Souci SW, Fachmann W, Kraut H (eds) (2016): *Die Zusammensetzung der Lebensmittel - Nährwert-Tabellen*, ed 8. Stuttgart, Wissenschaftliche Verlagsgesellschaft Stuttgart.
- Spedding S, Vanlint S, Morris H, Scragg R (2013): Does vitamin D sufficiency equate to a single serum 25-hydroxyvitamin D level or are different levels required for non-skeletal diseases? *Nutrients* 5 (12):5127–5139. DOI: 10.3390/nu5125127.
- Stamp TCB (1975): Factors in human vitamin D nutrition and in the production and cure of classical rickets. *Proceedings of the Nutrition Society* 34 (2):119–130.
- Strushkevich N, Usanov SA, Plotnikov AN, Jones G, Park H-W (2008): Structural analysis of CYP2R1 in complex with vitamin D₃. *Journal of molecular biology* 380 (1):95–106. DOI: 10.1016/j.jmb.2008.03.065.
- Swedish National Food Agency: Swedish food composition table. www.livsmedelsverket.se. Accessed 11 November 2020.
- Tanaka Y, Castillo L, DeLuca HF (1977): The 24-hydroxylation of 1, 25-dihydroxyvitamin D₃. *Journal of Biological Chemistry* 252 (4):1421–1424.
- Tangestani H, Djafarian K, Emamat H, Arabzadegan N, Shab-Bidar S (2019): Efficacy of vitamin D fortified foods on bone mineral density and serum bone biomarkers: A systematic review and meta-analysis of interventional studies. *Critical reviews in food science and nutrition*:1–10. DOI: 10.1080/10408398.2018.1558172.

- Thompson GR, Lewis B, Booth CC (1966): Absorption of vitamin D₃-³H in control subjects and patients with intestinal malabsorption. *The Journal of clinical investigation* 45 (1):94–102.
- Tian Y, Ke S, Thomas T, Meeker RJ, Gallo MA (1998): Transcriptional suppression of estrogen receptor gene expression by 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD). *The Journal of steroid biochemistry and molecular biology* 67 (1):17–24. DOI: 10.1016/S0960-0760(98)00067-3.
- Tjellesen L, Hummer L, Christiansen C, Rødbro P (1986): Serum concentration of vitamin D metabolites during treatment with vitamin D₂ and D₃ in normal premenopausal women. *Bone and mineral* 1 (5):407–413.
- Totland TH, Melnæs BK, Lundberg-Hallén N, Helland-Kigen KM, Lund-Blix NA, Myhre JB, Johansen AMW, Løken EB, Andersen LF (2012): Norkost 3. En landsomfattende kostholdsundersøkelse blant menn og kvinner i Norge i alderen 18 (70):2010–2011.
- Trang HM, Cole DE, Rubin LA, Pierratos A, Siu S, Vieth R (1998): Evidence that vitamin D₃ increases serum 25-hydroxyvitamin D more efficiently than does vitamin D₂. *The American journal of clinical nutrition* 68 (4):854–858. DOI: 10.1093/ajcn/68.4.854.
- Tripkovic L, Lambert H, Hart K, Smith CP, Bucca G, Penson S, Chope G, Hyppönen E, Berry J, Vieth R (2012): Comparison of vitamin D₂ and vitamin D₃ supplementation in raising serum 25-hydroxyvitamin D status: a systematic review and meta-analysis. *The American journal of clinical nutrition* 95 (6):1357–1364.
- Tuckey RC, Cheng CYS, Slominski AT (2019): The serum vitamin D metabolome: What we know and what is still to discover. *The Journal of steroid biochemistry and molecular biology* 186:4–21. DOI: 10.1016/j.jsbmb.2018.09.003.
- Urbain P, Singler F, Ihorst G, Biesalski H-K, Bertz H (2011): Bioavailability of vitamin D₂ from UV-B-irradiated button mushrooms in healthy adults deficient in serum 25-hydroxyvitamin D: a randomized controlled trial. *European journal of clinical nutrition* 65 (8):965–971. DOI: 10.1038/ejcn.2011.53.

- Vaes AMM, Tieland M, Regt MF de, Wittwer J, van Loon LJC, Groot LCPGM de (2018): Dose-response effects of supplementation with calcifediol on serum 25-hydroxyvitamin D status and its metabolites: A randomized controlled trial in older adults. *Clinical nutrition* (Edinburgh, Scotland) 37 (3):808–814. DOI: 10.1016/j.clnu.2017.03.029.
- Vieth R (1990): The mechanisms of vitamin D toxicity. *Bone and mineral* 11 (3):267–272.
- Viljakainen HT, Natri A-M, Kärkkäinen M, Huttunen MM, Palssa A, Jakobsen J, Cashman KD, Mølgaard C, Lamberg-Allardt C (2006): A Positive Dose–Response Effect of Vitamin D Supplementation on Site-Specific Bone Mineral Augmentation in Adolescent Girls: A Double-Blinded Randomized Placebo-Controlled 1-Year Intervention. *Journal of Bone and Mineral Research* 21 (6):836–844.
- Wang J, Lv S, Chen G, Gao C, He J, Zhong H, Xu Y (2015): Meta-analysis of the association between vitamin D and autoimmune thyroid disease. *Nutrients* 7 (4):2485–2498.
- Webb AR, Kline L, Holick MF (1988): Influence of season and latitude on the cutaneous synthesis of vitamin D₃: exposure to winter sunlight in Boston and Edmonton will not promote vitamin D₃ synthesis in human skin. *The Journal of Clinical Endocrinology & Metabolism* 67 (2):373–378.
- Wecksler WR, Okamura WH, Norman AW (1978): Studies on the mode of action of vitamin D—XIV. Quantitative assessment of the structural requirements for the interaction of 1 α , 25-dihydroxyvitamin D₃ with its chick intestinal mucosa receptor system. *Journal of steroid biochemistry* 9 (10):929–937.
- Whiting SJ, Bonjour J-P, Payen FD, Rousseau B (2015): Moderate amounts of vitamin D₃ in supplements are effective in raising serum 25-hydroxyvitamin D from low baseline levels in adults: a systematic review. *Nutrients* 7 (4):2311–2323. DOI: 10.3390/nu7042311.
- WHO & FAO (2004): *Vitamin and mineral requirements in human nutrition*, 2nd ed. Geneva, Rome, World Health Organization; FAO.
- Willnow TE, Nykjaer A (2002): Pathways for kidney-specific uptake of the steroid hormone 25-hydroxyvitamin D₃. *Current opinion in lipidology* 13 (3):255–260. DOI: 10.1097/00041433-200206000-00004.

References

- Wu-Wong JR, Nakane M, Ma J, Ruan X, Kroeger PE (2007): VDR-mediated gene expression patterns in resting human coronary artery smooth muscle cells. *Journal of cellular biochemistry* 100 (6):1395–1405. DOI: 10.1002/jcb.21133.
- Yang L, Ma J, Zhang X, Fan Y, Wang L (2012): Protective role of the vitamin D receptor. *Cellular immunology* 279 (2):160–166.
- Zhang M-X, Pan G-T, Guo J-F, Li B-Y, Qin L-Q, Zhang Z-L (2015): Vitamin D deficiency increases the risk of gestational diabetes mellitus: a meta-analysis of observational studies. *Nutrients* 7 (10):8366–8375.
- Zittermann A (2017): The Biphasic Effect of Vitamin D on the Musculoskeletal and Cardiovascular System. *International journal of endocrinology* 2017:3206240. DOI: 10.1155/2017/3206240.
- Zittermann A, Ernst JB, Prokop S, Fuchs U, Dreier J, Kuhn J, Knabbe C, Birschmann I, Schulz U, Berthold HK, Pilz S, Gouni-Berthold I, Gummert JF, Dittrich M, Börgermann J (2017): Effect of vitamin D on all-cause mortality in heart failure (EVITA): a 3-year randomized clinical trial with 4000 IU vitamin D daily. *European heart journal* 38 (29):2279–2286. DOI: 10.1093/eurheartj/ehx235.
- Zittermann A, Ernst JB, Prokop S, Fuchs U, Dreier J, Kuhn J, Knabbe C, Borgermann J, Berthold HK, Pilz S, Gouni-Berthold I, Gummert JF (2018): Effects of Vitamin D Supplementation on Renin and Aldosterone Concentrations in Patients with Advanced Heart Failure: The EVITA Trial. *International journal of endocrinology* 2018:5015417. DOI: 10.1155/2018/5015417.
- Zittermann A, Koerfer R (2008): Vitamin D in the prevention and treatment of coronary heart disease. *Current Opinion in Clinical Nutrition & Metabolic Care* 11 (6):752–757.

Danksagung

An dieser Stelle möchte ich mich herzlich bei all jenen bedanken, die mich in den Jahren intensiver Arbeit an dieser Dissertation unterstützt haben.

Ein besonderer Dank gebührt meinen beiden Professorinnen in Halle und Bergen für die unermüdliche Motivation, Kraft, Hingabe und Freude für unsere gemeinsamen Projekte. Ich möchte Frau Prof. Gabriele I. Stangl herzlich für Ihre stets „offene Tür“, die unerschöpflichen wissenschaftlichen Anregungen und die Möglichkeit danken mich im Rahmen meiner akademischen Laufbahn weiterzuentwickeln. Ein besonderer Dank gilt Frau Prof. Jutta Dierkes für Ihre wissenschaftliche und emotionale Unterstützung, die stetige Förderung und berufliche Herausforderung und Ihr unendliches Vertrauen in mich und meine Arbeit. Danke, dass Sie beide mir ermöglicht haben an diesem wunderbaren Projekt zu arbeiten.

Ich danke allen Mitarbeitern und ehemaligen Kollegen des Instituts für Agrar- und Ernährungswissenschaften für Ihre Mitarbeit und Unterstützung bei der Durchführung dieser Arbeit.

Ein besonderer Dank gilt allen Co-Autoren für Ihr produktives Feedback und die gemeinsame Freude an Wissenschaft.

Der größte Dank gilt meiner Familie, besonders meinen Eltern und meiner Tante, und meinen Freunden für die liebevolle Unterstützung auch in schwierigen Phasen mein persönliches Ziel nicht aus den Augen zu verlieren.

Ich widme diese Arbeit meinem Großvater.

Ulrike Spielau

Leipzig, November 2020

Curriculum Vitae

Persönliche Daten

Name	Ulrike Spielau
Geburtsdatum und -ort	25.06.1986 in Bad Saarow
Geburtsname	Lehmann
Geschlecht	weiblich

Studium und Schulbildung

10/2005 – 09/2010	Studium der Ernährungswissenschaften Martin-Luther-Universität Halle-Wittenberg Abschluss: Diplom Diplomarbeit zur “Exkretion von Katecholaminen in normal- und übergewichtigen Kindern”
09/1998 – 06/2005	Geschwister-Scholl-Gymnasium, Fürstenwalde (Spree) Abschluss: Abitur

Beruflicher Werdegang

Seit 07/2015	Wissenschaftliche Mitarbeiterin Pädiatrisches Forschungszentrum Universitätsklinikum Leipzig
07/2014 – 06/2015	Studienkoordinatorin (<i>Elternzeitvertretung</i>) Pädiatrisches Forschungszentrum Universitätsklinikum Leipzig
01/2014 – 04/2014	Gastwissenschaftlerin Stipendiatin des Norwegischen Forschungsrates Universität Bergen (Norwegen)
10/2010 – 12/2013	Promotionsstudentin Martin-Luther-Universität Halle-Wittenberg Institut für Agrar- und Ernährungswissenschaften

Publikationen

Rosendahl-Riise, H., Spielau, U., Ranhoff, A. H., Gudbrandsen O. A., & Dierkes, J. (2017). Vitamin D supplementation and its influence on muscle strength and mobility in community-dwelling older persons: a systematic review and meta-analysis. *Hum Nutr Diet*, 30(1):3-15.

Lehmann, U., Riedel, A., Hirche, F., Brandsch, C., Girndt, M., Ulrich, C., Seibert, E., Henning, C., Glomb, M.A., Dierkes, J., & Stangl, G. I. (2016). Vitamin D 3 supplementation: Response and predictors of vitamin D 3 metabolites—A randomized controlled trial. *Clinical Nutrition*, 35(2), 351-358.

Lehmann, U., Gjessing, H. R., Hirche, F., Mueller-Belecke, A., Gudbrandsen, O. A., Ueland, P. M., Middtun, Ø., Mellgren, G., Lauritzen, L., Lindqvist, H., Hansen, A.L., Erkkilä, A. T., Pot, G. K., Stangl, G. I., & Dierkes, J. (2015). Efficacy of fish intake on vitamin D status: a meta-analysis of randomized controlled trials. *The American journal of clinical nutrition*, 102(4), 837-847.

Brandsch, C., Zibolka, J., Frommhagen, M., Lehmann, U., Dierkes, J., Kühne, H., Hirche, F., & Stangl, G. I. (2014). Vitamin D is not linked to folate status and mRNA expression of intestinal proton-coupled folate transporter. *European journal of nutrition*, 53(4), 1115-1122.

Lehmann, U., Hirche, F., Stangl, G. I., Hinz, K., Westphal, S., & Dierkes, J. (2013). Bioavailability of vitamin D2 and D3 in healthy volunteers, a randomized placebo-controlled trial. *The Journal of Clinical Endocrinology & Metabolism*, 98(11), 4339-4345.

Ort, Datum

Unterschrift

Eidesstattliche Erklärung

Hiermit versichere ich an Eides statt, dass ich die vorliegende Arbeit selbstständig angefertigt und diese nicht bereits für eine Promotion oder ähnliche Zwecke an einer anderen Universität eingereicht habe. Weiterhin versichere ich, dass ich die verwendeten wissenschaftlichen Arbeiten und Hilfsmittel genau und vollständig angegeben habe.

Ort, Datum

Unterschrift